

GROMACS

Groningen Machine for Chemical Simulations



Reference Manual

Version 2018.4

GROMACS

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Version 2018.4

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More information can be found on our website: www.gromacs.org.

Preface & Disclaimer

This manual is not complete and has no pretention to be so due to lack of time of the contributors – our first priority is to improve the software. It is worked on continuously, which in some cases might mean the information is not entirely correct.

Comments on form and content are welcome, please send them to one of the mailing lists (see www.gromacs.org), or open an issue at redmine.gromacs.org. Corrections can also be made in the GROMACS git source repository and uploaded to gerrit.gromacs.org.

We release an updated version of the manual whenever we release a new version of the software, so in general it is a good idea to use a manual with the same major and minor release number as your GROMACS installation.

On-line Resources

You can find more documentation and other material at our homepage www.gromacs.org. Among other things there is an on-line reference, several GROMACS mailing lists with archives and contributed topologies/force fields.

Citation information

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Chapter 1

Introduction

1.1 Computational Chemistry and Molecular Modeling

GROMACS is an engine to perform molecular dynamics simulations and energy minimization. These are two of the many techniques that belong to the realm of computational chemistry and molecular modeling. *Computational chemistry* is just a name to indicate the use of computational techniques in chemistry, ranging from quantum mechanics of molecules to dynamics of large complex molecular aggregates. *Molecular modeling* indicates the general process of describing complex chemical systems in terms of a realistic atomic model, with the goal being to understand and predict macroscopic properties based on detailed knowledge on an atomic scale. Often, molecular modeling is used to design new materials, for which the accurate prediction of physical properties of realistic systems is required.

Macroscopic physical properties can be distinguished by (a) *static equilibrium properties*, such as the binding constant of an inhibitor to an enzyme, the average potential energy of a system, or the radial distribution function of a liquid, and (b) *dynamic or non-equilibrium properties*, such as the viscosity of a liquid, diffusion processes in membranes, the dynamics of phase changes, reaction kinetics, or the dynamics of defects in crystals. The choice of technique depends on the question asked and on the feasibility of the method to yield reliable results at the present state of the art. Ideally, the (relativistic) time-dependent Schrödinger equation describes the properties of molecular systems with high accuracy, but anything more complex than the equilibrium state of a few atoms cannot be handled at this *ab initio* level. Thus, approximations are necessary; the higher the complexity of a system and the longer the time span of the processes of interest is, the more severe the required approximations are. At a certain point (reached very much earlier than one would wish), the *ab initio* approach must be augmented or replaced by *empirical* parameterization of the model used. Where simulations based on physical principles of atomic interactions still fail due to the complexity of the system, molecular modeling is based entirely on a similarity analysis of known structural and chemical data. The QSAR methods (Quantitative Structure-Activity Relations) and many homology-based protein structure predictions belong to the latter category.

Macroscopic properties are always ensemble averages over a representative statistical ensemble

(either equilibrium or non-equilibrium) of molecular systems. For molecular modeling, this has two important consequences:

- The knowledge of a single structure, even if it is the structure of the global energy minimum, is not sufficient. It is necessary to generate a representative ensemble at a given temperature, in order to compute macroscopic properties. But this is not enough to compute thermodynamic equilibrium properties that are based on free energies, such as phase equilibria, binding constants, solubilities, relative stability of molecular conformations, etc. The computation of free energies and thermodynamic potentials requires special extensions of molecular simulation techniques.
- While molecular simulations, in principle, provide atomic details of the structures and motions, such details are often not relevant for the macroscopic properties of interest. This opens the way to simplify the description of interactions and average over irrelevant details. The science of statistical mechanics provides the theoretical framework for such simplifications. There is a hierarchy of methods ranging from considering groups of atoms as one unit, describing motion in a reduced number of collective coordinates, averaging over solvent molecules with potentials of mean force combined with stochastic dynamics [9], to *mesoscopic dynamics* describing densities rather than atoms and fluxes as response to thermodynamic gradients rather than velocities or accelerations as response to forces [10].

For the generation of a representative equilibrium ensemble two methods are available: (a) *Monte Carlo simulations* and (b) *Molecular Dynamics simulations*. For the generation of non-equilibrium ensembles and for the analysis of dynamic events, only the second method is appropriate. While Monte Carlo simulations are more simple than MD (they do not require the computation of forces), they do not yield significantly better statistics than MD in a given amount of computer time. Therefore, MD is the more universal technique. If a starting configuration is very far from equilibrium, the forces may be excessively large and the MD simulation may fail. In those cases, a robust *energy minimization* is required. Another reason to perform an energy minimization is the removal of all kinetic energy from the system: if several “snapshots” from dynamic simulations must be compared, energy minimization reduces the thermal noise in the structures and potential energies so that they can be compared better.

1.2 Molecular Dynamics Simulations

MD simulations solve Newton’s equations of motion for a system of N interacting atoms:

$$m_i \frac{\partial^2 \mathbf{r}_i}{\partial t^2} = \mathbf{F}_i, \quad i = 1 \dots N. \quad (1.1)$$

The forces are the negative derivatives of a potential function $V(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_N)$:

$$\mathbf{F}_i = -\frac{\partial V}{\partial \mathbf{r}_i} \quad (1.2)$$

The equations are solved simultaneously in small time steps. The system is followed for some time, taking care that the temperature and pressure remain at the required values, and the coordinates are written to an output file at regular intervals. The coordinates as a function of time

type of bond	type of vibration	wavenumber (cm ⁻¹)
C-H, O-H, N-H	stretch	3000–3500
C=C, C=O	stretch	1700–2000
HOH	bending	1600
C-C	stretch	1400–1600
H ₂ CX	sciss, rock	1000–1500
CCC	bending	800–1000
O-H···O	libration	400– 700
O-H···O	stretch	50– 200

Table 1.1: Typical vibrational frequencies (wavenumbers) in molecules and hydrogen-bonded liquids. Compare $kT/h = 200 \text{ cm}^{-1}$ at 300 K.

represent a *trajectory* of the system. After initial changes, the system will usually reach an *equilibrium state*. By averaging over an equilibrium trajectory, many macroscopic properties can be extracted from the output file.

It is useful at this point to consider the limitations of MD simulations. The user should be aware of those limitations and always perform checks on known experimental properties to assess the accuracy of the simulation. We list the approximations below.

The simulations are classical

Using Newton’s equation of motion automatically implies the use of *classical mechanics* to describe the motion of atoms. This is all right for most atoms at normal temperatures, but there are exceptions. Hydrogen atoms are quite light and the motion of protons is sometimes of essential quantum mechanical character. For example, a proton may *tunnel* through a potential barrier in the course of a transfer over a hydrogen bond. Such processes cannot be properly treated by classical dynamics! Helium liquid at low temperature is another example where classical mechanics breaks down. While helium may not deeply concern us, the high frequency vibrations of covalent bonds should make us worry! The statistical mechanics of a classical harmonic oscillator differs appreciably from that of a real quantum oscillator when the resonance frequency ν approximates or exceeds $k_B T/h$. Now at room temperature the wavenumber $\sigma = 1/\lambda = \nu/c$ at which $h\nu = k_B T$ is approximately 200 cm^{-1} . Thus, all frequencies higher than, say, 100 cm^{-1} may misbehave in classical simulations. This means that practically all bond and bond-angle vibrations are suspect, and even hydrogen-bonded motions as translational or librational H-bond vibrations are beyond the classical limit (see Table 1.1). What can we do?

Well, apart from real quantum-dynamical simulations, we can do one of two things:

(a) If we perform MD simulations using harmonic oscillators for bonds, we should make corrections to the total internal energy $U = E_{kin} + E_{pot}$ and specific heat C_V (and to entropy S and free energy A or G if those are calculated). The corrections to the energy and specific heat of a one-dimensional oscillator with frequency ν are: [11]

$$U^{QM} = U^{cl} + kT \left(\frac{1}{2}x - 1 + \frac{x}{e^x - 1} \right) \quad (1.3)$$

$$C_V^{QM} = C_V^{cl} + k \left(\frac{x^2 e^x}{(e^x - 1)^2} - 1 \right), \quad (1.4)$$

where $x = h\nu/kT$. The classical oscillator absorbs too much energy (kT), while the high-frequency quantum oscillator is in its ground state at the zero-point energy level of $\frac{1}{2}h\nu$.

(b) We can treat the bonds (and bond angles) as *constraints* in the equations of motion. The rationale behind this is that a quantum oscillator in its ground state resembles a constrained bond more closely than a classical oscillator. A good practical reason for this choice is that the algorithm can use larger time steps when the highest frequencies are removed. In practice the time step can be made four times as large when bonds are constrained than when they are oscillators [12]. GROMACS has this option for the bonds and bond angles. The flexibility of the latter is rather essential to allow for the realistic motion and coverage of configurational space [13].

Electrons are in the ground state

In MD we use a *conservative* force field that is a function of the positions of atoms only. This means that the electronic motions are not considered: the electrons are supposed to adjust their dynamics instantly when the atomic positions change (the *Born-Oppenheimer* approximation), and remain in their ground state. This is really all right, almost always. But of course, electron transfer processes and electronically excited states can not be treated. Neither can chemical reactions be treated properly, but there are other reasons to shy away from reactions for the time being.

Force fields are approximate

Force fields provide the forces. They are not really a part of the simulation method and their parameters can be modified by the user as the need arises or knowledge improves. But the form of the forces that can be used in a particular program is subject to limitations. The force field that is incorporated in GROMACS is described in Chapter 4. In the present version the force field is pair-additive (apart from long-range Coulomb forces), it cannot incorporate polarizabilities, and it does not contain fine-tuning of bonded interactions. This urges the inclusion of some limitations in this list below. For the rest it is quite useful and fairly reliable for biologically-relevant macromolecules in aqueous solution!

The force field is pair-additive

This means that all *non-bonded* forces result from the sum of non-bonded pair interactions. Non pair-additive interactions, the most important example of which is interaction through atomic polarizability, are represented by *effective pair potentials*. Only average non pair-additive contributions are incorporated. This also means that the pair interactions are not pure, *i.e.*, they are not valid for isolated pairs or for situations that differ appreciably from the test systems on which the models were parameterized. In fact, the effective pair potentials are not that bad in practice. But the omission of polarizability also means that electrons in atoms do not provide a dielectric constant as they should. For example, real liquid alkanes have a dielectric constant of slightly more than 2, which reduce the long-range electrostatic interaction between (partial) charges. Thus, the simulations will exaggerate the long-range Coulomb terms. Luckily, the next item compensates this effect a bit.

Long-range interactions are cut off

In this version, GROMACS always uses a cut-off radius for the Lennard-Jones interactions

and sometimes for the Coulomb interactions as well. The “minimum-image convention” used by GROMACS requires that only one image of each particle in the periodic boundary conditions is considered for a pair interaction, so the cut-off radius cannot exceed half the box size. That is still pretty big for large systems, and trouble is only expected for systems containing charged particles. But then truly bad things can happen, like accumulation of charges at the cut-off boundary or very wrong energies! For such systems, you should consider using one of the implemented long-range electrostatic algorithms, such as particle-mesh Ewald [14, 15].

Boundary conditions are unnatural

Since system size is small (even 10,000 particles is small), a cluster of particles will have a lot of unwanted boundary with its environment (vacuum). We must avoid this condition if we wish to simulate a bulk system. As such, we use periodic boundary conditions to avoid real phase boundaries. Since liquids are not crystals, something unnatural remains. This item is mentioned last because it is the least of the evils. For large systems, the errors are small, but for small systems with a lot of internal spatial correlation, the periodic boundaries may enhance internal correlation. In that case, beware of, and test, the influence of system size. This is especially important when using lattice sums for long-range electrostatics, since these are known to sometimes introduce extra ordering.

1.3 Energy Minimization and Search Methods

As mentioned in sec. 1.1, in many cases energy minimization is required. GROMACS provides a number of methods for local energy minimization, as detailed in sec. 3.10.

The potential energy function of a (macro)molecular system is a very complex landscape (or *hypersurface*) in a large number of dimensions. It has one deepest point, the *global minimum* and a very large number of *local minima*, where all derivatives of the potential energy function with respect to the coordinates are zero and all second derivatives are non-negative. The matrix of second derivatives, which is called the *Hessian matrix*, has non-negative eigenvalues; only the collective coordinates that correspond to translation and rotation (for an isolated molecule) have zero eigenvalues. In between the local minima there are *saddle points*, where the Hessian matrix has only one negative eigenvalue. These points are the mountain passes through which the system can migrate from one local minimum to another.

Knowledge of all local minima, including the global one, and of all saddle points would enable us to describe the relevant structures and conformations and their free energies, as well as the dynamics of structural transitions. Unfortunately, the dimensionality of the configurational space and the number of local minima is so high that it is impossible to sample the space at a sufficient number of points to obtain a complete survey. In particular, no minimization method exists that guarantees the determination of the global minimum in any practical amount of time. Impractical methods exist, some much faster than others [16]. However, given a starting configuration, it is possible to find the *nearest local minimum*. “Nearest” in this context does not always imply “nearest” in a geometrical sense (*i.e.*, the least sum of square coordinate differences), but means the minimum that can be reached by systematically moving down the steepest local gradient. Finding this nearest local minimum is all that GROMACS can do for you, sorry! If you want to find other

minima and hope to discover the global minimum in the process, the best advice is to experiment with temperature-coupled MD: run your system at a high temperature for a while and then quench it slowly down to the required temperature; do this repeatedly! If something as a melting or glass transition temperature exists, it is wise to stay for some time slightly below that temperature and cool down slowly according to some clever scheme, a process called *simulated annealing*. Since no physical truth is required, you can use your imagination to speed up this process. One trick that often works is to make hydrogen atoms heavier (mass 10 or so): although that will slow down the otherwise very rapid motions of hydrogen atoms, it will hardly influence the slower motions in the system, while enabling you to increase the time step by a factor of 3 or 4. You can also modify the potential energy function during the search procedure, *e.g.* by removing barriers (remove dihedral angle functions or replace repulsive potentials by *soft-core* potentials [17]), but always take care to restore the correct functions slowly. The best search method that allows rather drastic structural changes is to allow excursions into four-dimensional space [18], but this requires some extra programming beyond the standard capabilities of GROMACS.

Three possible energy minimization methods are:

- Those that require only function evaluations. Examples are the simplex method and its variants. A step is made on the basis of the results of previous evaluations. If derivative information is available, such methods are inferior to those that use this information.
- Those that use derivative information. Since the partial derivatives of the potential energy with respect to all coordinates are known in MD programs (these are equal to minus the forces) this class of methods is very suitable as modification of MD programs.
- Those that use second derivative information as well. These methods are superior in their convergence properties near the minimum: a quadratic potential function is minimized in one step! The problem is that for N particles a $3N \times 3N$ matrix must be computed, stored, and inverted. Apart from the extra programming to obtain second derivatives, for most systems of interest this is beyond the available capacity. There are intermediate methods that build up the Hessian matrix on the fly, but they also suffer from excessive storage requirements. So GROMACS will shy away from this class of methods.

The *steepest descent* method, available in GROMACS, is of the second class. It simply takes a step in the direction of the negative gradient (hence in the direction of the force), without any consideration of the history built up in previous steps. The step size is adjusted such that the search is fast, but the motion is always downhill. This is a simple and sturdy, but somewhat stupid, method: its convergence can be quite slow, especially in the vicinity of the local minimum! The faster-converging *conjugate gradient method* (see *e.g.* [19]) uses gradient information from previous steps. In general, steepest descents will bring you close to the nearest local minimum very quickly, while conjugate gradients brings you *very* close to the local minimum, but performs worse far away from the minimum. GROMACS also supports the L-BFGS minimizer, which is mostly comparable to *conjugate gradient method*, but in some cases converges faster.

Chapter 2

Definitions and Units

2.1 Notation

The following conventions for mathematical typesetting are used throughout this document:

Item	Notation	Example
Vector	Bold italic	\mathbf{r}_i
Vector Length	Italic	r_i

We define the *lowercase* subscripts i , j , k and l to denote particles: \mathbf{r}_i is the *position vector* of particle i , and using this notation:

$$\mathbf{r}_{ij} = \mathbf{r}_j - \mathbf{r}_i \quad (2.1)$$

$$r_{ij} = |\mathbf{r}_{ij}| \quad (2.2)$$

The force on particle i is denoted by \mathbf{F}_i and

$$\mathbf{F}_{ij} = \text{force on } i \text{ exerted by } j \quad (2.3)$$

Please note that we changed notation as of version 2.0 to $\mathbf{r}_{ij} = \mathbf{r}_j - \mathbf{r}_i$ since this is the notation commonly used. If you encounter an error, let us know.

2.2 MD units

GROMACS uses a consistent set of units that produce values in the vicinity of unity for most relevant molecular quantities. Let us call them *MD units*. The basic units in this system are nm, ps, K, electron charge (e) and atomic mass unit (u), see Table 2.1. The values used in GROMACS are taken from the CODATA Internationally recommended 2010 values of fundamental physical constants (see <http://nist.gov>).

Consistent with these units are a set of derived units, given in Table 2.2.

The **electric conversion factor** $f = \frac{1}{4\pi\epsilon_0} = 138.935\,458 \text{ kJ mol}^{-1} \text{ nm e}^{-2}$. It relates the mechan-

Quantity	Symbol	Unit
length	r	nm = 10^{-9} m
mass	m	u (unified atomic mass unit) = $1.660\,538\,921 \times 10^{-27}$ kg
time	t	ps = 10^{-12} s
charge	q	e = elementary charge = $1.602\,176\,565(\times 10^{-19})$ C
temperature	T	K

Table 2.1: Basic units used in GROMACS.

Quantity	Symbol	Unit
energy	E, V	kJ mol^{-1}
Force	\mathbf{F}	$\text{kJ mol}^{-1} \text{ nm}^{-1}$
pressure	p	bar
velocity	v	$\text{nm ps}^{-1} = 1000 \text{ m s}^{-1}$
dipole moment	μ	$e \text{ nm}$
electric potential	Φ	$\text{kJ mol}^{-1} e^{-1} = 0.010\,364\,269\,19 \text{ Volt}$
electric field	E	$\text{kJ mol}^{-1} \text{ nm}^{-1} e^{-1} = 1.036\,426\,919 \times 10^7 \text{ V m}^{-1}$

Table 2.2: Derived units. Note that an additional conversion factor of 10^{28} a.m.u (≈ 16.6) is applied to get bar instead of internal MD units in the energy and log files.

ical quantities to the electrical quantities as in

$$V = f \frac{q^2}{r} \quad \text{or} \quad F = f \frac{q^2}{r^2} \quad (2.4)$$

Electric potentials Φ and electric fields \mathbf{E} are intermediate quantities in the calculation of energies and forces. They do not occur inside GROMACS. If they are used in evaluations, there is a choice of equations and related units. We strongly recommend following the usual practice of including the factor f in expressions that evaluate Φ and \mathbf{E} :

$$\Phi(\mathbf{r}) = f \sum_j \frac{q_j}{|\mathbf{r} - \mathbf{r}_j|} \quad (2.5)$$

$$\mathbf{E}(\mathbf{r}) = f \sum_j q_j \frac{(\mathbf{r} - \mathbf{r}_j)}{|\mathbf{r} - \mathbf{r}_j|^3} \quad (2.6)$$

With these definitions, $q\Phi$ is an energy and $q\mathbf{E}$ is a force. The units are those given in Table 2.2: about 10 mV for potential. Thus, the potential of an electronic charge at a distance of 1 nm equals $f \approx 140$ units ≈ 1.4 V. (exact value: 1.439 964 5 V)

Note that these units are mutually consistent; changing any of the units is likely to produce inconsistencies and is therefore *strongly discouraged!* In particular: if Å are used instead of nm, the unit of time changes to 0.1 ps. If kcal mol^{-1} ($= 4.184 \text{ kJ mol}^{-1}$) is used instead of kJ mol^{-1} for energy, the unit of time becomes 0.488882 ps and the unit of temperature changes to 4.184 K. But in both cases all electrical energies go wrong, because they will still be computed in kJ mol^{-1} , expecting nm as the unit of length. Although careful rescaling of charges may still yield consistency, it is clear that such confusions must be rigidly avoided.

Symbol	Name	Value
N_{AV}	Avogadro's number	$6.022\,141\,29 \times 10^{23} \text{ mol}^{-1}$
R	gas constant	$8.314\,462\,1 \times 10^{-3} \text{ kJ mol}^{-1} \text{ K}^{-1}$
k_B	Boltzmann's constant	<i>idem</i>
h	Planck's constant	$0.399\,031\,271 \text{ kJ mol}^{-1} \text{ ps}$
\hbar	Dirac's constant	$0.063\,507\,799\,3 \text{ kJ mol}^{-1} \text{ ps}$
c	velocity of light	$299\,792.458 \text{ nm ps}^{-1}$

Table 2.3: Some Physical Constants

Quantity	Symbol	Relation to SI
Length	r^*	$r \sigma^{-1}$
Mass	m^*	$m M^{-1}$
Time	t^*	$t \sigma^{-1} \sqrt{\epsilon/M}$
Temperature	T^*	$k_B T \epsilon^{-1}$
Energy	E^*	$E \epsilon^{-1}$
Force	F^*	$F \sigma \epsilon^{-1}$
Pressure	P^*	$P \sigma^3 \epsilon^{-1}$
Velocity	v^*	$v \sqrt{M/\epsilon}$
Density	ρ^*	$N \sigma^3 V^{-1}$

Table 2.4: Reduced Lennard-Jones quantities

In terms of the MD units, the usual physical constants take on different values (see Table 2.3). All quantities are per mol rather than per molecule. There is no distinction between Boltzmann's constant k and the gas constant R : their value is $0.008\,314\,462\,1 \text{ kJ mol}^{-1} \text{ K}^{-1}$.

2.3 Reduced units

When simulating Lennard-Jones (LJ) systems, it might be advantageous to use reduced units (*i.e.*, setting $\epsilon_{ii} = \sigma_{ii} = m_i = k_B = 1$ for one type of atoms). This is possible. When specifying the input in reduced units, the output will also be in reduced units. The one exception is the *temperature*, which is expressed in $0.008\,314\,462\,1$ reduced units. This is a consequence of using Boltzmann's constant in the evaluation of temperature in the code. Thus not T , but $k_B T$, is the reduced temperature. A GROMACS temperature $T = 1$ means a reduced temperature of $0.008 \dots$ units; if a reduced temperature of 1 is required, the GROMACS temperature should be $120.272\,36$.

In Table 2.4 quantities are given for LJ potentials:

$$V_{LJ} = 4\epsilon \left[\left(\frac{\sigma}{r} \right)^{12} - \left(\frac{\sigma}{r} \right)^6 \right] \quad (2.7)$$

2.4 Mixed or Double precision

GROMACS can be compiled in either mixed or double precision. Documentation of previous GROMACS versions referred to “single precision”, but the implementation has made selective use of double precision for many years. Using single precision for all variables would lead to a significant reduction in accuracy. Although in “mixed precision” all state vectors, i.e. particle coordinates, velocities and forces, are stored in single precision, critical variables are double precision. A typical example of the latter is the virial, which is a sum over all forces in the system, which have varying signs. In addition, in many parts of the code we managed to avoid double precision for arithmetic, by paying attention to summation order or reorganization of mathematical expressions. The default configuration uses mixed precision, but it is easy to turn on double precision by adding the option `-DGMX_DOUBLE=on` to `cmake`. Double precision will be 20 to 100% slower than mixed precision depending on the architecture you are running on. Double precision will use somewhat more memory and run input, energy and full-precision trajectory files will be almost twice as large.

The energies in mixed precision are accurate up to the last decimal, the last one or two decimals of the forces are non-significant. The virial is less accurate than the forces, since the virial is only one order of magnitude larger than the size of each element in the sum over all atoms (sec. A.1). In most cases this is not really a problem, since the fluctuations in the virial can be two orders of magnitude larger than the average. Using cut-offs for the Coulomb interactions cause large errors in the energies, forces, and virial. Even when using a reaction-field or lattice sum method, the errors are larger than, or comparable to, the errors due to the partial use of single precision. Since MD is chaotic, trajectories with very similar starting conditions will diverge rapidly, the divergence is faster in mixed precision than in double precision.

For most simulations, mixed precision is accurate enough. In some cases double precision is required to get reasonable results:

- normal mode analysis, for the conjugate gradient or l-bfgs minimization and the calculation and diagonalization of the Hessian
- long-term energy conservation, especially for large systems

Chapter 3

Algorithms

3.1 Introduction

In this chapter we first give describe some general concepts used in GROMACS: *periodic boundary conditions* (sec. 3.2) and the *group concept* (sec. 3.3). The MD algorithm is described in sec. 3.4: first a global form of the algorithm is given, which is refined in subsequent subsections. The (simple) EM (Energy Minimization) algorithm is described in sec. 3.10. Some other algorithms for special purpose dynamics are described after this.

A few issues are of general interest. In all cases the *system* must be defined, consisting of molecules. Molecules again consist of particles with defined interaction functions. The detailed description of the *topology* of the molecules and of the *force field* and the calculation of forces is given in chapter 4. In the present chapter we describe other aspects of the algorithm, such as pair list generation, update of velocities and positions, coupling to external temperature and pressure, conservation of constraints. The *analysis* of the data generated by an MD simulation is treated in chapter 8.

3.2 Periodic boundary conditions

The classical way to minimize edge effects in a finite system is to apply *periodic boundary conditions*. The atoms of the system to be simulated are put into a space-filling box, which is surrounded by translated copies of itself (Fig. 3.1). Thus there are no boundaries of the system; the artifact caused by unwanted boundaries in an isolated cluster is now replaced by the artifact of periodic conditions. If the system is crystalline, such boundary conditions are desired (although motions are naturally restricted to periodic motions with wavelengths fitting into the box). If one wishes to simulate non-periodic systems, such as liquids or solutions, the periodicity by itself causes errors. The errors can be evaluated by comparing various system sizes; they are expected to be less severe than the errors resulting from an unnatural boundary with vacuum.

There are several possible shapes for space-filling unit cells. Some, like the *rhombic dodecahedron* and the *truncated octahedron* [20] are closer to being a sphere than a cube is, and are therefore

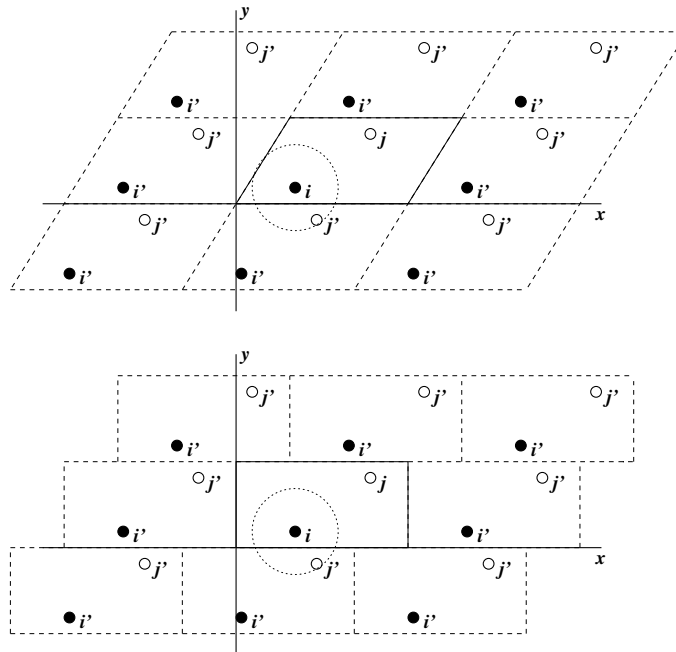


Figure 3.1: Periodic boundary conditions in two dimensions.

better suited to the study of an approximately spherical macromolecule in solution, since fewer solvent molecules are required to fill the box given a minimum distance between macromolecular images. At the same time, rhombic dodecahedra and truncated octahedra are special cases of *triclinic* unit cells; the most general space-filling unit cells that comprise all possible space-filling shapes [21]. For this reason, GROMACS is based on the triclinic unit cell.

GROMACS uses periodic boundary conditions, combined with the *minimum image convention*: only one – the nearest – image of each particle is considered for short-range non-bonded interaction terms. For long-range electrostatic interactions this is not always accurate enough, and GROMACS therefore also incorporates lattice sum methods such as Ewald Sum, PME and PPPM.

GROMACS supports triclinic boxes of any shape. The simulation box (unit cell) is defined by the 3 box vectors \mathbf{a} , \mathbf{b} and \mathbf{c} . The box vectors must satisfy the following conditions:

$$a_y = a_z = b_z = 0 \quad (3.1)$$

$$a_x > 0, \quad b_y > 0, \quad c_z > 0 \quad (3.2)$$

$$|b_x| \leq \frac{1}{2} a_x, \quad |c_x| \leq \frac{1}{2} a_x, \quad |c_y| \leq \frac{1}{2} b_y \quad (3.3)$$

Equations 3.1 can always be satisfied by rotating the box. Inequalities (3.2) and (3.3) can always be satisfied by adding and subtracting box vectors.

Even when simulating using a triclinic box, GROMACS always keeps the particles in a brick-shaped volume for efficiency, as illustrated in Fig. 3.1 for a 2-dimensional system. Therefore, from the output trajectory it might seem that the simulation was done in a rectangular box. The program `trjconv` can be used to convert the trajectory to a different unit-cell representation.

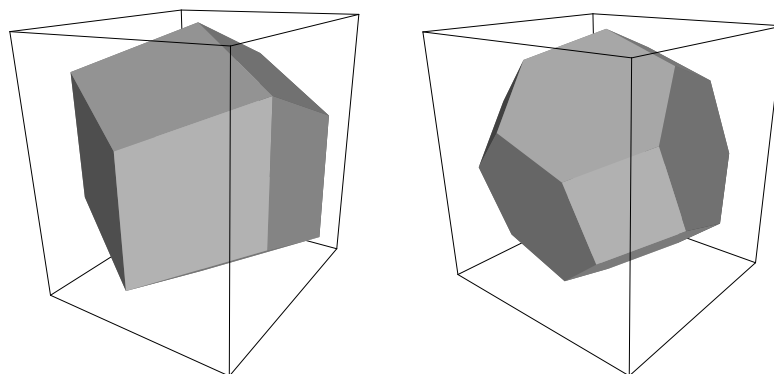


Figure 3.2: A rhombic dodecahedron and truncated octahedron (arbitrary orientations).

box type	image distance	box volume	box vectors			box vector angles		
			a	b	c	$\angle bc$	$\angle ac$	$\angle ab$
cubic	d	d^3	d 0 0	0 d 0	0 0 d	90°	90°	90°
rhombic dodecahedron (xy-square)	d	$\frac{1}{2}\sqrt{2}d^3$ $0.707d^3$	d 0 0	0 d 0	$\frac{1}{2}d$ $\frac{1}{2}d$ $\frac{1}{2}\sqrt{2}d$	60°	60°	90°
rhombic dodecahedron (xy-hexagon)	d	$\frac{1}{2}\sqrt{2}d^3$ $0.707d^3$	d 0 0	$\frac{1}{2}d$ $\frac{1}{2}\sqrt{3}d$ 0	$\frac{1}{2}d$ $\frac{1}{6}\sqrt{3}d$ $\frac{1}{3}\sqrt{6}d$	60°	60°	60°
truncated octahedron	d	$\frac{4}{9}\sqrt{3}d^3$ $0.770d^3$	d 0 0	$\frac{1}{3}d$ $\frac{2}{3}\sqrt{2}d$ 0	$-\frac{1}{3}d$ $\frac{1}{3}\sqrt{2}d$ $\frac{1}{3}\sqrt{6}d$	71.53°	109.47°	71.53°

Table 3.1: The cubic box, the rhombic dodecahedron and the truncated octahedron.

It is also possible to simulate without periodic boundary conditions, but it is usually more efficient to simulate an isolated cluster of molecules in a large periodic box, since fast grid searching can only be used in a periodic system.

3.2.1 Some useful box types

The three most useful box types for simulations of solvated systems are described in Table 3.1. The rhombic dodecahedron (Fig. 3.2) is the smallest and most regular space-filling unit cell. Each of the 12 image cells is at the same distance. The volume is 71% of the volume of a cube having the same image distance. This saves about 29% of CPU-time when simulating a spherical or flexible molecule in solvent. There are two different orientations of a rhombic dodecahedron that satisfy equations 3.1, 3.2 and 3.3. The program `editconf` produces the orientation which has a square intersection with the xy-plane. This orientation was chosen because the first two box vectors coincide with the x and y-axis, which is easier to comprehend. The other orientation can

be useful for simulations of membrane proteins. In this case the cross-section with the xy-plane is a hexagon, which has an area which is 14% smaller than the area of a square with the same image distance. The height of the box (c_z) should be changed to obtain an optimal spacing. This box shape not only saves CPU time, it also results in a more uniform arrangement of the proteins.

3.2.2 Cut-off restrictions

The minimum image convention implies that the cut-off radius used to truncate non-bonded interactions may not exceed half the shortest box vector:

$$R_c < \frac{1}{2} \min(\|\mathbf{a}\|, \|\mathbf{b}\|, \|\mathbf{c}\|), \quad (3.4)$$

because otherwise more than one image would be within the cut-off distance of the force. When a macromolecule, such as a protein, is studied in solution, this restriction alone is not sufficient: in principle, a single solvent molecule should not be able to ‘see’ both sides of the macromolecule. This means that the length of each box vector must exceed the length of the macromolecule in the direction of that edge *plus* two times the cut-off radius R_c . It is, however, common to compromise in this respect, and make the solvent layer somewhat smaller in order to reduce the computational cost. For efficiency reasons the cut-off with triclinic boxes is more restricted. For grid search the extra restriction is weak:

$$R_c < \min(a_x, b_y, c_z) \quad (3.5)$$

For simple search the extra restriction is stronger:

$$R_c < \frac{1}{2} \min(a_x, b_y, c_z) \quad (3.6)$$

Each unit cell (cubic, rectangular or triclinic) is surrounded by 26 translated images. A particular image can therefore always be identified by an index pointing to one of 27 *translation vectors* and constructed by applying a translation with the indexed vector (see 3.4.3). Restriction (3.5) ensures that only 26 images need to be considered.

3.3 The group concept

The GROMACS MD and analysis programs use user-defined *groups* of atoms to perform certain actions on. The maximum number of groups is 256, but each atom can only belong to six different groups, one each of the following:

temperature-coupling group The temperature coupling parameters (reference temperature, time constant, number of degrees of freedom, see 3.4.4) can be defined for each T-coupling group separately. For example, in a solvated macromolecule the solvent (that tends to generate more heating by force and integration errors) can be coupled with a shorter time constant to a bath than is a macromolecule, or a surface can be kept cooler than an adsorbing molecule. Many different T-coupling groups may be defined. See also center of mass groups below.

freeze group Atoms that belong to a freeze group are kept stationary in the dynamics. This is useful during equilibration, *e.g.* to avoid badly placed solvent molecules giving unreasonable kicks to protein atoms, although the same effect can also be obtained by putting a restraining potential on the atoms that must be protected. The freeze option can be used, if desired, on just one or two coordinates of an atom, thereby freezing the atoms in a plane or on a line. When an atom is partially frozen, constraints will still be able to move it, even in a frozen direction. A fully frozen atom can not be moved by constraints. Many freeze groups can be defined. Frozen coordinates are unaffected by pressure scaling; in some cases this can produce unwanted results, particularly when constraints are also used (in this case you will get very large pressures). Accordingly, it is recommended to avoid combining freeze groups with constraints and pressure coupling. For the sake of equilibration it could suffice to start with freezing in a constant volume simulation, and afterward use position restraints in conjunction with constant pressure.

accelerate group On each atom in an “accelerate group” an acceleration a^g is imposed. This is equivalent to an external force. This feature makes it possible to drive the system into a non-equilibrium state and enables the performance of non-equilibrium MD and hence to obtain transport properties.

energy-monitor group Mutual interactions between all energy-monitor groups are compiled during the simulation. This is done separately for Lennard-Jones and Coulomb terms. In principle up to 256 groups could be defined, but that would lead to 256×256 items! Better use this concept sparingly.

All non-bonded interactions between pairs of energy-monitor groups can be excluded (see details in the User Guide). Pairs of particles from excluded pairs of energy-monitor groups are not put into the pair list. This can result in a significant speedup for simulations where interactions within or between parts of the system are not required.

center of mass group In GROMACS the center of mass (COM) motion can be removed, for either the complete system or for groups of atoms. The latter is useful, *e.g.* for systems where there is limited friction (*e.g.* gas systems) to prevent center of mass motion to occur. It makes sense to use the same groups for temperature coupling and center of mass motion removal.

Compressed position output group In order to further reduce the size of the compressed trajectory file (`.xtc` or `.trj`), it is possible to store only a subset of all particles. All x-compression groups that are specified are saved, the rest are not. If no such groups are specified, then all atoms are saved to the compressed trajectory file.

The use of groups in GROMACS tools is described in sec. 8.1.

3.4 Molecular Dynamics

A global flow scheme for MD is given in Fig. 3.3. Each MD or EM run requires as input a set of initial coordinates and – optionally – initial velocities of all particles involved. This chapter does not describe how these are obtained; for the setup of an actual MD run check the online manual at www.gromacs.org.

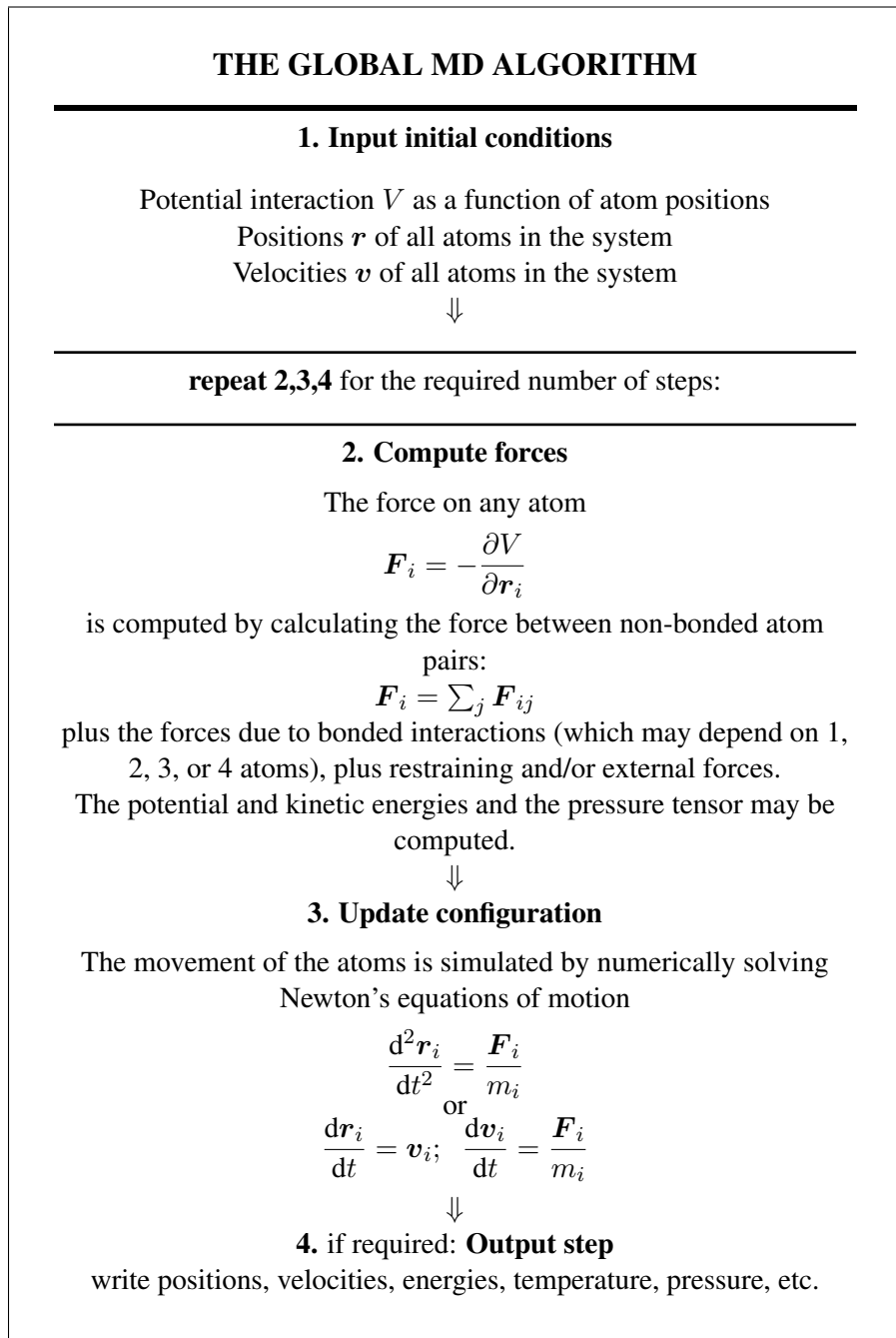


Figure 3.3: The global MD algorithm

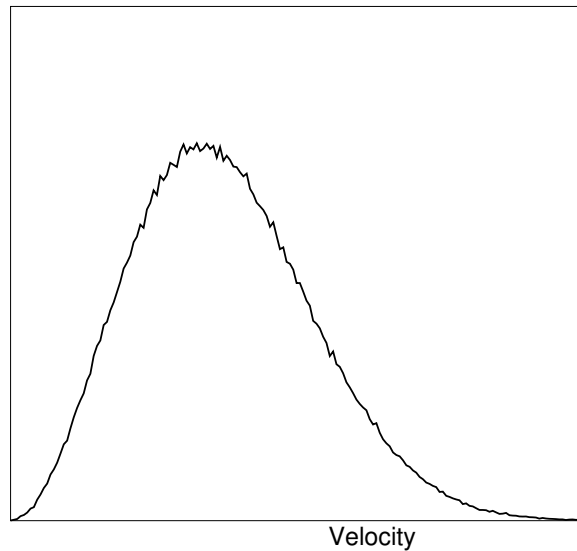


Figure 3.4: A Maxwell-Boltzmann velocity distribution, generated from random numbers.

3.4.1 Initial conditions

Topology and force field

The system topology, including a description of the force field, must be read in. Force fields and topologies are described in chapter 4 and 5, respectively. All this information is static; it is never modified during the run.

Coordinates and velocities

Then, before a run starts, the box size and the coordinates and velocities of all particles are required. The box size and shape is determined by three vectors (nine numbers) $\mathbf{b}_1, \mathbf{b}_2, \mathbf{b}_3$, which represent the three basis vectors of the periodic box.

If the run starts at $t = t_0$, the coordinates at $t = t_0$ must be known. The *leap-frog algorithm*, the default algorithm used to update the time step with Δt (see 3.4.4), also requires that the velocities at $t = t_0 - \frac{1}{2}\Delta t$ are known. If velocities are not available, the program can generate initial atomic velocities $v_i, i = 1 \dots 3N$ with a (Fig. 3.4) at a given absolute temperature T :

$$p(v_i) = \sqrt{\frac{m_i}{2\pi kT}} \exp\left(-\frac{m_i v_i^2}{2kT}\right) \quad (3.7)$$

where k is Boltzmann's constant (see chapter 2). To accomplish this, normally distributed random numbers are generated by adding twelve random numbers R_k in the range $0 \leq R_k < 1$ and subtracting 6.0 from their sum. The result is then multiplied by the standard deviation of the velocity distribution $\sqrt{kT/m_i}$. Since the resulting total energy will not correspond exactly to the required temperature T , a correction is made: first the center-of-mass motion is removed and then all velocities are scaled so that the total energy corresponds exactly to T (see eqn. 3.18).

Center-of-mass motion

The center-of-mass velocity is normally set to zero at every step; there is (usually) no net external force acting on the system and the center-of-mass velocity should remain constant. In practice, however, the update algorithm introduces a very slow change in the center-of-mass velocity, and therefore in the total kinetic energy of the system – especially when temperature coupling is used. If such changes are not quenched, an appreciable center-of-mass motion can develop in long runs, and the temperature will be significantly misinterpreted. Something similar may happen due to overall rotational motion, but only when an isolated cluster is simulated. In periodic systems with filled boxes, the overall rotational motion is coupled to other degrees of freedom and does not cause such problems.

3.4.2 Neighbor searching

As mentioned in chapter 4, internal forces are either generated from fixed (static) lists, or from dynamic lists. The latter consist of non-bonded interactions between any pair of particles. When calculating the non-bonded forces, it is convenient to have all particles in a rectangular box. As shown in Fig. 3.1, it is possible to transform a triclinic box into a rectangular box. The output coordinates are always in a rectangular box, even when a dodecahedron or triclinic box was used for the simulation. Equation 3.1 ensures that we can reset particles in a rectangular box by first shifting them with box vector \mathbf{c} , then with \mathbf{b} and finally with \mathbf{a} . Equations 3.3, 3.4 and 3.5 ensure that we can find the 14 nearest triclinic images within a linear combination that does not involve multiples of box vectors.

Pair lists generation

The non-bonded pair forces need to be calculated only for those pairs i, j for which the distance r_{ij} between i and the nearest image of j is less than a given cut-off radius R_c . Some of the particle pairs that fulfill this criterion are excluded, when their interaction is already fully accounted for by bonded interactions. GROMACS employs a *pair list* that contains those particle pairs for which non-bonded forces must be calculated. The pair list contains particles i , a displacement vector for particle i , and all particles j that are within `rlist` of this particular image of particle i . The list is updated every `nstlist` steps.

To make the neighbor list, all particles that are close (*i.e.* within the neighbor list cut-off) to a given particle must be found. This searching, usually called neighbor search (NS) or pair search, involves periodic boundary conditions and determining the *image* (see sec. 3.2). The search algorithm is $O(N)$, although a simpler $O(N^2)$ algorithm is still available under some conditions.

Cut-off schemes: group versus Verlet

From version 4.6, GROMACS supports two different cut-off scheme setups: the original one based on particle groups and one using a Verlet buffer. There are some important differences that affect results, performance and feature support. The group scheme can be made to work (almost) like the Verlet scheme, but this will lead to a decrease in performance. The group scheme is especially fast for water molecules, which are abundant in many simulations, but on the most recent x86 processors, this advantage is negated by the better instruction-level parallelism available in the Verlet-scheme implementation. The group scheme is deprecated in version 5.0, and will be removed in a future version. For practical details of choosing and setting up cut-off schemes, please see the User Guide.

In the group scheme, a neighbor list is generated consisting of pairs of groups of at least one particle. These groups were originally charge groups (see sec. 3.4.2), but with a proper treatment of long-range electrostatics, performance in unbuffered simulations is their only advantage. A pair of groups is put into the neighbor list when their center of geometry is within the cut-off distance. Interactions between all particle pairs (one from each charge group) are calculated for a certain number of MD steps, until the neighbor list is updated. This setup is efficient, as the neighbor search only checks distance between charge-group pair, not particle pairs (saves a factor of $3 \times 3 = 9$ with a three-particle water model) and the non-bonded force kernels can be optimized for, say, a water molecule “group”. Without explicit buffering, this setup leads to energy drift as some particle pairs which are within the cut-off don’t interact and some outside the cut-off do interact. This can be caused by

- particles moving across the cut-off between neighbor search steps, and/or
- for charge groups consisting of more than one particle, particle pairs moving in/out of the cut-off when their charge group center of geometry distance is outside/inside of the cut-off.

Explicitly adding a buffer to the neighbor list will remove such artifacts, but this comes at a high computational cost. How severe the artifacts are depends on the system, the properties in which you are interested, and the cut-off setup.

The Verlet cut-off scheme uses a buffered pair list by default. It also uses clusters of particles, but these are not static as in the group scheme. Rather, the clusters are defined spatially and consist of 4 or 8 particles, which is convenient for stream computing, using e.g. SSE, AVX or CUDA on GPUs. At neighbor search steps, a pair list is created with a Verlet buffer, ie. the pair-list cut-off is larger than the interaction cut-off. In the non-bonded kernels, interactions are only computed when a particle pair is within the cut-off distance at that particular time step. This ensures that as particles move between pair search steps, forces between nearly all particles within the cut-off distance are calculated. We say *nearly* all particles, because GROMACS uses a fixed pair list update frequency for efficiency. A particle-pair, whose distance was outside the cut-off, could possibly move enough during this fixed number of steps that its distance is now within the cut-off. This small chance results in a small energy drift, and the size of the chance depends on the temperature. When temperature coupling is used, the buffer size can be determined automatically, given a certain tolerance on the energy drift.

The Verlet cut-off scheme is implemented in a very efficient fashion based on clusters of particles. The simplest example is a cluster size of 4 particles. The pair list is then constructed based on

cluster pairs. The cluster-pair search is much faster searching based on particle pairs, because $4 \times 4 = 16$ particle pairs are put in the list at once. The non-bonded force calculation kernel can then calculate many particle-pair interactions at once, which maps nicely to SIMD or SIMT units on modern hardware, which can perform multiple floating operations at once. These non-bonded kernels are much faster than the kernels used in the group scheme for most types of systems, particularly on newer hardware.

Additionally, when the list buffer is determined automatically as described below, we also apply dynamic pair list pruning. The pair list can be constructed infrequently, but that can lead to a lot of pairs in the list that are outside the cut-off range for all or most of the life time of this pair list. Such pairs can be pruned out by applying a cluster-pair kernel that only determines which clusters are in range. Because of the way the non-bonded data is regularized in GROMACS, this kernel is an order of magnitude faster than the search and the interaction kernel. On the GPU this pruning is overlapped with the integration on the CPU, so it is free in most cases. Therefore we can prune every 4-10 integration steps with little overhead and significantly reduce the number of cluster pairs in the interaction kernel. This procedure is applied automatically, unless the user set the pair-list buffer size manually.

Energy drift and pair-list buffering

For a canonical (NVT) ensemble, the average energy error caused by diffusion of j particles from outside the pair-list cut-off r_ℓ to inside the interaction cut-off r_c over the lifetime of the list can be determined from the atomic displacements and the shape of the potential at the cut-off. The displacement distribution along one dimension for a freely moving particle with mass m over time t at temperature T is a Gaussian $G(x)$ of zero mean and variance $\sigma^2 = t^2 k_B T / m$. For the distance between two particles, the variance changes to $\sigma^2 = \sigma_{12}^2 = t^2 k_B T (1/m_1 + 1/m_2)$. Note that in practice particles usually interact with (bump into) other particles over time t and therefore the real displacement distribution is much narrower. Given a non-bonded interaction cut-off distance of r_c and a pair-list cut-off $r_\ell = r_c + r_b$ for r_b the Verlet buffer size, we can then write the average energy error after time t for all missing pair interactions between a single i particle of type 1 surrounded by all j particles that are of type 2 with number density ρ_2 , when the inter-particle distance changes from r_0 to r_t , as:

$$\langle \Delta V \rangle = \int_0^{r_c} \int_{r_\ell}^{\infty} 4\pi r_0^2 \rho_2 V(r_t) G\left(\frac{r_t - r_0}{\sigma}\right) dr_0 dr_t \quad (3.8)$$

To evaluate this analytically, we need to make some approximations. First we replace $V(r_t)$ by a Taylor expansion around r_c , then we can move the lower bound of the integral over r_0 to $-\infty$ which will simplify the result:

$$\begin{aligned} \langle \Delta V \rangle \approx \int_{-\infty}^{r_c} \int_{r_\ell}^{\infty} 4\pi r_0^2 \rho_2 \left[V'(r_c)(r_t - r_c) + \right. \\ V''(r_c) \frac{1}{2} (r_t - r_c)^2 + \\ V'''(r_c) \frac{1}{6} (r_t - r_c)^3 + \\ \left. O((r_t - r_c)^4) \right] G\left(\frac{r_t - r_0}{\sigma}\right) dr_0 dr_t \quad (3.9) \end{aligned}$$

Replacing the factor r_0^2 by $(r_\ell + \sigma)^2$, which results in a slight overestimate, allows us to calculate the integrals analytically:

$$\begin{aligned}
\langle \Delta V \rangle &\approx 4\pi(r_\ell + \sigma)^2 \rho_2 \int_{-\infty}^{r_c} \int_{r_\ell}^{\infty} \left[V'(r_c)(r_t - r_c) + \right. \\
&\quad \left. V''(r_c) \frac{1}{2}(r_t - r_c)^2 + \right. \\
&\quad \left. V'''(r_c) \frac{1}{6}(r_t - r_c)^3 \right] G\left(\frac{r_t - r_0}{\sigma}\right) dr_0 dr_t \quad (3.10) \\
&= 4\pi(r_\ell + \sigma)^2 \rho_2 \left\{ \frac{1}{2} V'(r_c) \left[r_b \sigma G\left(\frac{r_b}{\sigma}\right) - (r_b^2 + \sigma^2) E\left(\frac{r_b}{\sigma}\right) \right] + \right. \\
&\quad \frac{1}{6} V''(r_c) \left[\sigma(r_b^2 + 2\sigma^2) G\left(\frac{r_b}{\sigma}\right) - r_b(r_b^2 + 3\sigma^2) E\left(\frac{r_b}{\sigma}\right) \right] + \\
&\quad \frac{1}{24} V'''(r_c) \left[r_b \sigma(r_b^2 + 5\sigma^2) G\left(\frac{r_b}{\sigma}\right) \right. \\
&\quad \left. \left. - (r_b^4 + 6r_b^2 \sigma^2 + 3\sigma^4) E\left(\frac{r_b}{\sigma}\right) \right] \right\} \quad (3.11)
\end{aligned}$$

where $G(x)$ is a Gaussian distribution with 0 mean and unit variance and $E(x) = \frac{1}{2} \operatorname{erfc}(x/\sqrt{2})$. We always want to achieve small energy error, so σ will be small compared to both r_c and r_ℓ , thus the approximations in the equations above are good, since the Gaussian distribution decays rapidly. The energy error needs to be averaged over all particle pair types and weighted with the particle counts. In GROMACS we don't allow cancellation of error between pair types, so we average the absolute values. To obtain the average energy error per unit time, it needs to be divided by the neighbor-list life time $t = (\text{nstlist} - 1) \times \text{dt}$. The function can not be inverted analytically, so we use bisection to obtain the buffer size r_b for a target drift. Again we note that in practice the error we usually be much smaller than this estimate, as in the condensed phase particle displacements will be much smaller than for freely moving particles, which is the assumption used here.

When (bond) constraints are present, some particles will have fewer degrees of freedom. This will reduce the energy errors. For simplicity, we only consider one constraint per particle, the heaviest particle in case a particle is involved in multiple constraints. This simplification overestimates the displacement. The motion of a constrained particle is a superposition of the 3D motion of the center of mass of both particles and a 2D rotation around the center of mass. The displacement in an arbitrary direction of a particle with 2 degrees of freedom is not Gaussian, but rather follows the complementary error function:

$$\frac{\sqrt{\pi}}{2\sqrt{2}\sigma} \operatorname{erfc}\left(\frac{|r|}{\sqrt{2}\sigma}\right) \quad (3.12)$$

where σ^2 is again $t^2 k_B T / m$. This distribution can no longer be integrated analytically to obtain the energy error. But we can generate a tight upper bound using a scaled and shifted Gaussian distribution (not shown). This Gaussian distribution can then be used to calculate the energy error as described above. The rotation displacement around the center of mass can not be more than the length of the arm. To take this into account, we scale σ in eqn. 3.12 (details not presented here) to obtain an overestimate of the real displacement. This latter effect significantly reduces the buffer

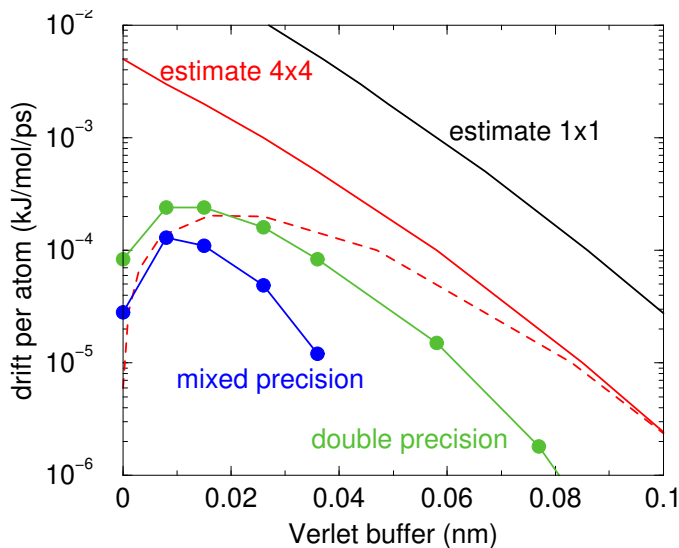


Figure 3.5: Energy drift per atom for an SPC/E water system at 300K with a time step of 2 fs and a pair-list update period of 10 steps (pair-list life time: 18 fs). PME was used with `ewald-rtol` set to 10^{-5} ; this parameter affects the shape of the potential at the cut-off. Error estimates due to finite Verlet buffer size are shown for a 1×1 atom pair list and 4×4 atom pair list without and with (dashed line) cancellation of positive and negative errors. Real energy drift is shown for simulations using double- and mixed-precision settings. Rounding errors in the SETTLE constraint algorithm from the use of single precision causes the drift to become negative at large buffer size. Note that at zero buffer size, the real drift is small because positive (H-H) and negative (O-H) energy errors cancel.

size for longer neighborlist lifetimes in e.g. water, as constrained hydrogens are by far the fastest particles, but they can not move further than 0.1 nm from the heavy atom they are connected to.

There is one important implementation detail that reduces the energy errors caused by the finite Verlet buffer list size. The derivation above assumes a particle pair-list. However, the GROMACS implementation uses a cluster pair-list for efficiency. The pair list consists of pairs of clusters of 4 particles in most cases, also called a 4×4 list, but the list can also be 4×8 (GPU CUDA kernels and AVX 256-bit single precision kernels) or 4×2 (SSE double-precision kernels). This means that the pair-list is effectively much larger than the corresponding 1×1 list. Thus slightly beyond the pair-list cut-off there will still be a large fraction of particle pairs present in the list. This fraction can be determined in a simulation and accurately estimated under some reasonable assumptions. The fraction decreases with increasing pair-list range, meaning that a smaller buffer can be used. For typical all-atom simulations with a cut-off of 0.9 nm this fraction is around 0.9, which gives a reduction in the energy errors of a factor of 10. This reduction is taken into account during the automatic Verlet buffer calculation and results in a smaller buffer size.

In Fig. 3.5 one can see that for small buffer sizes the drift of the total energy is much smaller than the pair energy error tolerance, due to cancellation of errors. For larger buffer size, the error estimate is a factor of 6 higher than drift of the total energy, or alternatively the buffer estimate is 0.024 nm too large. This is because the protons don't move freely over 18 fs, but rather vibrate.

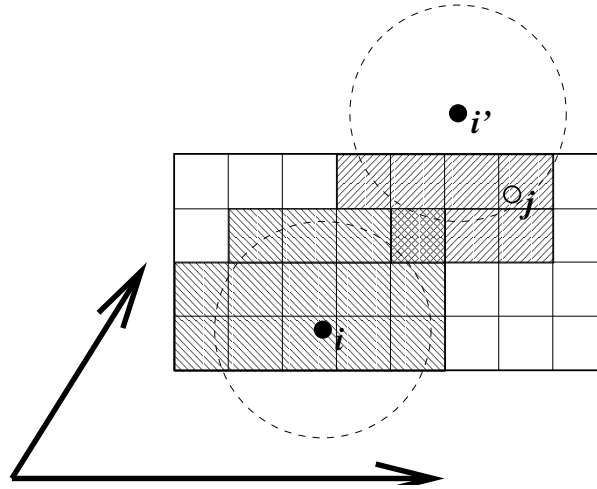


Figure 3.6: Grid search in two dimensions. The arrows are the box vectors.

Cut-off artifacts and switched interactions

With the Verlet scheme, the pair potentials are shifted to be zero at the cut-off, which makes the potential the integral of the force. This is only possible in the group scheme if the shape of the potential is such that its value is zero at the cut-off distance. However, there can still be energy drift when the forces are non-zero at the cut-off. This effect is extremely small and often not noticeable, as other integration errors (e.g. from constraints) may dominate. To completely avoid cut-off artifacts, the non-bonded forces can be switched exactly to zero at some distance smaller than the neighbor list cut-off (there are several ways to do this in GROMACS, see sec. 4.1.5). One then has a buffer with the size equal to the neighbor list cut-off less the longest interaction cut-off.

Simple search

Due to eqns. 3.1 and 3.6, the vector \mathbf{r}_{ij} connecting images within the cut-off R_c can be found by constructing:

$$\mathbf{r}''' = \mathbf{r}_j - \mathbf{r}_i \quad (3.13)$$

$$\mathbf{r}'' = \mathbf{r}''' - \mathbf{c} * \text{round}(r_z'''/c_z) \quad (3.14)$$

$$\mathbf{r}' = \mathbf{r}'' - \mathbf{b} * \text{round}(r_y''/b_y) \quad (3.15)$$

$$\mathbf{r}_{ij} = \mathbf{r}' - \mathbf{a} * \text{round}(r_x'/a_x) \quad (3.16)$$

When distances between two particles in a triclinic box are needed that do not obey eqn. 3.1, many shifts of combinations of box vectors need to be considered to find the nearest image.

Grid search

The grid search is schematically depicted in Fig. 3.6. All particles are put on the NS grid, with the smallest spacing $\geq R_c/2$ in each of the directions. In the direction of each box vector, a particle

i has three images. For each direction the image may be -1, 0 or 1, corresponding to a translation over -1, 0 or +1 box vector. We do not search the surrounding NS grid cells for neighbors of i and then calculate the image, but rather construct the images first and then search neighbors corresponding to that image of i . As Fig. 3.6 shows, some grid cells may be searched more than once for different images of i . This is not a problem, since, due to the minimum image convention, at most one image will “see” the j -particle. For every particle, fewer than 125 (5^3) neighboring cells are searched. Therefore, the algorithm scales linearly with the number of particles. Although the prefactor is large, the scaling behavior makes the algorithm far superior over the standard $O(N^2)$ algorithm when there are more than a few hundred particles. The grid search is equally fast for rectangular and triclinic boxes. Thus for most protein and peptide simulations the rhombic dodecahedron will be the preferred box shape.

Charge groups

Charge groups were originally introduced to reduce cut-off artifacts of Coulomb interactions. When a plain cut-off is used, significant jumps in the potential and forces arise when atoms with (partial) charges move in and out of the cut-off radius. When all chemical moieties have a net charge of zero, these jumps can be reduced by moving groups of atoms with net charge zero, called charge groups, in and out of the neighbor list. This reduces the cut-off effects from the charge-charge level to the dipole-dipole level, which decay much faster. With the advent of full range electrostatics methods, such as particle-mesh Ewald (sec. 4.8.2), the use of charge groups is no longer required for accuracy. It might even have a slight negative effect on the accuracy or efficiency, depending on how the neighbor list is made and the interactions are calculated.

But there is still an important reason for using “charge groups”: efficiency with the group cut-off scheme. Where applicable, neighbor searching is carried out on the basis of charge groups which are defined in the molecular topology. If the nearest image distance between the *geometrical centers* of the atoms of two charge groups is less than the cut-off radius, all atom pairs between the charge groups are included in the pair list. The neighbor searching for a water system, for instance, is $3^2 = 9$ times faster when each molecule is treated as a charge group. Also the highly optimized water force loops (see sec. A.2.1) only work when all atoms in a water molecule form a single charge group. Currently the name *neighbor-search group* would be more appropriate, but the name charge group is retained for historical reasons. When developing a new force field, the advice is to use charge groups of 3 to 4 atoms for optimal performance. For all-atom force fields this is relatively easy, as one can simply put hydrogen atoms, and in some case oxygen atoms, in the same charge group as the heavy atom they are connected to; for example: CH₃, CH₂, CH, NH₂, NH, OH, CO₂, CO.

With the Verlet cut-off scheme, charge groups are ignored.

3.4.3 Compute forces

Potential energy

When forces are computed, the potential energy of each interaction term is computed as well. The total potential energy is summed for various contributions, such as Lennard-Jones, Coulomb, and

bonded terms. It is also possible to compute these contributions for *energy-monitor groups* of atoms that are separately defined (see sec. 3.3).

Kinetic energy and temperature

The temperature is given by the total kinetic energy of the N -particle system:

$$E_{kin} = \frac{1}{2} \sum_{i=1}^N m_i v_i^2 \quad (3.17)$$

From this the absolute temperature T can be computed using:

$$\frac{1}{2} N_{df} k T = E_{kin} \quad (3.18)$$

where k is Boltzmann's constant and N_{df} is the number of degrees of freedom which can be computed from:

$$N_{df} = 3N - N_c - N_{com} \quad (3.19)$$

Here N_c is the number of *constraints* imposed on the system. When performing molecular dynamics $N_{com} = 3$ additional degrees of freedom must be removed, because the three center-of-mass velocities are constants of the motion, which are usually set to zero. When simulating in vacuo, the rotation around the center of mass can also be removed, in this case $N_{com} = 6$. When more than one temperature-coupling group is used, the number of degrees of freedom for group i is:

$$N_{df}^i = (3N^i - N_c^i) \frac{3N - N_c - N_{com}}{3N - N_c} \quad (3.20)$$

The kinetic energy can also be written as a tensor, which is necessary for pressure calculation in a triclinic system, or systems where shear forces are imposed:

$$\mathbf{E}_{kin} = \frac{1}{2} \sum_i^N m_i \mathbf{v}_i \otimes \mathbf{v}_i \quad (3.21)$$

Pressure and virial

The pressure tensor \mathbf{P} is calculated from the difference between kinetic energy E_{kin} and the virial $\mathbf{\Xi}$:

$$\mathbf{P} = \frac{2}{V} (\mathbf{E}_{kin} - \mathbf{\Xi}) \quad (3.22)$$

where V is the volume of the computational box. The scalar pressure P , which can be used for pressure coupling in the case of isotropic systems, is computed as:

$$P = \text{trace}(\mathbf{P})/3 \quad (3.23)$$

The virial $\mathbf{\Xi}$ tensor is defined as:

$$\mathbf{\Xi} = -\frac{1}{2} \sum_{i<j} \mathbf{r}_{ij} \otimes \mathbf{F}_{ij} \quad (3.24)$$

The GROMACS implementation of the virial computation is described in sec. A.1.

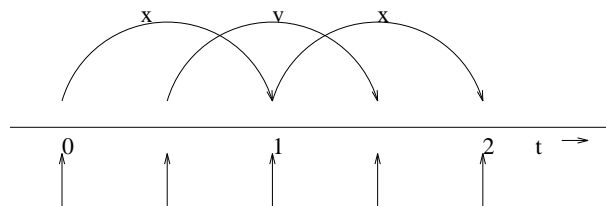


Figure 3.7: The Leap-Frog integration method. The algorithm is called Leap-Frog because \mathbf{r} and \mathbf{v} are leaping like frogs over each other's backs.

3.4.4 The leap-frog integrator

The default MD integrator in GROMACS is the so-called *leap-frog* algorithm [22] for the integration of the equations of motion. When extremely accurate integration with temperature and/or pressure coupling is required, the velocity Verlet integrators are also present and may be preferable (see 3.4.5). The leap-frog algorithm uses positions \mathbf{r} at time t and velocities \mathbf{v} at time $t - \frac{1}{2}\Delta t$; it updates positions and velocities using the forces $\mathbf{F}(t)$ determined by the positions at time t using these relations:

$$\mathbf{v}(t + \frac{1}{2}\Delta t) = \mathbf{v}(t - \frac{1}{2}\Delta t) + \frac{\Delta t}{m}\mathbf{F}(t) \quad (3.25)$$

$$\mathbf{r}(t + \Delta t) = \mathbf{r}(t) + \Delta t\mathbf{v}(t + \frac{1}{2}\Delta t) \quad (3.26)$$

The algorithm is visualized in Fig. 3.7. It produces trajectories that are identical to the Verlet [23] algorithm, whose position-update relation is

$$\mathbf{r}(t + \Delta t) = 2\mathbf{r}(t) - \mathbf{r}(t - \Delta t) + \frac{1}{m}\mathbf{F}(t)\Delta t^2 + O(\Delta t^4) \quad (3.27)$$

The algorithm is of third order in \mathbf{r} and is time-reversible. See ref. [24] for the merits of this algorithm and comparison with other time integration algorithms.

The equations of motion are modified for temperature coupling and pressure coupling, and extended to include the conservation of constraints, all of which are described below.

3.4.5 The velocity Verlet integrator

The velocity Verlet algorithm [25] is also implemented in GROMACS, though it is not yet fully integrated with all sets of options. In velocity Verlet, positions \mathbf{r} and velocities \mathbf{v} at time t are used to integrate the equations of motion; velocities at the previous half step are not required.

$$\mathbf{v}(t + \frac{1}{2}\Delta t) = \mathbf{v}(t) + \frac{\Delta t}{2m}\mathbf{F}(t) \quad (3.28)$$

$$\mathbf{r}(t + \Delta t) = \mathbf{r}(t) + \Delta t\mathbf{v}(t + \frac{1}{2}\Delta t) \quad (3.29)$$

$$\mathbf{v}(t + \Delta t) = \mathbf{v}(t + \frac{1}{2}\Delta t) + \frac{\Delta t}{2m}\mathbf{F}(t + \Delta t) \quad (3.30)$$

or, equivalently,

$$\mathbf{r}(t + \Delta t) = \mathbf{r}(t) + \Delta t \mathbf{v} + \frac{\Delta t^2}{2m} \mathbf{F}(t) \quad (3.31)$$

$$\mathbf{v}(t + \Delta t) = \mathbf{v}(t) + \frac{\Delta t}{2m} [\mathbf{F}(t) + \mathbf{F}(t + \Delta t)] \quad (3.32)$$

With no temperature or pressure coupling, and with *corresponding* starting points, leap-frog and velocity Verlet will generate identical trajectories, as can easily be verified by hand from the equations above. Given a single starting file with the *same* starting point $\mathbf{x}(0)$ and $\mathbf{v}(0)$, leap-frog and velocity Verlet will *not* give identical trajectories, as leap-frog will interpret the velocities as corresponding to $t = -\frac{1}{2}\Delta t$, while velocity Verlet will interpret them as corresponding to the timepoint $t = 0$.

3.4.6 Understanding reversible integrators: The Trotter decomposition

To further understand the relationship between velocity Verlet and leap-frog integration, we introduce the reversible Trotter formulation of dynamics, which is also useful to understanding implementations of thermostats and barostats in GROMACS.

A system of coupled, first-order differential equations can be evolved from time $t = 0$ to time t by applying the evolution operator

$$\begin{aligned} \Gamma(t) &= \exp(iLt)\Gamma(0) \\ iL &= \dot{\Gamma} \cdot \nabla_{\Gamma}, \end{aligned} \quad (3.33)$$

where L is the Liouville operator, and Γ is the multidimensional vector of independent variables (positions and velocities). A short-time approximation to the true operator, accurate at time $\Delta t = t/P$, is applied P times in succession to evolve the system as

$$\Gamma(t) = \prod_{i=1}^P \exp(iL\Delta t)\Gamma(0) \quad (3.34)$$

For NVE dynamics, the Liouville operator is

$$iL = \sum_{i=1}^N \mathbf{v}_i \cdot \nabla_{\mathbf{r}_i} + \sum_{i=1}^N \frac{1}{m_i} \mathbf{F}(r_i) \cdot \nabla_{\mathbf{v}_i}. \quad (3.35)$$

This can be split into two additive operators

$$\begin{aligned} iL_1 &= \sum_{i=1}^N \frac{1}{m_i} \mathbf{F}(r_i) \cdot \nabla_{\mathbf{v}_i} \\ iL_2 &= \sum_{i=1}^N \mathbf{v}_i \cdot \nabla_{\mathbf{r}_i} \end{aligned} \quad (3.36)$$

Then a short-time, symmetric, and thus reversible approximation of the true dynamics will be

$$\exp(iL\Delta t) = \exp(iL_2\frac{1}{2}\Delta t) \exp(iL_1\Delta t) \exp(iL_2\frac{1}{2}\Delta t) + \mathcal{O}(\Delta t^3). \quad (3.37)$$

This corresponds to velocity Verlet integration. The first exponential term over $\frac{1}{2}\Delta t$ corresponds to a velocity half-step, the second exponential term over Δt corresponds to a full velocity step, and the last exponential term over $\frac{1}{2}\Delta t$ is the final velocity half step. For future times $t = n\Delta t$, this becomes

$$\begin{aligned} \exp(iLn\Delta t) &\approx \left(\exp(iL_2\frac{1}{2}\Delta t) \exp(iL_1\Delta t) \exp(iL_2\frac{1}{2}\Delta t) \right)^n \\ &\approx \exp(iL_2\frac{1}{2}\Delta t) \left(\exp(iL_1\Delta t) \exp(iL_2\Delta t) \right)^{n-1} \\ &\quad \exp(iL_1\Delta t) \exp(iL_2\frac{1}{2}\Delta t) \end{aligned} \quad (3.38)$$

This formalism allows us to easily see the difference between the different flavors of Verlet integrators. The leap-frog integrator can be seen as starting with Eq. 3.37 with the $\exp(iL_1\Delta t)$ term, instead of the half-step velocity term, yielding

$$\exp(iLn\Delta t) = \exp(iL_1\Delta t) \exp(iL_2\Delta t) + \mathcal{O}(\Delta t^3). \quad (3.39)$$

Here, the full step in velocity is between $t - \frac{1}{2}\Delta t$ and $t + \frac{1}{2}\Delta t$, since it is a combination of the velocity half steps in velocity Verlet. For future times $t = n\Delta t$, this becomes

$$\exp(iLn\Delta t) \approx \left(\exp(iL_1\Delta t) \exp(iL_2\Delta t) \right)^n. \quad (3.40)$$

Although at first this does not appear symmetric, as long as the full velocity step is between $t - \frac{1}{2}\Delta t$ and $t + \frac{1}{2}\Delta t$, then this is simply a way of starting velocity Verlet at a different place in the cycle.

Even though the trajectory and thus potential energies are identical between leap-frog and velocity Verlet, the kinetic energy and temperature will not necessarily be the same. Standard velocity Verlet uses the velocities at the t to calculate the kinetic energy and thus the temperature only at time t ; the kinetic energy is then a sum over all particles

$$\begin{aligned} KE_{\text{full}}(t) &= \sum_i \left(\frac{1}{2m_i} \mathbf{v}_i(t) \right)^2 \\ &= \sum_i \frac{1}{2m_i} \left(\frac{1}{2} \mathbf{v}_i(t - \frac{1}{2}\Delta t) + \frac{1}{2} \mathbf{v}_i(t + \frac{1}{2}\Delta t) \right)^2, \end{aligned} \quad (3.41)$$

with the square on the *outside* of the average. Standard leap-frog calculates the kinetic energy at time t based on the average kinetic energies at the timesteps $t + \frac{1}{2}\Delta t$ and $t - \frac{1}{2}\Delta t$, or the sum over all particles

$$KE_{\text{average}}(t) = \sum_i \frac{1}{2m_i} \left(\frac{1}{2} \mathbf{v}_i(t - \frac{1}{2}\Delta t)^2 + \frac{1}{2} \mathbf{v}_i(t + \frac{1}{2}\Delta t)^2 \right), \quad (3.42)$$

where the square is *inside* the average.

A non-standard variant of velocity Verlet which averages the kinetic energies $KE(t + \frac{1}{2}\Delta t)$ and $KE(t - \frac{1}{2}\Delta t)$, exactly like leap-frog, is also now implemented in GROMACS (as `.mdp` file option `md-vv-avek`). Without temperature and pressure coupling, velocity Verlet with half-step-averaged kinetic energies and leap-frog will be identical up to numerical precision. For

temperature- and pressure-control schemes, however, velocity Verlet with half-step-averaged kinetic energies and leap-frog will be different, as will be discussed in the section on thermostats and barostats.

The half-step-averaged kinetic energy and temperature are slightly more accurate for a given step size; the difference in average kinetic energies using the half-step-averaged kinetic energies (*md* and *md-vv-avek*) will be closer to the kinetic energy obtained in the limit of small step size than will the full-step kinetic energy (using *md-vv*). For NVE simulations, this difference is usually not significant, since the positions and velocities of the particles are still identical; it makes a difference in the way the temperature of the simulations are *interpreted*, but *not* in the trajectories that are produced. Although the kinetic energy is more accurate with the half-step-averaged method, meaning that it changes less as the timestep gets large, it is also more noisy. The RMS deviation of the total energy of the system (sum of kinetic plus potential) in the half-step-averaged kinetic energy case will be higher (about twice as high in most cases) than the full-step kinetic energy. The drift will still be the same, however, as again, the trajectories are identical.

For NVT simulations, however, there *will* be a difference, as discussed in the section on temperature control, since the velocities of the particles are adjusted such that kinetic energies of the simulations, which can be calculated either way, reach the distribution corresponding to the set temperature. In this case, the three methods will not give identical results.

Because the velocity and position are both defined at the same time t the velocity Verlet integrator can be used for some methods, especially rigorously correct pressure control methods, that are not actually possible with leap-frog. The integration itself takes negligibly more time than leap-frog, but twice as many communication calls are currently required. In most cases, and especially for large systems where communication speed is important for parallelization and differences between thermodynamic ensembles vanish in the $1/N$ limit, and when only NVT ensembles are required, leap-frog will likely be the preferred integrator. For pressure control simulations where the fine details of the thermodynamics are important, only velocity Verlet allows the true ensemble to be calculated. In either case, simulation with double precision may be required to get fine details of thermodynamics correct.

3.4.7 Multiple time stepping

Several other simulation packages use multiple time stepping for bonds and/or the PME mesh forces. In GROMACS we have not implemented this (yet), since we use a different philosophy. Bonds can be constrained (which is also a more sound approximation of a physical quantum oscillator), which allows the smallest time step to be increased to the larger one. This not only halves the number of force calculations, but also the update calculations. For even larger time steps, angle vibrations involving hydrogen atoms can be removed using virtual interaction sites (see sec. 6.10), which brings the shortest time step up to PME mesh update frequency of a multiple time stepping scheme.

3.4.8 Temperature coupling

While direct use of molecular dynamics gives rise to the NVE (constant number, constant volume, constant energy ensemble), most quantities that we wish to calculate are actually from a

constant temperature (NVT) ensemble, also called the canonical ensemble. GROMACS can use the *weak-coupling* scheme of Berendsen [26], stochastic randomization through the Andersen thermostat [27], the extended ensemble Nosé-Hoover scheme [28, 29], or a velocity-rescaling scheme [30] to simulate constant temperature, with advantages of each of the schemes laid out below.

There are several other reasons why it might be necessary to control the temperature of the system (drift during equilibration, drift as a result of force truncation and integration errors, heating due to external or frictional forces), but this is not entirely correct to do from a thermodynamic standpoint, and in some cases only masks the symptoms (increase in temperature of the system) rather than the underlying problem (deviations from correct physics in the dynamics). For larger systems, errors in ensemble averages and structural properties incurred by using temperature control to remove slow drifts in temperature appear to be negligible, but no completely comprehensive comparisons have been carried out, and some caution must be taken in interpreting the results.

When using temperature and/or pressure coupling the total energy is no longer conserved. Instead there is a conserved energy quantity the formula of which will depend on the combination of temperature and pressure coupling algorithm used. For all coupling algorithms, except for Andersen temperature coupling and Parrinello-Rahman pressure coupling combined with shear stress, the conserved energy quantity is computed and stored in the energy and log file. Note that this quantity will not be conserved when external forces are applied to the system, such as pulling on a group with a changing distance or an electric field. Furthermore, how well the energy is conserved depends on the accuracy of all algorithms involved in the simulation. Usually the algorithms that cause most drift are constraints and the pair-list buffer, depending on the parameters used.

Berendsen temperature coupling

The Berendsen algorithm mimics weak coupling with first-order kinetics to an external heat bath with given temperature T_0 . See ref. [31] for a comparison with the Nosé-Hoover scheme. The effect of this algorithm is that a deviation of the system temperature from T_0 is slowly corrected according to:

$$\frac{dT}{dt} = \frac{T_0 - T}{\tau} \quad (3.43)$$

which means that a temperature deviation decays exponentially with a time constant τ . This method of coupling has the advantage that the strength of the coupling can be varied and adapted to the user requirement: for equilibration purposes the coupling time can be taken quite short (*e.g.* 0.01 ps), but for reliable equilibrium runs it can be taken much longer (*e.g.* 0.5 ps) in which case it hardly influences the conservative dynamics.

The Berendsen thermostat suppresses the fluctuations of the kinetic energy. This means that one does not generate a proper canonical ensemble, so rigorously, the sampling will be incorrect. This error scales with $1/N$, so for very large systems most ensemble averages will not be affected significantly, except for the distribution of the kinetic energy itself. However, fluctuation properties, such as the heat capacity, will be affected. A similar thermostat which does produce a correct ensemble is the velocity rescaling thermostat [30] described below.

The heat flow into or out of the system is affected by scaling the velocities of each particle every

step, or every n_{TC} steps, with a time-dependent factor λ , given by:

$$\lambda = \left[1 + \frac{n_{TC}\Delta t}{\tau_T} \left\{ \frac{T_0}{T(t - \frac{1}{2}\Delta t)} - 1 \right\} \right]^{1/2} \quad (3.44)$$

The parameter τ_T is close, but not exactly equal, to the time constant τ of the temperature coupling (eqn. 3.43):

$$\tau = 2C_V\tau_T/N_{df}k \quad (3.45)$$

where C_V is the total heat capacity of the system, k is Boltzmann's constant, and N_{df} is the total number of degrees of freedom. The reason that $\tau \neq \tau_T$ is that the kinetic energy change caused by scaling the velocities is partly redistributed between kinetic and potential energy and hence the change in temperature is less than the scaling energy. In practice, the ratio τ/τ_T ranges from 1 (gas) to 2 (harmonic solid) to 3 (water). When we use the term "temperature coupling time constant," we mean the parameter τ_T . **Note** that in practice the scaling factor λ is limited to the range of $0.8 \leq \lambda \leq 1.25$, to avoid scaling by very large numbers which may crash the simulation. In normal use, λ will always be much closer to 1.0.

The thermostat modifies the kinetic energy at each scaling step by:

$$\Delta E_k = (\lambda - 1)^2 E_k \quad (3.46)$$

The sum of these changes over the run needs to be subtracted from the total energy to obtain the conserved energy quantity.

Velocity-rescaling temperature coupling

The velocity-rescaling thermostat [30] is essentially a Berendsen thermostat (see above) with an additional stochastic term that ensures a correct kinetic energy distribution by modifying it according to

$$dK = (K_0 - K) \frac{dt}{\tau_T} + 2\sqrt{\frac{KK_0}{N_f}} \frac{dW}{\sqrt{\tau_T}}, \quad (3.47)$$

where K is the kinetic energy, N_f the number of degrees of freedom and dW a Wiener process. There are no additional parameters, except for a random seed. This thermostat produces a correct canonical ensemble and still has the advantage of the Berendsen thermostat: first order decay of temperature deviations and no oscillations.

Andersen thermostat

One simple way to maintain a thermostatted ensemble is to take an NVE integrator and periodically re-select the velocities of the particles from a Maxwell-Boltzmann distribution. [27] This can either be done by randomizing all the velocities simultaneously (massive collision) every $\tau_T/\Delta t$ steps (andersen-massive), or by randomizing every particle with some small probability every timestep (andersen), equal to $\Delta t/\tau$, where in both cases Δt is the timestep and τ_T is a characteristic coupling time scale. Because of the way constraints operate, all particles in the same constraint group must be randomized simultaneously. Because of parallelization issues, the `andersen` version cannot currently (5.0) be used in systems with constraints.

`andersen-massive` can be used regardless of constraints. This thermostat is also currently only possible with velocity Verlet algorithms, because it operates directly on the velocities at each timestep.

This algorithm completely avoids some of the ergodicity issues of other thermostating algorithms, as energy cannot flow back and forth between energetically decoupled components of the system as in velocity scaling motions. However, it can slow down the kinetics of system by randomizing correlated motions of the system, including slowing sampling when τ_T is at moderate levels (less than 10 ps). This algorithm should therefore generally not be used when examining kinetics or transport properties of the system. [32]

Nosé-Hoover temperature coupling

The Berendsen weak-coupling algorithm is extremely efficient for relaxing a system to the target temperature, but once the system has reached equilibrium it might be more important to probe a correct canonical ensemble. This is unfortunately not the case for the weak-coupling scheme.

To enable canonical ensemble simulations, GROMACS also supports the extended-ensemble approach first proposed by Nosé [28] and later modified by Hoover [29]. The system Hamiltonian is extended by introducing a thermal reservoir and a friction term in the equations of motion. The friction force is proportional to the product of each particle’s velocity and a friction parameter, ξ . This friction parameter (or “heat bath” variable) is a fully dynamic quantity with its own momentum (p_ξ) and equation of motion; the time derivative is calculated from the difference between the current kinetic energy and the reference temperature.

In this formulation, the particles’ equations of motion in Fig. 3.3 are replaced by:

$$\frac{d^2 \mathbf{r}_i}{dt^2} = \frac{\mathbf{F}_i}{m_i} - \frac{p_\xi}{Q} \frac{d\mathbf{r}_i}{dt}, \quad (3.48)$$

where the equation of motion for the heat bath parameter ξ is:

$$\frac{dp_\xi}{dt} = (T - T_0). \quad (3.49)$$

The reference temperature is denoted T_0 , while T is the current instantaneous temperature of the system. The strength of the coupling is determined by the constant Q (usually called the “mass parameter” of the reservoir) in combination with the reference temperature.¹

The conserved quantity for the Nosé-Hoover equations of motion is not the total energy, but rather

$$H = \sum_{i=1}^N \frac{\mathbf{p}_i^2}{2m_i} + U(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_N) + \frac{p_\xi^2}{2Q} + N_f k T \xi, \quad (3.50)$$

where N_f is the total number of degrees of freedom.

In our opinion, the mass parameter is a somewhat awkward way of describing coupling strength, especially due to its dependence on reference temperature (and some implementations even include the number of degrees of freedom in your system when defining Q). To maintain the coupling strength, one would have to change Q in proportion to the change in reference temperature.

¹Note that some derivations, an alternative notation $\xi_{\text{alt}} = v_\xi = p_\xi/Q$ is used.

For this reason, we prefer to let the GROMACS user work instead with the period τ_T of the oscillations of kinetic energy between the system and the reservoir instead. It is directly related to Q and T_0 via:

$$Q = \frac{\tau_T^2 T_0}{4\pi^2}. \quad (3.51)$$

This provides a much more intuitive way of selecting the Nosé-Hoover coupling strength (similar to the weak-coupling relaxation), and in addition τ_T is independent of system size and reference temperature.

It is however important to keep the difference between the weak-coupling scheme and the Nosé-Hoover algorithm in mind: Using weak coupling you get a strongly damped *exponential relaxation*, while the Nosé-Hoover approach produces an *oscillatory relaxation*. The actual time it takes to relax with Nosé-Hoover coupling is several times larger than the period of the oscillations that you select. These oscillations (in contrast to exponential relaxation) also means that the time constant normally should be 4–5 times larger than the relaxation time used with weak coupling, but your mileage may vary.

Nosé-Hoover dynamics in simple systems such as collections of harmonic oscillators, can be *non-ergodic*, meaning that only a subsection of phase space is ever sampled, even if the simulations were to run for infinitely long. For this reason, the Nosé-Hoover chain approach was developed, where each of the Nosé-Hoover thermostats has its own Nosé-Hoover thermostat controlling its temperature. In the limit of an infinite chain of thermostats, the dynamics are guaranteed to be ergodic. Using just a few chains can greatly improve the ergodicity, but recent research has shown that the system will still be nonergodic, and it is still not entirely clear what the practical effect of this [33]. Currently, the default number of chains is 10, but this can be controlled by the user. In the case of chains, the equations are modified in the following way to include a chain of thermostating particles [34]:

$$\begin{aligned} \frac{d^2 \mathbf{r}_i}{dt^2} &= \frac{\mathbf{F}_i}{m_i} - \frac{p_{\xi_1}}{Q_1} \frac{d\mathbf{r}_i}{dt} \\ \frac{dp_{\xi_1}}{dt} &= (T - T_0) - p_{\xi_1} \frac{p_{\xi_2}}{Q_2} \\ \frac{dp_{\xi_{i=2\dots N}}}{dt} &= \left(\frac{p_{\xi_{i-1}}^2}{Q_{i-1}} - kT \right) - p_{\xi_i} \frac{p_{\xi_{i+1}}}{Q_{i+1}} \\ \frac{dp_{\xi_N}}{dt} &= \left(\frac{p_{\xi_{N-1}}^2}{Q_{N-1}} - kT \right) \end{aligned} \quad (3.52)$$

The conserved quantity for Nosé-Hoover chains is

$$H = \sum_{i=1}^N \frac{\mathbf{p}_i^2}{2m_i} + U(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_N) + \sum_{k=1}^M \frac{p_{\xi_k}^2}{2Q'_k} + N_f kT \xi_1 + kT \sum_{k=2}^M \xi_k \quad (3.53)$$

The values and velocities of the Nosé-Hoover thermostat variables are generally not included in the output, as they take up a fair amount of space and are generally not important for analysis of simulations, but by setting an `mdp` option the values of all the positions and velocities of all

Nosé-Hoover particles in the chain are written to the `.edr` file. Leap-frog simulations currently can only have Nosé-Hoover chain lengths of 1, but this will likely be updated in later version.

As described in the integrator section, for temperature coupling, the temperature that the algorithm attempts to match to the reference temperature is calculated differently in velocity Verlet and leap-frog dynamics. Velocity Verlet (*md-vv*) uses the full-step kinetic energy, while leap-frog and *md-vv-avek* use the half-step-averaged kinetic energy.

We can examine the Trotter decomposition again to better understand the differences between these constant-temperature integrators. In the case of Nosé-Hoover dynamics (for simplicity, using a chain with $N = 1$, with more details in Ref. [35]), we split the Liouville operator as

$$iL = iL_1 + iL_2 + iL_{\text{NHC}}, \quad (3.54)$$

where

$$\begin{aligned} iL_1 &= \sum_{i=1}^N \left[\frac{\mathbf{p}_i}{m_i} \right] \cdot \frac{\partial}{\partial \mathbf{r}_i} \\ iL_2 &= \sum_{i=1}^N \mathbf{F}_i \cdot \frac{\partial}{\partial \mathbf{p}_i} \\ iL_{\text{NHC}} &= \sum_{i=1}^N -\frac{p_\xi}{Q} \mathbf{v}_i \cdot \nabla_{\mathbf{v}_i} + \frac{p_\xi}{Q} \frac{\partial}{\partial \xi} + (T - T_0) \frac{\partial}{\partial p_\xi} \end{aligned} \quad (3.55)$$

For standard velocity Verlet with Nosé-Hoover temperature control, this becomes

$$\begin{aligned} \exp(iL\Delta t) &= \exp(iL_{\text{NHC}}\Delta t/2) \exp(iL_2\Delta t/2) \\ &\quad \exp(iL_1\Delta t) \exp(iL_2\Delta t/2) \exp(iL_{\text{NHC}}\Delta t/2) + \mathcal{O}(\Delta t^3). \end{aligned} \quad (3.56)$$

For half-step-averaged temperature control using *md-vv-avek*, this decomposition will not work, since we do not have the full step temperature until after the second velocity step. However, we can construct an alternate decomposition that is still reversible, by switching the place of the NHC and velocity portions of the decomposition:

$$\begin{aligned} \exp(iL\Delta t) &= \exp(iL_2\Delta t/2) \exp(iL_{\text{NHC}}\Delta t/2) \exp(iL_1\Delta t) \\ &\quad \exp(iL_{\text{NHC}}\Delta t/2) \exp(iL_2\Delta t/2) + \mathcal{O}(\Delta t^3) \end{aligned} \quad (3.57)$$

This formalism allows us to easily see the difference between the different flavors of velocity Verlet integrator. The leap-frog integrator can be seen as starting with Eq. 3.57 just before the $\exp(iL_1\Delta t)$ term, yielding:

$$\begin{aligned} \exp(iL\Delta t) &= \exp(iL_1\Delta t) \exp(iL_{\text{NHC}}\Delta t/2) \\ &\quad \exp(iL_2\Delta t) \exp(iL_{\text{NHC}}\Delta t/2) + \mathcal{O}(\Delta t^3) \end{aligned} \quad (3.58)$$

and then using some algebra tricks to solve for some quantities are required before they are actually calculated [36].

Group temperature coupling

In GROMACS temperature coupling can be performed on groups of atoms, typically a protein and solvent. The reason such algorithms were introduced is that energy exchange between different components is not perfect, due to different effects including cut-offs etc. If now the whole system is coupled to one heat bath, water (which experiences the largest cut-off noise) will tend to heat up and the protein will cool down. Typically 100 K differences can be obtained. With the use of proper electrostatic methods (PME) these difference are much smaller but still not negligible. The parameters for temperature coupling in groups are given in the `mdp` file. Recent investigation has shown that small temperature differences between protein and water may actually be an artifact of the way temperature is calculated when there are finite timesteps, and very large differences in temperature are likely a sign of something else seriously going wrong with the system, and should be investigated carefully [37].

One special case should be mentioned: it is possible to temperature-couple only part of the system, leaving other parts without temperature coupling. This is done by specifying -1 for the time constant τ_T for the group that should not be thermostatted. If only part of the system is thermostatted, the system will still eventually converge to an NVT system. In fact, one suggestion for minimizing errors in the temperature caused by discretized timesteps is that if constraints on the water are used, then only the water degrees of freedom should be thermostatted, not protein degrees of freedom, as the higher frequency modes in the protein can cause larger deviations from the “true” temperature, the temperature obtained with small timesteps [37].

3.4.9 Pressure coupling

In the same spirit as the temperature coupling, the system can also be coupled to a “pressure bath.” GROMACS supports both the Berendsen algorithm [26] that scales coordinates and box vectors every step, the extended-ensemble Parrinello-Rahman approach [38, 39], and for the velocity Verlet variants, the Martyna-Tuckerman-Tobias-Klein (MTTK) implementation of pressure control [35]. Parrinello-Rahman and Berendsen can be combined with any of the temperature coupling methods above. MTTK can only be used with Nosé-Hoover temperature control. From 5.1 afterwards, it can only used when the system does not have constraints.

Berendsen pressure coupling

The Berendsen algorithm rescales the coordinates and box vectors every step, or every n_{PC} steps, with a matrix μ , which has the effect of a first-order kinetic relaxation of the pressure towards a given reference pressure \mathbf{P}_0 according to

$$\frac{d\mathbf{P}}{dt} = \frac{\mathbf{P}_0 - \mathbf{P}}{\tau_p}. \quad (3.59)$$

The scaling matrix μ is given by

$$\mu_{ij} = \delta_{ij} - \frac{n_{PC}\Delta t}{3\tau_p}\beta_{ij}\{P_{0ij} - P_{ij}(t)\}. \quad (3.60)$$

Here, β is the isothermal compressibility of the system. In most cases this will be a diagonal matrix, with equal elements on the diagonal, the value of which is generally not known. It suffices to take a rough estimate because the value of β only influences the non-critical time constant of the pressure relaxation without affecting the average pressure itself. For water at 1 atm and 300 K $\beta = 4.6 \times 10^{-10} \text{ Pa}^{-1} = 4.6 \times 10^{-5} \text{ bar}^{-1}$, which is 7.6×10^{-4} MD units (see chapter 2). Most other liquids have similar values. When scaling completely anisotropically, the system has to be rotated in order to obey eqn. 3.1. This rotation is approximated in first order in the scaling, which is usually less than 10^{-4} . The actual scaling matrix $\boldsymbol{\mu}'$ is

$$\boldsymbol{\mu}' = \begin{pmatrix} \mu_{xx} & \mu_{xy} + \mu_{yx} & \mu_{xz} + \mu_{zx} \\ 0 & \mu_{yy} & \mu_{yz} + \mu_{zy} \\ 0 & 0 & \mu_{zz} \end{pmatrix}. \quad (3.61)$$

The velocities are neither scaled nor rotated. Since the equations of motion are modified by pressure coupling, the conserved energy quantity also needs to be modified. For first order pressure coupling, the work the barostat applies to the system every step needs to be subtracted from the total energy to obtain the conserved energy quantity:

$$-\sum_{i,j} (\mu_{ij} - \delta_{ij}) P_{ij} V = \sum_{i,j} 2(\mu_{ij} - \delta_{ij}) \Xi_{ij} \quad (3.62)$$

where δ_{ij} is the Kronecker delta and Ξ is the virial. Note that the factor 2 originates from the factor $\frac{1}{2}$ in the virial definition (eqn. 3.24).

In GROMACS, the Berendsen scaling can also be done isotropically, which means that instead of \boldsymbol{P} a diagonal matrix with elements of size $\text{trace}(\boldsymbol{P})/3$ is used. For systems with interfaces, semi-isotropic scaling can be useful. In this case, the x/y -directions are scaled isotropically and the z direction is scaled independently. The compressibility in the x/y or z -direction can be set to zero, to scale only in the other direction(s).

If you allow full anisotropic deformations and use constraints you might have to scale more slowly or decrease your timestep to avoid errors from the constraint algorithms. It is important to note that although the Berendsen pressure control algorithm yields a simulation with the correct average pressure, it does not yield the exact NPT ensemble, and it is not yet clear exactly what errors this approximation may yield.

Parrinello-Rahman pressure coupling

In cases where the fluctuations in pressure or volume are important *per se* (e.g. to calculate thermodynamic properties), especially for small systems, it may be a problem that the exact ensemble is not well defined for the weak-coupling scheme, and that it does not simulate the true NPT ensemble.

GROMACS also supports constant-pressure simulations using the Parrinello-Rahman approach [38, 39], which is similar to the Nosé-Hoover temperature coupling, and in theory gives the true NPT ensemble. With the Parrinello-Rahman barostat, the box vectors as represented by the matrix \boldsymbol{b}

obey the matrix equation of motion²

$$\frac{d\mathbf{b}^2}{dt^2} = V\mathbf{W}^{-1}\mathbf{b}'^{-1}(\mathbf{P} - \mathbf{P}_{ref}). \quad (3.63)$$

The volume of the box is denoted V , and \mathbf{W} is a matrix parameter that determines the strength of the coupling. The matrices \mathbf{P} and \mathbf{P}_{ref} are the current and reference pressures, respectively.

The equations of motion for the particles are also changed, just as for the Nosé-Hoover coupling. In most cases you would combine the Parrinello-Rahman barostat with the Nosé-Hoover thermostat, but to keep it simple we only show the Parrinello-Rahman modification here. The modified Hamiltonian, which will be conserved, is:

$$E_{\text{pot}} + E_{\text{kin}} + \sum_i P_{ii}V + \sum_{i,j} \frac{1}{2}W_{ij} \left(\frac{db_{ij}}{dt} \right)^2 \quad (3.64)$$

The equations of motion for the atoms, obtained from the Hamiltonian are:

$$\frac{d^2\mathbf{r}_i}{dt^2} = \frac{\mathbf{F}_i}{m_i} - \mathbf{M} \frac{d\mathbf{r}_i}{dt}, \quad (3.65)$$

$$\mathbf{M} = \mathbf{b}^{-1} \left[\mathbf{b} \frac{d\mathbf{b}'}{dt} + \frac{d\mathbf{b}}{dt} \mathbf{b}' \right] \mathbf{b}'^{-1}. \quad (3.66)$$

This extra term has the appearance of a friction, but it should be noted that it is fictitious, and rather an effect of the Parrinello-Rahman equations of motion being defined with all particle coordinates represented relative to the box vectors, while GROMACS] uses normal Cartesian coordinates for positions, velocities and forces. It is worth noting that the kinetic energy too should formally be calculated based on velocities relative to the box vectors. This can have an effect e.g. for external constant stress, but for now we only support coupling to constant external pressures, and for any normal simulation the velocities of box vectors should be extremely small compared to particle velocities. Gang Liu has done some work on deriving this for Cartesian coordinates[40] that we will try to implement at some point in the future together with support for external stress.

The (inverse) mass parameter matrix \mathbf{W}^{-1} determines the strength of the coupling, and how the box can be deformed. The box restriction (3.1) will be fulfilled automatically if the corresponding elements of \mathbf{W}^{-1} are zero. Since the coupling strength also depends on the size of your box, we prefer to calculate it automatically in GROMACS. You only have to provide the approximate isothermal compressibilities β and the pressure time constant τ_p in the input file (L is the largest box matrix element):

$$\left(\mathbf{W}^{-1} \right)_{ij} = \frac{4\pi^2\beta_{ij}}{3\tau_p^2 L}. \quad (3.67)$$

Just as for the Nosé-Hoover thermostat, you should realize that the Parrinello-Rahman time constant is *not* equivalent to the relaxation time used in the Berendsen pressure coupling algorithm. In most cases you will need to use a 4–5 times larger time constant with Parrinello-Rahman coupling. If your pressure is very far from equilibrium, the Parrinello-Rahman coupling may result in very large box oscillations that could even crash your run. In that case you would have to increase

²The box matrix representation \mathbf{b} in GROMACS corresponds to the transpose of the box matrix representation \mathbf{h} in the paper by Nosé and Klein. Because of this, some of our equations will look slightly different.

the time constant, or (better) use the weak-coupling scheme to reach the target pressure, and then switch to Parrinello-Rahman coupling once the system is in equilibrium. Additionally, using the leap-frog algorithm, the pressure at time t is not available until after the time step has completed, and so the pressure from the previous step must be used, which makes the algorithm not directly reversible, and may not be appropriate for high precision thermodynamic calculations.

Surface-tension coupling

When a periodic system consists of more than one phase, separated by surfaces which are parallel to the xy -plane, the surface tension and the z -component of the pressure can be coupled to a pressure bath. Presently, this only works with the Berendsen pressure coupling algorithm in GROMACS. The average surface tension $\gamma(t)$ can be calculated from the difference between the normal and the lateral pressure

$$\gamma(t) = \frac{1}{n} \int_0^{L_z} \left\{ P_{zz}(z, t) - \frac{P_{xx}(z, t) + P_{yy}(z, t)}{2} \right\} dz \quad (3.68)$$

$$= \frac{L_z}{n} \left\{ P_{zz}(t) - \frac{P_{xx}(t) + P_{yy}(t)}{2} \right\}, \quad (3.69)$$

where L_z is the height of the box and n is the number of surfaces. The pressure in the z -direction is corrected by scaling the height of the box with μ_{zz}

$$\Delta P_{zz} = \frac{\Delta t}{\tau_p} \{ P_{0zz} - P_{zz}(t) \} \quad (3.70)$$

$$\mu_{zz} = 1 + \beta_{zz} \Delta P_{zz} \quad (3.71)$$

This is similar to normal pressure coupling, except that the factor of $1/3$ is missing. The pressure correction in the z -direction is then used to get the correct convergence for the surface tension to the reference value γ_0 . The correction factor for the box length in the x/y -direction is

$$\mu_{x/y} = 1 + \frac{\Delta t}{2 \tau_p} \beta_{x/y} \left(\frac{n \gamma_0}{\mu_{zz} L_z} - \left\{ P_{zz}(t) + \Delta P_{zz} - \frac{P_{xx}(t) + P_{yy}(t)}{2} \right\} \right) \quad (3.72)$$

The value of β_{zz} is more critical than with normal pressure coupling. Normally an incorrect compressibility will just scale τ_p , but with surface tension coupling it affects the convergence of the surface tension. When β_{zz} is set to zero (constant box height), ΔP_{zz} is also set to zero, which is necessary for obtaining the correct surface tension.

MTTK pressure control algorithms

As mentioned in the previous section, one weakness of leap-frog integration is in constant pressure simulations, since the pressure requires a calculation of both the virial and the kinetic energy at the full time step; for leap-frog, this information is not available until *after* the full timestep. Velocity Verlet does allow the calculation, at the cost of an extra round of global communication, and can compute, mod any integration errors, the true NPT ensemble.

The full equations, combining both pressure coupling and temperature coupling, are taken from Martyna *et al.* [35] and Tuckerman [41] and are referred to here as MTTK equations (Martyna-Tuckerman-Tobias-Klein). We introduce for convenience $\epsilon = (1/3) \ln(V/V_0)$, where V_0 is a

reference volume. The momentum of ϵ is $v_\epsilon = p_\epsilon/W = \dot{\epsilon} = \dot{V}/3V$, and define $\alpha = 1 + 3/N_{dof}$ (see Ref [41])

The isobaric equations are

$$\begin{aligned}\dot{\mathbf{r}}_i &= \frac{\mathbf{p}_i}{m_i} + \frac{p_\epsilon}{W} \mathbf{r}_i \\ \frac{\dot{\mathbf{p}}_i}{m_i} &= \frac{1}{m_i} \mathbf{F}_i - \alpha \frac{p_\epsilon}{W} \frac{\mathbf{p}_i}{m_i} \\ \dot{\epsilon} &= \frac{p_\epsilon}{W} \\ \frac{\dot{p}_\epsilon}{W} &= \frac{3V}{W} (P_{\text{int}} - P) + (\alpha - 1) \left(\sum_{n=1}^N \frac{\mathbf{p}_i^2}{m_i} \right),\end{aligned}\tag{3.73}$$

$$\tag{3.74}$$

where

$$P_{\text{int}} = P_{\text{kin}} - P_{\text{vir}} = \frac{1}{3V} \left[\sum_{i=1}^N \left(\frac{\mathbf{p}_i^2}{2m_i} - \mathbf{r}_i \cdot \mathbf{F}_i \right) \right].\tag{3.75}$$

The terms including α are required to make phase space incompressible [41]. The ϵ acceleration term can be rewritten as

$$\frac{\dot{p}_\epsilon}{W} = \frac{3V}{W} (\alpha P_{\text{kin}} - P_{\text{vir}} - P)\tag{3.76}$$

In terms of velocities, these equations become

$$\begin{aligned}\dot{\mathbf{r}}_i &= \mathbf{v}_i + v_\epsilon \mathbf{r}_i \\ \dot{\mathbf{v}}_i &= \frac{1}{m_i} \mathbf{F}_i - \alpha v_\epsilon \mathbf{v}_i \\ \dot{\epsilon} &= v_\epsilon \\ \dot{v}_\epsilon &= \frac{3V}{W} (P_{\text{int}} - P) + (\alpha - 1) \left(\sum_{n=1}^N \frac{1}{2} m_i \mathbf{v}_i^2 \right) \\ P_{\text{int}} &= P_{\text{kin}} - P_{\text{vir}} = \frac{1}{3V} \left[\sum_{i=1}^N \left(\frac{1}{2} m_i \mathbf{v}_i^2 - \mathbf{r}_i \cdot \mathbf{F}_i \right) \right]\end{aligned}\tag{3.77}$$

For these equations, the conserved quantity is

$$H = \sum_{i=1}^N \frac{\mathbf{p}_i^2}{2m_i} + U(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_N) + \frac{p_\epsilon}{2W} + PV\tag{3.78}$$

The next step is to add temperature control. Adding Nosé-Hoover chains, including to the barostat degree of freedom, where we use η for the barostat Nosé-Hoover variables, and Q' for the coupling constants of the thermostats of the barostats, we get

$$\dot{\mathbf{r}}_i = \frac{\mathbf{p}_i}{m_i} + \frac{p_\epsilon}{W} \mathbf{r}_i$$

$$\begin{aligned}
\frac{\dot{\mathbf{p}}_i}{m_i} &= \frac{1}{m_i} \mathbf{F}_i - \alpha \frac{p_\epsilon}{W} \frac{\mathbf{p}_i}{m_i} - \frac{p_{\xi_1}}{Q_1} \frac{\mathbf{p}_i}{m_i} \\
\dot{\epsilon} &= \frac{p_\epsilon}{W} \\
\frac{\dot{p}_\epsilon}{W} &= \frac{3V}{W} (\alpha P_{\text{kin}} - P_{\text{vir}} - P) - \frac{p_{\eta_1}}{Q'_1} p_\epsilon \\
\dot{\xi}_k &= \frac{p_{\xi_k}}{Q_k} \\
\dot{\eta}_k &= \frac{p_{\eta_k}}{Q'_k} \\
\dot{p}_{\xi_k} &= G_k - \frac{p_{\xi_{k+1}}}{Q_{k+1}} \quad k = 1, \dots, M-1 \\
\dot{p}_{\eta_k} &= G'_k - \frac{p_{\eta_{k+1}}}{Q'_{k+1}} \quad k = 1, \dots, M-1 \\
\dot{p}_{\xi_M} &= G_M \\
\dot{p}_{\eta_M} &= G'_M,
\end{aligned} \tag{3.79}$$

where

$$\begin{aligned}
P_{\text{int}} &= P_{\text{kin}} - P_{\text{vir}} = \frac{1}{3V} \left[\sum_{i=1}^N \left(\frac{\mathbf{p}_i^2}{2m_i} - \mathbf{r}_i \cdot \mathbf{F}_i \right) \right] \\
G_1 &= \sum_{i=1}^N \frac{\mathbf{p}_i^2}{m_i} - N_f kT \\
G_k &= \frac{p_{\xi_{k-1}}^2}{2Q_{k-1}} - kT \quad k = 2, \dots, M \\
G'_1 &= \frac{p_\epsilon^2}{2W} - kT \\
G'_k &= \frac{p_{\eta_{k-1}}^2}{2Q'_{k-1}} - kT \quad k = 2, \dots, M
\end{aligned} \tag{3.80}$$

The conserved quantity is now

$$\begin{aligned}
H &= \sum_{i=1}^N \frac{\mathbf{p}_i^2}{2m_i} + U(\mathbf{r}_1, \mathbf{r}_2, \dots, \mathbf{r}_N) + \frac{p_\epsilon^2}{2W} + PV + \\
&\sum_{k=1}^M \frac{p_{\xi_k}^2}{2Q_k} + \sum_{k=1}^M \frac{p_{\eta_k}^2}{2Q'_k} + N_f kT \xi_1 + kT \sum_{i=2}^M \xi_k + kT \sum_{k=1}^M \eta_k
\end{aligned} \tag{3.81}$$

Returning to the Trotter decomposition formalism, for pressure control and temperature control [35] we get:

$$iL = iL_1 + iL_2 + iL_{\epsilon,1} + iL_{\epsilon,2} + iL_{\text{NHC-baro}} + iL_{\text{NHC}} \tag{3.82}$$

where ‘‘NHC-baro’’ corresponds to the Nosè-Hoover chain of the barostat, and NHC corresponds to the NHC of the particles,

$$iL_1 = \sum_{i=1}^N \left[\frac{\mathbf{p}_i}{m_i} + \frac{p_\epsilon}{W} \mathbf{r}_i \right] \cdot \frac{\partial}{\partial \mathbf{r}_i} \tag{3.83}$$

$$iL_2 = \sum_{i=1}^N \mathbf{F}_i - \alpha \frac{p_\epsilon}{W} \mathbf{p}_i \cdot \frac{\partial}{\partial \mathbf{p}_i} \quad (3.84)$$

$$iL_{\epsilon,1} = \frac{p_\epsilon}{W} \frac{\partial}{\partial \epsilon} \quad (3.85)$$

$$iL_{\epsilon,2} = G_\epsilon \frac{\partial}{\partial p_\epsilon} \quad (3.86)$$

and where

$$G_\epsilon = 3V (\alpha P_{\text{kin}} - P_{\text{vir}} - P) \quad (3.87)$$

Using the Trotter decomposition, we get

$$\begin{aligned} \exp(iL\Delta t) &= \exp(iL_{\text{NHC-baro}}\Delta t/2) \exp(iL_{\text{NHC}}\Delta t/2) \\ &\exp(iL_{\epsilon,2}\Delta t/2) \exp(iL_2\Delta t/2) \\ &\exp(iL_{\epsilon,1}\Delta t) \exp(iL_1\Delta t) \\ &\exp(iL_2\Delta t/2) \exp(iL_{\epsilon,2}\Delta t/2) \\ &\exp(iL_{\text{NHC}}\Delta t/2) \exp(iL_{\text{NHC-baro}}\Delta t/2) + \mathcal{O}(\Delta t^3) \end{aligned} \quad (3.88)$$

The action of $\exp(iL_1\Delta t)$ comes from the solution of the differential equation $\dot{\mathbf{r}}_i = \mathbf{v}_i + v_\epsilon \mathbf{r}_i$ with $\mathbf{v}_i = \mathbf{p}_i/m_i$ and v_ϵ constant with initial condition $\mathbf{r}_i(0)$, evaluate at $t = \Delta t$. This yields the evolution

$$\mathbf{r}_i(\Delta t) = \mathbf{r}_i(0)e^{v_\epsilon\Delta t} + \Delta t \mathbf{v}_i(0)e^{v_\epsilon\Delta t/2} \frac{\sinh(v_\epsilon\Delta t/2)}{v_\epsilon\Delta t/2}. \quad (3.89)$$

The action of $\exp(iL_2\Delta t/2)$ comes from the solution of the differential equation $\dot{\mathbf{v}}_i = \frac{\mathbf{F}_i}{m_i} - \alpha v_\epsilon \mathbf{v}_i$, yielding

$$\mathbf{v}_i(\Delta t/2) = \mathbf{v}_i(0)e^{-\alpha v_\epsilon\Delta t/2} + \frac{\Delta t}{2m_i} \mathbf{F}_i(0)e^{-\alpha v_\epsilon\Delta t/4} \frac{\sinh(\alpha v_\epsilon\Delta t/4)}{\alpha v_\epsilon\Delta t/4}. \quad (3.90)$$

md-vv-avek uses the full step kinetic energies for determining the pressure with the pressure control, but the half-step-averaged kinetic energy for the temperatures, which can be written as a Trotter decomposition as

$$\begin{aligned} \exp(iL\Delta t) &= \exp(iL_{\text{NHC-baro}}\Delta t/2) \exp(iL_{\epsilon,2}\Delta t/2) \exp(iL_2\Delta t/2) \\ &\exp(iL_{\text{NHC}}\Delta t/2) \exp(iL_{\epsilon,1}\Delta t) \exp(iL_1\Delta t) \exp(iL_{\text{NHC}}\Delta t/2) \\ &\exp(iL_2\Delta t/2) \exp(iL_{\epsilon,2}\Delta t/2) \exp(iL_{\text{NHC-baro}}\Delta t/2) + \mathcal{O}(\Delta t^3) \end{aligned} \quad (3.91)$$

With constraints, the equations become significantly more complicated, in that each of these equations need to be solved iteratively for the constraint forces. Before GROMACS 5.1, these iterative constraints were solved as described in [42]. From GROMACS 5.1 onward, MTTK with constraints has been removed because of numerical stability issues with the iterations.

Infrequent evaluation of temperature and pressure coupling

Temperature and pressure control require global communication to compute the kinetic energy and virial, which can become costly if performed every step for large systems. We can rearrange the

Trotter decomposition to give alternate symplectic, reversible integrator with the coupling steps every n steps instead of every steps. These new integrators will diverge if the coupling time step is too large, as the auxiliary variable integrations will not converge. However, in most cases, long coupling times are more appropriate, as they disturb the dynamics less [35].

Standard velocity Verlet with Nosé-Hoover temperature control has a Trotter expansion

$$\begin{aligned} \exp(iL\Delta t) \approx & \exp(iL_{\text{NHC}}\Delta t/2) \exp(iL_2\Delta t/2) \\ & \exp(iL_1\Delta t) \exp(iL_2\Delta t/2) \exp(iL_{\text{NHC}}\Delta t/2). \end{aligned} \quad (3.92)$$

If the Nosé-Hoover chain is sufficiently slow with respect to the motions of the system, we can write an alternate integrator over n steps for velocity Verlet as

$$\begin{aligned} \exp(iL\Delta t) \approx & (\exp(iL_{\text{NHC}}(n\Delta t/2)) [\exp(iL_2\Delta t/2) \\ & \exp(iL_1\Delta t) \exp(iL_2\Delta t/2)]^n \exp(iL_{\text{NHC}}(n\Delta t/2))). \end{aligned} \quad (3.93)$$

For pressure control, this becomes

$$\begin{aligned} \exp(iL\Delta t) \approx & \exp(iL_{\text{NHC-baro}}(n\Delta t/2)) \exp(iL_{\text{NHC}}(n\Delta t/2)) \\ & \exp(iL_{\epsilon,2}(n\Delta t/2)) [\exp(iL_2\Delta t/2) \\ & \exp(iL_{\epsilon,1}\Delta t) \exp(iL_1\Delta t) \\ & \exp(iL_2\Delta t/2)]^n \exp(iL_{\epsilon,2}(n\Delta t/2)) \\ & \exp(iL_{\text{NHC}}(n\Delta t/2)) \exp(iL_{\text{NHC-baro}}(n\Delta t/2)), \end{aligned} \quad (3.94)$$

where the box volume integration occurs every step, but the auxiliary variable integrations happen every n steps.

3.4.10 The complete update algorithm

The complete algorithm for the update of velocities and coordinates is given using leap-frog in Fig. 3.8. The SHAKE algorithm of step 4 is explained below.

GROMACS has a provision to “freeze” (prevent motion of) selected particles, which must be defined as a “freeze group.” This is implemented using a *freeze factor* \mathbf{f}_g , which is a vector, and differs for each freeze group (see sec. 3.3). This vector contains only zero (freeze) or one (don’t freeze). When we take this freeze factor and the external acceleration \mathbf{a}_h into account the update algorithm for the velocities becomes

$$\mathbf{v}(t + \frac{\Delta t}{2}) = \mathbf{f}_g * \lambda * \left[\mathbf{v}(t - \frac{\Delta t}{2}) + \frac{\mathbf{F}(t)}{m} \Delta t + \mathbf{a}_h \Delta t \right], \quad (3.95)$$

where g and h are group indices which differ per atom.

3.4.11 Output step

The most important output of the MD run is the *trajectory file*, which contains particle coordinates and (optionally) velocities at regular intervals. The trajectory file contains frames that could

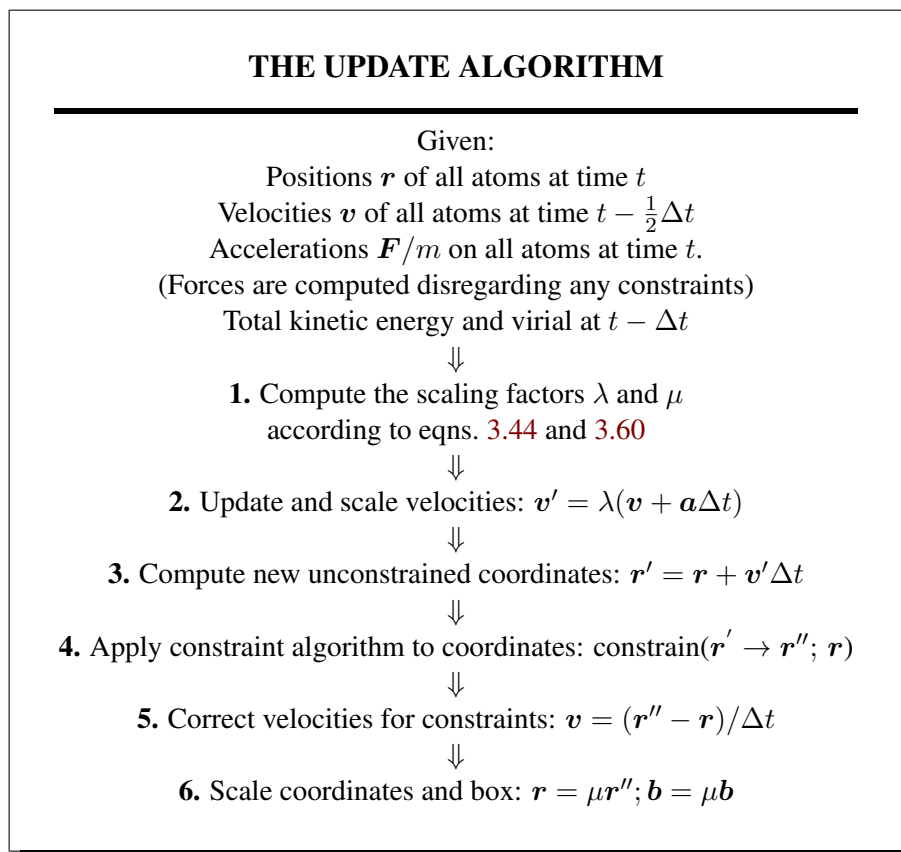


Figure 3.8: The MD update algorithm with the leap-frog integrator

include positions, velocities and/or forces, as well as information about the dimensions of the simulation volume, integration step, integration time, etc. The interpretation of the time varies with the integrator chosen, as described above. For Velocity Verlet integrators, velocities labeled at time t are for that time. For other integrators (e.g. leap-frog, stochastic dynamics), the velocities labeled at time t are for time $t - \frac{1}{2}\Delta t$.

Since the trajectory files are lengthy, one should not save every step! To retain all information it suffices to write a frame every 15 steps, since at least 30 steps are made per period of the highest frequency in the system, and Shannon's sampling theorem states that two samples per period of the highest frequency in a band-limited signal contain all available information. But that still gives very long files! So, if the highest frequencies are not of interest, 10 or 20 samples per ps may suffice. Be aware of the distortion of high-frequency motions by the *stroboscopic effect*, called *aliasing*: higher frequencies are mirrored with respect to the sampling frequency and appear as lower frequencies.

GROMACS can also write reduced-precision coordinates for a subset of the simulation system to a special compressed trajectory file format. All the other tools can read and write this format. See the User Guide for details on how to set up your `.mdp` file to have `mdrun` use this feature.

3.5 Shell molecular dynamics

GROMACS can simulate polarizability using the shell model of Dick and Overhauser [43]. In such models a shell particle representing the electronic degrees of freedom is attached to a nucleus by a spring. The potential energy is minimized with respect to the shell position at every step of the simulation (see below). Successful applications of shell models in GROMACS have been published for N_2 [44] and water [45].

3.5.1 Optimization of the shell positions

The force \mathbf{F}_S on a shell particle S can be decomposed into two components

$$\mathbf{F}_S = \mathbf{F}_{bond} + \mathbf{F}_{nb} \quad (3.96)$$

where \mathbf{F}_{bond} denotes the component representing the polarization energy, usually represented by a harmonic potential and \mathbf{F}_{nb} is the sum of Coulomb and van der Waals interactions. If we assume that \mathbf{F}_{nb} is almost constant we can analytically derive the optimal position of the shell, i.e. where $\mathbf{F}_S = 0$. If we have the shell S connected to atom A we have

$$\mathbf{F}_{bond} = k_b (\mathbf{x}_S - \mathbf{x}_A). \quad (3.97)$$

In an iterative solver, we have positions $\mathbf{x}_S(n)$ where n is the iteration count. We now have at iteration n

$$\mathbf{F}_{nb} = \mathbf{F}_S - k_b (\mathbf{x}_S(n) - \mathbf{x}_A) \quad (3.98)$$

and the optimal position for the shells $\mathbf{x}_S(n+1)$ thus follows from

$$\mathbf{F}_S - k_b (\mathbf{x}_S(n) - \mathbf{x}_A) + k_b (\mathbf{x}_S(n+1) - \mathbf{x}_A) = 0 \quad (3.99)$$

if we write

$$\Delta \mathbf{x}_S = \mathbf{x}_S(n+1) - \mathbf{x}_S(n) \quad (3.100)$$

we finally obtain

$$\Delta \mathbf{x}_S = \mathbf{F}_S/k_b \quad (3.101)$$

which then yields the algorithm to compute the next trial in the optimization of shell positions

$$\mathbf{x}_S(n+1) = \mathbf{x}_S(n) + \mathbf{F}_S/k_b. \quad (3.102)$$

3.6 Constraint algorithms

Constraints can be imposed in GROMACS using LINCS (default) or the traditional SHAKE method.

3.6.1 SHAKE

The SHAKE [46] algorithm changes a set of unconstrained coordinates \mathbf{r}' to a set of coordinates \mathbf{r}'' that fulfill a list of distance constraints, using a set \mathbf{r} reference, as

$$\text{SHAKE}(\mathbf{r}' \rightarrow \mathbf{r}''; \mathbf{r}) \quad (3.103)$$

This action is consistent with solving a set of Lagrange multipliers in the constrained equations of motion. SHAKE needs a *relative tolerance*; it will continue until all constraints are satisfied within that relative tolerance. An error message is given if SHAKE cannot reset the coordinates because the deviation is too large, or if a given number of iterations is surpassed.

Assume the equations of motion must fulfill K holonomic constraints, expressed as

$$\sigma_k(\mathbf{r}_1 \dots \mathbf{r}_N) = 0; \quad k = 1 \dots K. \quad (3.104)$$

For example, $(\mathbf{r}_1 - \mathbf{r}_2)^2 - b^2 = 0$. Then the forces are defined as

$$-\frac{\partial}{\partial \mathbf{r}_i} \left(V + \sum_{k=1}^K \lambda_k \sigma_k \right), \quad (3.105)$$

where λ_k are Lagrange multipliers which must be solved to fulfill the constraint equations. The second part of this sum determines the *constraint forces* \mathbf{G}_i , defined by

$$\mathbf{G}_i = - \sum_{k=1}^K \lambda_k \frac{\partial \sigma_k}{\partial \mathbf{r}_i} \quad (3.106)$$

The displacement due to the constraint forces in the leap-frog or Verlet algorithm is equal to $(\mathbf{G}_i/m_i)(\Delta t)^2$. Solving the Lagrange multipliers (and hence the displacements) requires the solution of a set of coupled equations of the second degree. These are solved iteratively by SHAKE. For the special case of rigid water molecules, that often make up more than 80% of the simulation system we have implemented the SETTLE algorithm [47] (sec. 5.6).

For velocity Verlet, an additional round of constraining must be done, to constrain the velocities of the second velocity half step, removing any component of the velocity parallel to the bond vector. This step is called RATTLE, and is covered in more detail in the original Andersen paper [48].

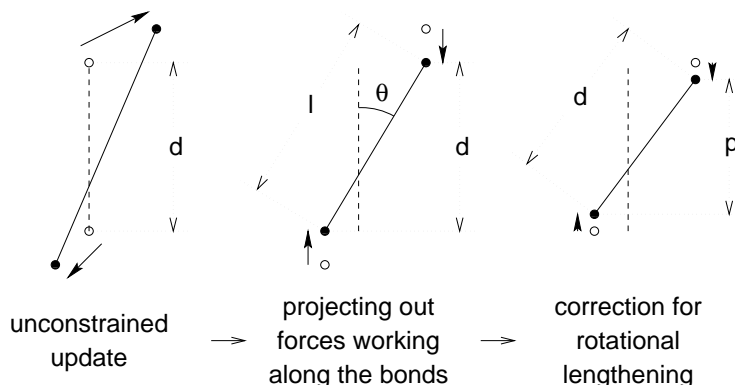


Figure 3.9: The three position updates needed for one time step. The dashed line is the old bond of length d , the solid lines are the new bonds. $l = d \cos \theta$ and $p = (2d^2 - l^2)^{\frac{1}{2}}$.

3.6.2 LINCS

The LINCS algorithm

LINCS is an algorithm that resets bonds to their correct lengths after an unconstrained update [49]. The method is non-iterative, as it always uses two steps. Although LINCS is based on matrices, no matrix-matrix multiplications are needed. The method is more stable and faster than SHAKE, but it can only be used with bond constraints and isolated angle constraints, such as the proton angle in OH. Because of its stability, LINCS is especially useful for Brownian dynamics. LINCS has two parameters, which are explained in the subsection parameters. The parallel version of LINCS, P-LINCS, is described in subsection 3.17.3.

The LINCS formulas

We consider a system of N particles, with positions given by a $3N$ vector $\mathbf{r}(t)$. For molecular dynamics the equations of motion are given by Newton's Law

$$\frac{d^2 \mathbf{r}}{dt^2} = \mathbf{M}^{-1} \mathbf{F}, \quad (3.107)$$

where \mathbf{F} is the $3N$ force vector and \mathbf{M} is a $3N \times 3N$ diagonal matrix, containing the masses of the particles. The system is constrained by K time-independent constraint equations

$$g_i(\mathbf{r}) = |\mathbf{r}_{i_1} - \mathbf{r}_{i_2}| - d_i = 0 \quad i = 1, \dots, K. \quad (3.108)$$

In a numerical integration scheme, LINCS is applied after an unconstrained update, just like SHAKE. The algorithm works in two steps (see figure Fig. 3.9). In the first step, the projections of the new bonds on the old bonds are set to zero. In the second step, a correction is applied for the lengthening of the bonds due to rotation. The numerics for the first step and the second step are very similar. A complete derivation of the algorithm can be found in [49]. Only a short description of the first step is given here.

A new notation is introduced for the gradient matrix of the constraint equations which appears on the right hand side of this equation:

$$B_{hi} = \frac{\partial g_h}{\partial r_i} \quad (3.109)$$

Notice that \mathbf{B} is a $K \times 3N$ matrix, it contains the directions of the constraints. The following equation shows how the new constrained coordinates \mathbf{r}_{n+1} are related to the unconstrained coordinates \mathbf{r}_{n+1}^{unc} by

$$\begin{aligned} \mathbf{r}_{n+1} &= (\mathbf{I} - \mathbf{T}_n \mathbf{B}_n) \mathbf{r}_{n+1}^{unc} + \mathbf{T}_n \mathbf{d} = \\ &\mathbf{r}_{n+1}^{unc} - \mathbf{M}^{-1} \mathbf{B}_n (\mathbf{B}_n \mathbf{M}^{-1} \mathbf{B}_n^T)^{-1} (\mathbf{B}_n \mathbf{r}_{n+1}^{unc} - \mathbf{d}) \end{aligned} \quad (3.110)$$

where $\mathbf{T} = \mathbf{M}^{-1} \mathbf{B}^T (\mathbf{B} \mathbf{M}^{-1} \mathbf{B}^T)^{-1}$. The derivation of this equation from eqns. 3.107 and 3.108 can be found in [49].

This first step does not set the real bond lengths to the prescribed lengths, but the projection of the new bonds onto the old directions of the bonds. To correct for the rotation of bond i , the projection of the bond, p_i , on the old direction is set to

$$p_i = \sqrt{2d_i^2 - l_i^2}, \quad (3.111)$$

where l_i is the bond length after the first projection. The corrected positions are

$$\mathbf{r}_{n+1}^* = (\mathbf{I} - \mathbf{T}_n \mathbf{B}_n) \mathbf{r}_{n+1} + \mathbf{T}_n \mathbf{p}. \quad (3.112)$$

This correction for rotational effects is actually an iterative process, but during MD only one iteration is applied. The relative constraint deviation after this procedure will be less than 0.0001 for every constraint. In energy minimization, this might not be accurate enough, so the number of iterations is equal to the order of the expansion (see below).

Half of the CPU time goes to inverting the constraint coupling matrix $\mathbf{B}_n \mathbf{M}^{-1} \mathbf{B}_n^T$, which has to be done every time step. This $K \times K$ matrix has $1/m_{i_1} + 1/m_{i_2}$ on the diagonal. The off-diagonal elements are only non-zero when two bonds are connected, then the element is $\cos \phi / m_c$, where m_c is the mass of the atom connecting the two bonds and ϕ is the angle between the bonds.

The matrix \mathbf{T} is inverted through a power expansion. A $K \times K$ matrix \mathbf{S} is introduced which is the inverse square root of the diagonal of $\mathbf{B}_n \mathbf{M}^{-1} \mathbf{B}_n^T$. This matrix is used to convert the diagonal elements of the coupling matrix to one:

$$\begin{aligned} (\mathbf{B}_n \mathbf{M}^{-1} \mathbf{B}_n^T)^{-1} &= \mathbf{S} \mathbf{S}^{-1} (\mathbf{B}_n \mathbf{M}^{-1} \mathbf{B}_n^T)^{-1} \mathbf{S}^{-1} \mathbf{S} \\ &= \mathbf{S} (\mathbf{S} \mathbf{B}_n \mathbf{M}^{-1} \mathbf{B}_n^T \mathbf{S})^{-1} \mathbf{S} = \mathbf{S} (\mathbf{I} - \mathbf{A}_n)^{-1} \mathbf{S} \end{aligned} \quad (3.113)$$

The matrix \mathbf{A}_n is symmetric and sparse and has zeros on the diagonal. Thus a simple trick can be used to calculate the inverse:

$$(\mathbf{I} - \mathbf{A}_n)^{-1} = \mathbf{I} + \mathbf{A}_n + \mathbf{A}_n^2 + \mathbf{A}_n^3 + \dots \quad (3.114)$$

This inversion method is only valid if the absolute values of all the eigenvalues of \mathbf{A}_n are smaller than one. In molecules with only bond constraints, the connectivity is so low that this will always be true, even if ring structures are present. Problems can arise in angle-constrained molecules. By

constraining angles with additional distance constraints, multiple small ring structures are introduced. This gives a high connectivity, leading to large eigenvalues. Therefore LINCS should NOT be used with coupled angle-constraints.

For molecules with all bonds constrained the eigenvalues of A are around 0.4. This means that with each additional order in the expansion eqn. 3.114 the deviations decrease by a factor 0.4. But for relatively isolated triangles of constraints the largest eigenvalue is around 0.7. Such triangles can occur when removing hydrogen angle vibrations with an additional angle constraint in alcohol groups or when constraining water molecules with LINCS, for instance with flexible constraints. The constraints in such triangles converge twice as slow as the other constraints. Therefore, starting with GROMACS 4, additional terms are added to the expansion for such triangles

$$(\mathbf{I} - \mathbf{A}_n)^{-1} \approx \mathbf{I} + \mathbf{A}_n + \dots + \mathbf{A}_n^{N_i} + (\mathbf{A}_n^* + \dots + \mathbf{A}_n^{*N_i}) \mathbf{A}_n^{N_i} \quad (3.115)$$

where N_i is the normal order of the expansion and \mathbf{A}^* only contains the elements of \mathbf{A} that couple constraints within rigid triangles, all other elements are zero. In this manner, the accuracy of angle constraints comes close to that of the other constraints, while the series of matrix vector multiplications required for determining the expansion only needs to be extended for a few constraint couplings. This procedure is described in the P-LINCS paper[50].

The LINCS Parameters

The accuracy of LINCS depends on the number of matrices used in the expansion eqn. 3.114. For MD calculations a fourth order expansion is enough. For Brownian dynamics with large time steps an eighth order expansion may be necessary. The order is a parameter in the `*.mdp` file. The implementation of LINCS is done in such a way that the algorithm will never crash. Even when it is impossible to reset the constraints LINCS will generate a conformation which fulfills the constraints as well as possible. However, LINCS will generate a warning when in one step a bond rotates over more than a predefined angle. This angle is set by the user in the `*.mdp` file.

3.7 Simulated Annealing

The well known simulated annealing (SA) protocol is supported in GROMACS, and you can even couple multiple groups of atoms separately with an arbitrary number of reference temperatures that change during the simulation. The annealing is implemented by simply changing the current reference temperature for each group in the temperature coupling, so the actual relaxation and coupling properties depends on the type of thermostat you use and how hard you are coupling it. Since we are changing the reference temperature it is important to remember that the system will NOT instantaneously reach this value - you need to allow for the inherent relaxation time in the coupling algorithm too. If you are changing the annealing reference temperature faster than the temperature relaxation you will probably end up with a crash when the difference becomes too large.

The annealing protocol is specified as a series of corresponding times and reference temperatures for each group, and you can also choose whether you only want a single sequence (after which the temperature will be coupled to the last reference value), or if the annealing should be periodic and

restart at the first reference point once the sequence is completed. You can mix and match both types of annealing and non-annealed groups in your simulation.

3.8 Stochastic Dynamics

Stochastic or velocity Langevin dynamics adds a friction and a noise term to Newton's equations of motion, as

$$m_i \frac{d^2 \mathbf{r}_i}{dt^2} = -m_i \gamma_i \frac{d\mathbf{r}_i}{dt} + \mathbf{F}_i(\mathbf{r}) + \dot{\mathbf{r}}_i, \quad (3.116)$$

where γ_i is the friction constant [1/ps] and $\dot{\mathbf{r}}_i(t)$ is a noise process with $\langle \dot{\mathbf{r}}_i(t) \dot{\mathbf{r}}_j(t+s) \rangle = 2m_i \gamma_i k_B T \delta(s) \delta_{ij}$. When $1/\gamma_i$ is large compared to the time scales present in the system, one could see stochastic dynamics as molecular dynamics with stochastic temperature-coupling. But any processes that take longer than $1/\gamma_i$, e.g. hydrodynamics, will be dampened. Since each degree of freedom is coupled independently to a heat bath, equilibration of fast modes occurs rapidly. For simulating a system in vacuum there is the additional advantage that there is no accumulation of errors for the overall translational and rotational degrees of freedom. When $1/\gamma_i$ is small compared to the time scales present in the system, the dynamics will be completely different from MD, but the sampling is still correct.

In GROMACS there is one simple and efficient implementation. Its accuracy is equivalent to the normal MD leap-frog and Velocity Verlet integrator. It is nearly identical to the common way of discretizing the Langevin equation, but the friction and velocity term are applied in an impulse fashion [51]. It can be described as:

$$\mathbf{v}' = \mathbf{v}(t - \frac{1}{2}\Delta t) + \frac{1}{m} \mathbf{F}(t) \Delta t \quad (3.117)$$

$$\Delta \mathbf{v} = -\alpha \mathbf{v}'(t + \frac{1}{2}\Delta t) + \sqrt{\frac{k_B T}{m} (1 - \alpha^2)} \mathbf{r}_i^G \quad (3.118)$$

$$\mathbf{r}(t + \Delta t) = \mathbf{r}(t) + \left(\mathbf{v}' + \frac{1}{2} \Delta \mathbf{v} \right) \Delta t \quad (3.119)$$

$$\mathbf{v}(t + \frac{1}{2}\Delta t) = \mathbf{v}' + \Delta \mathbf{v} \quad (3.120)$$

$$\alpha = 1 - e^{-\gamma \Delta t} \quad (3.121)$$

where \mathbf{r}_i^G is Gaussian distributed noise with $\mu = 0$, $\sigma = 1$. The velocity is first updated a full time step without friction and noise to get \mathbf{v}' , identical to the normal update in leap-frog. The friction and noise are then applied as an impulse at step $t + \Delta t$. The advantage of this scheme is that the velocity-dependent terms act at the full time step, which makes the correct integration of forces that depend on both coordinates and velocities, such as constraints and dissipative particle dynamics (DPD, not implemented yet), straightforward. With constraints, the coordinate update eqn. 3.119 is split into a normal leap-frog update and a $\Delta \mathbf{v}$. After both of these updates the constraints are applied to coordinates and velocities.

When using SD as a thermostat, an appropriate value for γ is e.g. 0.5 ps^{-1} , since this results in a friction that is lower than the internal friction of water, while it still provides efficient thermostating.

3.9 Brownian Dynamics

In the limit of high friction, stochastic dynamics reduces to Brownian dynamics, also called position Langevin dynamics. This applies to over-damped systems, *i.e.* systems in which the inertia effects are negligible. The equation is

$$\frac{d\mathbf{r}_i}{dt} = \frac{1}{\gamma_i} \mathbf{F}_i(\mathbf{r}) + \dot{\mathbf{r}}_i \quad (3.122)$$

where γ_i is the friction coefficient [amu/ps] and $\dot{\mathbf{r}}_i(t)$ is a noise process with $\langle \dot{\mathbf{r}}_i(t) \dot{\mathbf{r}}_j(t+s) \rangle = 2\delta(s)\delta_{ij}k_B T/\gamma_i$. In GROMACS the equations are integrated with a simple, explicit scheme

$$\mathbf{r}_i(t + \Delta t) = \mathbf{r}_i(t) + \frac{\Delta t}{\gamma_i} \mathbf{F}_i(\mathbf{r}(t)) + \sqrt{2k_B T \frac{\Delta t}{\gamma_i}} \mathbf{r}_i^G, \quad (3.123)$$

where \mathbf{r}_i^G is Gaussian distributed noise with $\mu = 0$, $\sigma = 1$. The friction coefficients γ_i can be chosen the same for all particles or as $\gamma_i = m_i \gamma_i$, where the friction constants γ_i can be different for different groups of atoms. Because the system is assumed to be over-damped, large timesteps can be used. LINCS should be used for the constraints since SHAKE will not converge for large atomic displacements. BD is an option of the `mdrun` program.

3.10 Energy Minimization

Energy minimization in GROMACS can be done using steepest descent, conjugate gradients, or l-bfgs (limited-memory Broyden-Fletcher-Goldfarb-Shanno quasi-Newtonian minimizer...we prefer the abbreviation). EM is just an option of the `mdrun` program.

3.10.1 Steepest Descent

Although steepest descent is certainly not the most efficient algorithm for searching, it is robust and easy to implement.

We define the vector \mathbf{r} as the vector of all $3N$ coordinates. Initially a maximum displacement h_0 (*e.g.* 0.01 nm) must be given.

First the forces \mathbf{F} and potential energy are calculated. New positions are calculated by

$$\mathbf{r}_{n+1} = \mathbf{r}_n + \frac{\mathbf{F}_n}{\max(|\mathbf{F}_n|)} h_n, \quad (3.124)$$

where h_n is the maximum displacement and \mathbf{F}_n is the force, or the negative gradient of the potential V . The notation $\max(|\mathbf{F}_n|)$ means the largest scalar force on any atom. The forces and energy are again computed for the new positions

If ($V_{n+1} < V_n$) the new positions are accepted and $h_{n+1} = 1.2h_n$.

If ($V_{n+1} \geq V_n$) the new positions are rejected and $h_n = 0.2h_n$.

The algorithm stops when either a user-specified number of force evaluations has been performed (*e.g.* 100), or when the maximum of the absolute values of the force (gradient) components is

smaller than a specified value ϵ . Since force truncation produces some noise in the energy evaluation, the stopping criterion should not be made too tight to avoid endless iterations. A reasonable value for ϵ can be estimated from the root mean square force f a harmonic oscillator would exhibit at a temperature T . This value is

$$f = 2\pi\nu\sqrt{2mkT}, \quad (3.125)$$

where ν is the oscillator frequency, m the (reduced) mass, and k Boltzmann's constant. For a weak oscillator with a wave number of 100 cm^{-1} and a mass of 10 atomic units, at a temperature of 1 K, $f = 7.7 \text{ kJ mol}^{-1} \text{ nm}^{-1}$. A value for ϵ between 1 and 10 is acceptable.

3.10.2 Conjugate Gradient

Conjugate gradient is slower than steepest descent in the early stages of the minimization, but becomes more efficient closer to the energy minimum. The parameters and stop criterion are the same as for steepest descent. In GROMACS conjugate gradient can not be used with constraints, including the SETTLE algorithm for water [47], as this has not been implemented. If water is present it must be of a flexible model, which can be specified in the `*.mdp` file by `define = -DFLEXIBLE`.

This is not really a restriction, since the accuracy of conjugate gradient is only required for minimization prior to a normal-mode analysis, which cannot be performed with constraints. For most other purposes steepest descent is efficient enough.

3.10.3 L-BFGS

The original BFGS algorithm works by successively creating better approximations of the inverse Hessian matrix, and moving the system to the currently estimated minimum. The memory requirements for this are proportional to the square of the number of particles, so it is not practical for large systems like biomolecules. Instead, we use the L-BFGS algorithm of Nocedal [52, 53], which approximates the inverse Hessian by a fixed number of corrections from previous steps. This sliding-window technique is almost as efficient as the original method, but the memory requirements are much lower - proportional to the number of particles multiplied with the correction steps. In practice we have found it to converge faster than conjugate gradients, but due to the correction steps it is not yet parallelized. It is also noteworthy that switched or shifted interactions usually improve the convergence, since sharp cut-offs mean the potential function at the current coordinates is slightly different from the previous steps used to build the inverse Hessian approximation.

3.11 Normal-Mode Analysis

Normal-mode analysis [54, 55, 56] can be performed using GROMACS, by diagonalization of the mass-weighted Hessian H :

$$R^T M^{-1/2} H M^{-1/2} R = \text{diag}(\lambda_1, \dots, \lambda_{3N}) \quad (3.126)$$

$$\lambda_i = (2\pi\omega_i)^2 \quad (3.127)$$

where M contains the atomic masses, R is a matrix that contains the eigenvectors as columns, λ_i are the eigenvalues and ω_i are the corresponding frequencies.

First the Hessian matrix, which is a $3N \times 3N$ matrix where N is the number of atoms, needs to be calculated:

$$H_{ij} = \frac{\partial^2 V}{\partial x_i \partial x_j} \quad (3.128)$$

where x_i and x_j denote the atomic x, y or z coordinates. In practice, this equation is not used, but the Hessian is calculated numerically from the force as:

$$H_{ij} = -\frac{f_i(\mathbf{x} + h\mathbf{e}_j) - f_i(\mathbf{x} - h\mathbf{e}_j)}{2h} \quad (3.129)$$

$$f_i = -\frac{\partial V}{\partial x_i} \quad (3.130)$$

where \mathbf{e}_j is the unit vector in direction j . It should be noted that for a usual normal-mode calculation, it is necessary to completely minimize the energy prior to computation of the Hessian. The tolerance required depends on the type of system, but a rough indication is $0.001 \text{ kJ mol}^{-1}$. Minimization should be done with conjugate gradients or L-BFGS in double precision.

A number of GROMACS programs are involved in these calculations. First, the energy should be minimized using `mdrun`. Then, `mdrun` computes the Hessian. **Note** that for generating the run input file, one should use the minimized conformation from the full precision trajectory file, as the structure file is not accurate enough. `gmx nmeig` does the diagonalization and the sorting of the normal modes according to their frequencies. Both `mdrun` and `gmx nmeig` should be run in double precision. The normal modes can be analyzed with the program `gmx ana eig`. Ensembles of structures at any temperature and for any subset of normal modes can be generated with `gmx nmens`. An overview of normal-mode analysis and the related principal component analysis (see sec. 8.11) can be found in [57].

3.12 Free energy calculations

3.12.1 Slow-growth methods

Free energy calculations can be performed in GROMACS using a number of methods, including “slow-growth.” An example problem might be calculating the difference in free energy of binding of an inhibitor **I** to an enzyme **E** and to a mutated enzyme **E'**. It is not feasible with computer simulations to perform a docking calculation for such a large complex, or even releasing the inhibitor from the enzyme in a reasonable amount of computer time with reasonable accuracy. However, if we consider the free energy cycle in Fig. 3.10A we can write:

$$\Delta G_1 - \Delta G_2 = \Delta G_3 - \Delta G_4 \quad (3.131)$$

If we are interested in the left-hand term we can equally well compute the right-hand term.

If we want to compute the difference in free energy of binding of two inhibitors **I** and **I'** to an enzyme **E** (Fig. 3.10B) we can again use eqn. 3.131 to compute the desired property.

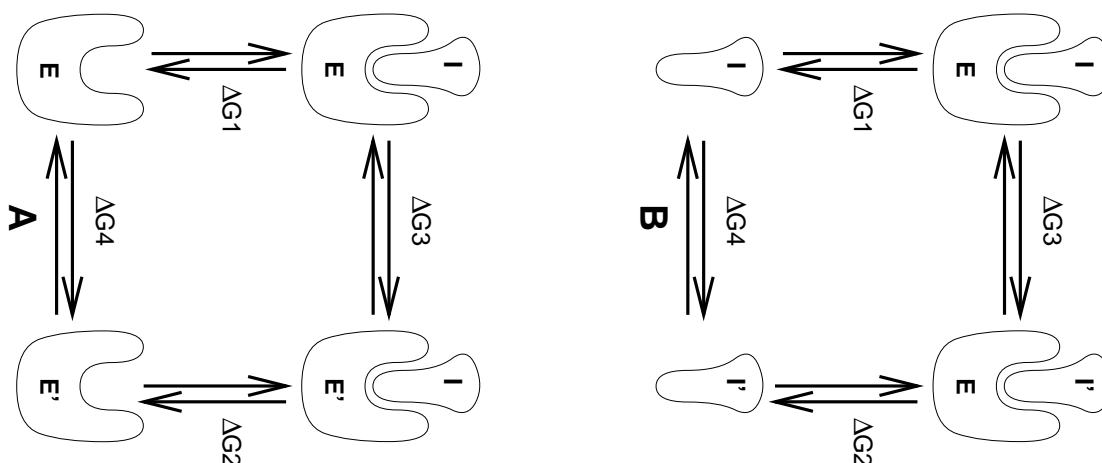


Figure 3.10: Free energy cycles. **A:** to calculate ΔG_{12} , the free energy difference between the binding of inhibitor **I** to enzymes **E** respectively **E'**. **B:** to calculate ΔG_{12} , the free energy difference for binding of inhibitors **I** respectively **I'** to enzyme **E**.

Free energy differences between two molecular species can be calculated in GROMACS using the “slow-growth” method. Such free energy differences between different molecular species are physically meaningless, but they can be used to obtain meaningful quantities employing a thermodynamic cycle. The method requires a simulation during which the Hamiltonian of the system changes slowly from that describing one system (A) to that describing the other system (B). The change must be so slow that the system remains in equilibrium during the process; if that requirement is fulfilled, the change is reversible and a slow-growth simulation from B to A will yield the same results (but with a different sign) as a slow-growth simulation from A to B. This is a useful check, but the user should be aware of the danger that equality of forward and backward growth results does not guarantee correctness of the results.

The required modification of the Hamiltonian H is realized by making H a function of a *coupling parameter* λ : $H = H(p, q; \lambda)$ in such a way that $\lambda = 0$ describes system A and $\lambda = 1$ describes system B:

$$H(p, q; 0) = H^A(p, q); \quad H(p, q; 1) = H^B(p, q). \quad (3.132)$$

In GROMACS, the functional form of the λ -dependence is different for the various force-field contributions and is described in section sec. 4.5.

The Helmholtz free energy A is related to the partition function Q of an N, V, T ensemble, which is assumed to be the equilibrium ensemble generated by a MD simulation at constant volume and temperature. The generally more useful Gibbs free energy G is related to the partition function Δ of an N, p, T ensemble, which is assumed to be the equilibrium ensemble generated by a MD simulation at constant pressure and temperature:

$$A(\lambda) = -k_B T \ln Q \quad (3.133)$$

$$Q = c \int \int \exp[-\beta H(p, q; \lambda)] dp dq \quad (3.134)$$

$$G(\lambda) = -k_B T \ln \Delta \quad (3.135)$$

$$\Delta = c \int \int \int \exp[-\beta H(p, q; \lambda) - \beta pV] dp dq dV \quad (3.136)$$

$$G = A + pV, \quad (3.137)$$

where $\beta = 1/(k_B T)$ and $c = (N!h^{3N})^{-1}$. These integrals over phase space cannot be evaluated from a simulation, but it is possible to evaluate the derivative with respect to λ as an ensemble average:

$$\frac{dA}{d\lambda} = \frac{\iint (\partial H / \partial \lambda) \exp[-\beta H(p, q; \lambda)] dp dq}{\iint \exp[-\beta H(p, q; \lambda)] dp dq} = \left\langle \frac{\partial H}{\partial \lambda} \right\rangle_{NVT; \lambda}, \quad (3.138)$$

with a similar relation for $dG/d\lambda$ in the N, p, T ensemble. The difference in free energy between A and B can be found by integrating the derivative over λ :

$$A^B(V, T) - A^A(V, T) = \int_0^1 \left\langle \frac{\partial H}{\partial \lambda} \right\rangle_{NVT; \lambda} d\lambda \quad (3.139)$$

$$G^B(p, T) - G^A(p, T) = \int_0^1 \left\langle \frac{\partial H}{\partial \lambda} \right\rangle_{NpT; \lambda} d\lambda. \quad (3.140)$$

If one wishes to evaluate $G^B(p, T) - G^A(p, T)$, the natural choice is a constant-pressure simulation. However, this quantity can also be obtained from a slow-growth simulation at constant volume, starting with system A at pressure p and volume V and ending with system B at pressure p_B , by applying the following small (but, in principle, exact) correction:

$$G^B(p) - G^A(p) = A^B(V) - A^A(V) - \int_p^{p_B} [V^B(p') - V] dp' \quad (3.141)$$

Here we omitted the constant T from the notation. This correction is roughly equal to $-\frac{1}{2}(p^B - p)\Delta V = (\Delta V)^2/(2\kappa V)$, where ΔV is the volume change at p and κ is the isothermal compressibility. This is usually small; for example, the growth of a water molecule from nothing in a bath of 1000 water molecules at constant volume would produce an additional pressure of as much as 22 bar, but a correction to the Helmholtz free energy of just -1 kJ mol^{-1} .

In Cartesian coordinates, the kinetic energy term in the Hamiltonian depends only on the momenta, and can be separately integrated and, in fact, removed from the equations. When masses do not change, there is no contribution from the kinetic energy at all; otherwise the integrated contribution to the free energy is $-\frac{3}{2}k_B T \ln(m^B/m^A)$. **Note** that this is only true in the absence of constraints.

3.12.2 Thermodynamic integration

GROMACS offers the possibility to integrate eq. 3.139 or eq. 3.140 in one simulation over the full range from A to B. However, if the change is large and insufficient sampling can be expected, the user may prefer to determine the value of $\langle dG/d\lambda \rangle$ accurately at a number of well-chosen intermediate values of λ . This can easily be done by setting the stepsize `delta_lambda` to zero. Each simulation can be equilibrated first, and a proper error estimate can be made for each value of $dG/d\lambda$ from the fluctuation of $\partial H/\partial \lambda$. The total free energy change is then determined afterward by an appropriate numerical integration procedure.

GROMACS now also supports the use of Bennett's Acceptance Ratio [58] for calculating values of ΔG for transformations from state A to state B using the program `gmx bar`. The same data can also be used to calculate free energies using MBAR [59], though the analysis currently requires external tools from the external `pymbar` package, at <https://SimTK.org/home/pymbar>.

The λ -dependence for the force-field contributions is described in detail in section sec. 4.5.

3.13 Replica exchange

Replica exchange molecular dynamics (REMD) is a method that can be used to speed up the sampling of any type of simulation, especially if conformations are separated by relatively high energy barriers. It involves simulating multiple replicas of the same system at different temperatures and randomly exchanging the complete state of two replicas at regular intervals with the probability:

$$P(1 \leftrightarrow 2) = \min \left(1, \exp \left[\left(\frac{1}{k_B T_1} - \frac{1}{k_B T_2} \right) (U_1 - U_2) \right] \right) \quad (3.142)$$

where T_1 and T_2 are the reference temperatures and U_1 and U_2 are the instantaneous potential energies of replicas 1 and 2 respectively. After exchange the velocities are scaled by $(T_1/T_2)^{\pm 0.5}$ and a neighbor search is performed the next step. This combines the fast sampling and frequent barrier-crossing of the highest temperature with correct Boltzmann sampling at all the different temperatures [60, 61]. We only attempt exchanges for neighboring temperatures as the probability decreases very rapidly with the temperature difference. One should not attempt exchanges for all possible pairs in one step. If, for instance, replicas 1 and 2 would exchange, the chance of exchange for replicas 2 and 3 not only depends on the energies of replicas 2 and 3, but also on the energy of replica 1. In GROMACS this is solved by attempting exchange for all “odd” pairs on “odd” attempts and for all “even” pairs on “even” attempts. If we have four replicas: 0, 1, 2 and 3, ordered in temperature and we attempt exchange every 1000 steps, pairs 0-1 and 2-3 will be tried at steps 1000, 3000 etc. and pair 1-2 at steps 2000, 4000 etc.

How should one choose the temperatures? The energy difference can be written as:

$$U_1 - U_2 = N_{df} \frac{c}{2} k_B (T_1 - T_2) \quad (3.143)$$

where N_{df} is the total number of degrees of freedom of one replica and c is 1 for harmonic potentials and around 2 for protein/water systems. If $T_2 = (1 + \epsilon)T_1$ the probability becomes:

$$P(1 \leftrightarrow 2) = \exp \left(-\frac{\epsilon^2 c N_{df}}{2(1 + \epsilon)} \right) \approx \exp \left(-\epsilon^2 \frac{c}{2} N_{df} \right) \quad (3.144)$$

Thus for a probability of $e^{-2} \approx 0.135$ one obtains $\epsilon \approx 2/\sqrt{c N_{df}}$. With all bonds constrained one has $N_{df} \approx 2 N_{atoms}$ and thus for $c = 2$ one should choose ϵ as $1/\sqrt{N_{atoms}}$. However there is one problem when using pressure coupling. The density at higher temperatures will decrease, leading to higher energy [62], which should be taken into account. The GROMACS website features a so-called “REMD calculator,” that lets you type in the temperature range and the number of atoms, and based on that proposes a set of temperatures.

An extension to the REMD for the isobaric-isothermal ensemble was proposed by Okabe *et al.* [63]. In this work the exchange probability is modified to:

$$P(1 \leftrightarrow 2) = \min \left(1, \exp \left[\left(\frac{1}{k_B T_1} - \frac{1}{k_B T_2} \right) (U_1 - U_2) + \left(\frac{P_1}{k_B T_1} - \frac{P_2}{k_B T_2} \right) (V_1 - V_2) \right] \right) \quad (3.145)$$

where P_1 and P_2 are the respective reference pressures and V_1 and V_2 are the respective instantaneous volumes in the simulations. In most cases the differences in volume are so small that the second term is negligible. It only plays a role when the difference between P_1 and P_2 is large or in phase transitions.

Hamiltonian replica exchange is also supported in GROMACS. In Hamiltonian replica exchange, each replica has a different Hamiltonian, defined by the free energy pathway specified for the simulation. The exchange probability to maintain the correct ensemble probabilities is:

$$P(1 \leftrightarrow 2) = \min \left(1, \exp \left[\left(\frac{1}{k_B T} - \frac{1}{k_B T} \right) ((U_1(x_2) - U_1(x_1)) + (U_2(x_1) - U_2(x_2))) \right] \right) \quad (3.146)$$

The separate Hamiltonians are defined by the free energy functionality of GROMACS, with swaps made between the different values of λ defined in the `mdp` file.

Hamiltonian and temperature replica exchange can also be performed simultaneously, using the acceptance criteria:

$$P(1 \leftrightarrow 2) = \min \left(1, \exp \left[\left(\frac{1}{k_B T} - \frac{1}{k_B T} \right) \left(\frac{U_1(x_2) - U_1(x_1)}{k_B T_1} + \frac{U_2(x_1) - U_2(x_2)}{k_B T_2} \right) \right] \right) \quad (3.147)$$

Gibbs sampling replica exchange has also been implemented in GROMACS [64]. In Gibbs sampling replica exchange, all possible pairs are tested for exchange, allowing swaps between replicas that are not neighbors.

Gibbs sampling replica exchange requires no additional potential energy calculations. However there is an additional communication cost in Gibbs sampling replica exchange, as for some permutations, more than one round of swaps must take place. In some cases, this extra communication cost might affect the efficiency.

All replica exchange variants are options of the `mdrun` program. It will only work when MPI is installed, due to the inherent parallelism in the algorithm. For efficiency each replica can run on a separate rank. See the manual page of `mdrun` on how to use these multinode features.

3.14 Essential Dynamics sampling

The results from Essential Dynamics (see sec. 8.11) of a protein can be used to guide MD simulations. The idea is that from an initial MD simulation (or from other sources) a definition of the collective fluctuations with largest amplitude is obtained. The position along one or more of these collective modes can be constrained in a (second) MD simulation in a number of ways for several purposes. For example, the position along a certain mode may be kept fixed to monitor the average force (free-energy gradient) on that coordinate in that position. Another application is to enhance sampling efficiency with respect to usual MD [65, 66]. In this case, the system is encouraged to sample its available configuration space more systematically than in a diffusion-like path that proteins usually take.

Another possibility to enhance sampling is flooding. Here a flooding potential is added to certain (collective) degrees of freedom to expel the system out of a region of phase space [67].

The procedure for essential dynamics sampling or flooding is as follows. First, the eigenvectors and eigenvalues need to be determined using covariance analysis (`gmx covar`) or normal-mode analysis (`gmx nmeig`). Then, this information is fed into `make_edi`, which has many options for selecting vectors and setting parameters, see `gmx make_edi -h`. The generated `edi` input file is then passed to `mdrun`.

3.15 Expanded Ensemble

In an expanded ensemble simulation [68], both the coordinates and the thermodynamic ensemble are treated as configuration variables that can be sampled over. The probability of any given state can be written as:

$$P(\vec{x}, k) \propto \exp(-\beta_k U_k + g_k), \quad (3.148)$$

where $\beta_k = \frac{1}{k_B T_k}$ is the β corresponding to the k th thermodynamic state, and g_k is a user-specified weight factor corresponding to the k th state. This space is therefore a *mixed, generalized, or expanded* ensemble which samples from multiple thermodynamic ensembles simultaneously. g_k is chosen to give a specific weighting of each subensemble in the expanded ensemble, and can either be fixed, or determined by an iterative procedure. The set of g_k is frequently chosen to give each thermodynamic ensemble equal probability, in which case g_k is equal to the free energy in non-dimensional units, but they can be set to arbitrary values as desired. Several different algorithms can be used to equilibrate these weights, described in the mdp option listings.

In GROMACS, this space is sampled by alternating sampling in the k and \vec{x} directions. Sampling in the \vec{x} direction is done by standard molecular dynamics sampling; sampling between the different thermodynamics states is done by Monte Carlo, with several different Monte Carlo moves supported. The k states can be defined by different temperatures, or choices of the free energy λ variable, or both. Expanded ensemble simulations thus represent a serialization of the replica exchange formalism, allowing a single simulation to explore many thermodynamic states.

3.16 Parallelization

The CPU time required for a simulation can be reduced by running the simulation in parallel over more than one core. Ideally, one would want to have linear scaling: running on N cores makes the simulation N times faster. In practice this can only be achieved for a small number of cores. The scaling will depend a lot on the algorithms used. Also, different algorithms can have different restrictions on the interaction ranges between atoms.

3.17 Domain decomposition

Since most interactions in molecular simulations are local, domain decomposition is a natural way to decompose the system. In domain decomposition, a spatial domain is assigned to each rank, which will then integrate the equations of motion for the particles that currently reside in its local domain. With domain decomposition, there are two choices that have to be made: the division of the unit cell into domains and the assignment of the forces to domains. Most molecular simulation packages use the half-shell method for assigning the forces. But there are two methods that always require less communication: the eighth shell [69] and the midpoint [70] method. GROMACS currently uses the eighth shell method, but for certain systems or hardware architectures it might be advantageous to use the midpoint method. Therefore, we might implement the midpoint method in the future. Most of the details of the domain decomposition can be found in the GROMACS 4 paper [5].

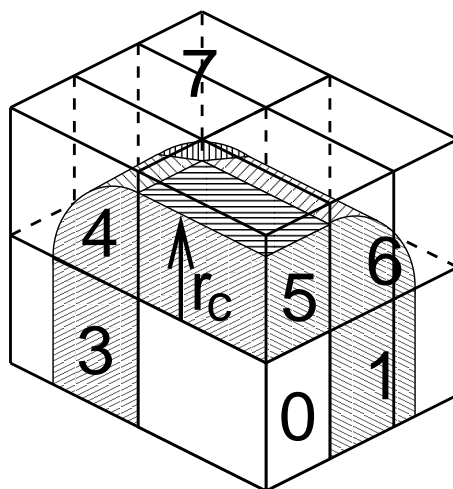


Figure 3.11: A non-staggered domain decomposition grid of $3 \times 2 \times 2$ cells. Coordinates in zones 1 to 7 are communicated to the corner cell that has its home particles in zone 0. r_c is the cut-off radius.

3.17.1 Coordinate and force communication

In the most general case of a triclinic unit cell, the space is divided with a 1-, 2-, or 3-D grid in parallelepipeds that we call domain decomposition cells. Each cell is assigned to a particle-particle rank. The system is partitioned over the ranks at the beginning of each MD step in which neighbor searching is performed. Since the neighbor searching is based on charge groups, charge groups are also the units for the domain decomposition. Charge groups are assigned to the cell where their center of geometry resides. Before the forces can be calculated, the coordinates from some neighboring cells need to be communicated, and after the forces are calculated, the forces need to be communicated in the other direction. The communication and force assignment is based on zones that can cover one or multiple cells. An example of a zone setup is shown in Fig. 3.11.

The coordinates are communicated by moving data along the “negative” direction in x , y or z to the next neighbor. This can be done in one or multiple pulses. In Fig. 3.11 two pulses in x are required, then one in y and then one in z . The forces are communicated by reversing this procedure. See the GROMACS 4 paper [5] for details on determining which non-bonded and bonded forces should be calculated on which rank.

3.17.2 Dynamic load balancing

When different ranks have a different computational load (load imbalance), all ranks will have to wait for the one that takes the most time. One would like to avoid such a situation. Load imbalance can occur due to four reasons:

- inhomogeneous particle distribution
- inhomogeneous interaction cost distribution (charged/uncharged, water/non-water due to GROMACS water innerloops)

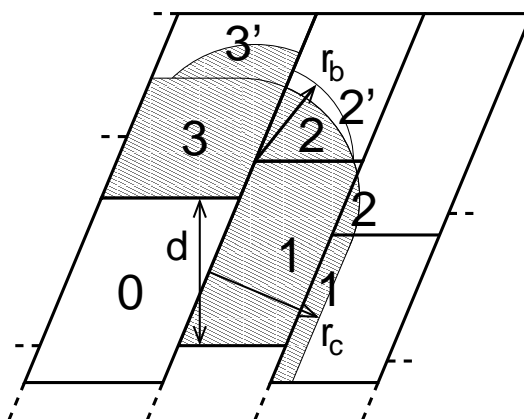


Figure 3.12: The zones to communicate to the rank of zone 0, see the text for details. r_c and r_b are the non-bonded and bonded cut-off radii respectively, d is an example of a distance between following, staggered boundaries of cells.

- statistical fluctuation (only with small particle numbers)
- differences in communication time, due to network topology and/or other jobs on the machine interfering with our communication

So we need a dynamic load balancing algorithm where the volume of each domain decomposition cell can be adjusted *independently*. To achieve this, the 2- or 3-D domain decomposition grids need to be staggered. Fig. 3.12 shows the most general case in 2-D. Due to the staggering, one might require two distance checks for deciding if a charge group needs to be communicated: a non-bonded distance and a bonded distance check.

By default, `mdrun` automatically turns on the dynamic load balancing during a simulation when the total performance loss due to the force calculation imbalance is 2% or more. **Note** that the reported force load imbalance numbers might be higher, since the force calculation is only part of work that needs to be done during an integration step. The load imbalance is reported in the log file at log output steps and when the `-v` option is used also on screen. The average load imbalance and the total performance loss due to load imbalance are reported at the end of the log file.

There is one important parameter for the dynamic load balancing, which is the minimum allowed scaling. By default, each dimension of the domain decomposition cell can scale down by at least a factor of 0.8. For 3-D domain decomposition this allows cells to change their volume by about a factor of 0.5, which should allow for compensation of a load imbalance of 100%. The minimum allowed scaling can be changed with the `-dds` option of `mdrun`.

The load imbalance is measured by timing a single region of the MD step on each MPI rank. This region can not include MPI communication, as timing of MPI calls does not allow separating wait due to imbalance from actual communication. The domain volumes are then scaled, with under-relaxation, inversely proportional with the measured time. This procedure will decrease the load imbalance when the change in load in the measured region correlates with the change in domain volume and the load outside the measured region does not depend strongly on the domain volume. In CPU-only simulations, the load is measured between the coordinate and the force communication. In simulations with non-bonded work on GPUs, we overlap communication and work on the

interaction	range	option	default
non-bonded	$r_c = \max(r_{\text{list}}, r_{\text{vdW}}, r_{\text{Coul}})$	mdp file	
two-body bonded	$\max(r_{\text{mb}}, r_c)$	mdrun -rdd	starting conf. + 10%
multi-body bonded	r_{mb}	mdrun -rdd	starting conf. + 10%
constraints	r_{con}	mdrun -rcon	est. from bond lengths
virtual sites	r_{con}	mdrun -rcon	0

Table 3.2: The interaction ranges with domain decomposition.

CPU with calculation on the GPU. Therefore we measure from the last communication before the force calculation to when the CPU or GPU is finished, whichever is last. When not using PME ranks, we subtract the time in PME from the CPU time, as this includes MPI calls and the PME load is independent of domain size. This generally works well, unless the non-bonded load is low and there is imbalance in the bonded interactions. Then two issues can arise. Dynamic load balancing can increase the imbalance in update and constraints and with PME the coordinate and force redistribution time can go up significantly. Although dynamic load balancing can significantly improve performance in cases where there is imbalance in the bonded interactions on the CPU, there are many situations in which some domains continue decreasing in size and the load imbalance increases and/or PME coordinate and force redistribution cost increases significantly. As of version 2016.1, `mdrun` disables the dynamic load balancing when measurement indicates that it deteriorates performance. This means that in most cases the user will get good performance with the default, automated dynamic load balancing setting.

3.17.3 Constraints in parallel

Since with domain decomposition parts of molecules can reside on different ranks, bond constraints can cross cell boundaries. Therefore a parallel constraint algorithm is required. GRO-MACS uses the P-LINCS algorithm [50], which is the parallel version of the LINCS algorithm [49] (see 3.6.2). The P-LINCS procedure is illustrated in Fig. 3.13. When molecules cross the cell boundaries, atoms in such molecules up to (`lincs_order + 1`) bonds away are communicated over the cell boundaries. Then, the normal LINCS algorithm can be applied to the local bonds plus the communicated ones. After this procedure, the local bonds are correctly constrained, even though the extra communicated ones are not. One coordinate communication step is required for the initial LINCS step and one for each iteration. Forces do not need to be communicated.

3.17.4 Interaction ranges

Domain decomposition takes advantage of the locality of interactions. This means that there will be limitations on the range of interactions. By default, `mdrun` tries to find the optimal balance between interaction range and efficiency. But it can happen that a simulation stops with an error message about missing interactions, or that a simulation might run slightly faster with shorter interaction ranges. A list of interaction ranges and their default values is given in Table 3.2.

In most cases the defaults of `mdrun` should not cause the simulation to stop with an error message of missing interactions. The range for the bonded interactions is determined from the distance be-

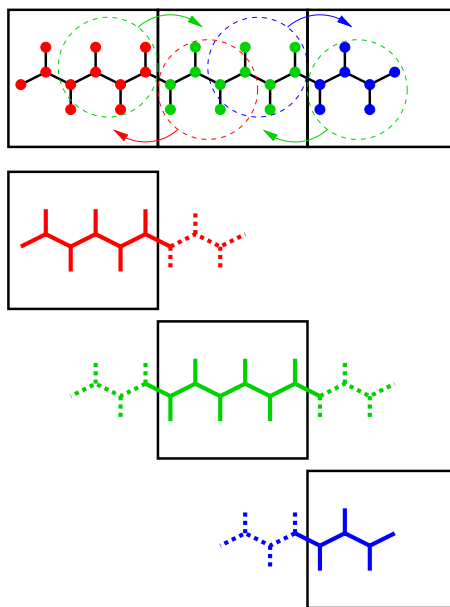


Figure 3.13: Example of the parallel setup of P-LINCS with one molecule split over three domain decomposition cells, using a matrix expansion order of 3. The top part shows which atom coordinates need to be communicated to which cells. The bottom parts show the local constraints (solid) and the non-local constraints (dashed) for each of the three cells.

tween bonded charge-groups in the starting configuration, with 10% added for headroom. For the constraints, the value of r_{con} is determined by taking the maximum distance that $(\text{lincs_order} + 1)$ bonds can cover when they all connect at angles of 120 degrees. The actual constraint communication is not limited by r_{con} , but by the minimum cell size L_C , which has the following lower limit:

$$L_C \geq \max(r_{\text{mb}}, r_{\text{con}}) \quad (3.149)$$

Without dynamic load balancing the system is actually allowed to scale beyond this limit when pressure scaling is used. **Note** that for triclinic boxes, L_C is not simply the box diagonal component divided by the number of cells in that direction, rather it is the shortest distance between the triclinic cells borders. For rhombic dodecahedra this is a factor of $\sqrt{3/2}$ shorter along x and y .

When $r_{\text{mb}} > r_c$, `mdrun` employs a smart algorithm to reduce the communication. Simply communicating all charge groups within r_{mb} would increase the amount of communication enormously. Therefore only charge-groups that are connected by bonded interactions to charge groups which are not locally present are communicated. This leads to little extra communication, but also to a slightly increased cost for the domain decomposition setup. In some cases, *e.g.* coarse-grained simulations with a very short cut-off, one might want to set r_{mb} by hand to reduce this cost.

3.17.5 Multiple-Program, Multiple-Data PME parallelization

Electrostatics interactions are long-range, therefore special algorithms are used to avoid summation over many atom pairs. In GROMACS this is usually PME (sec. 4.8.2). Since with PME all

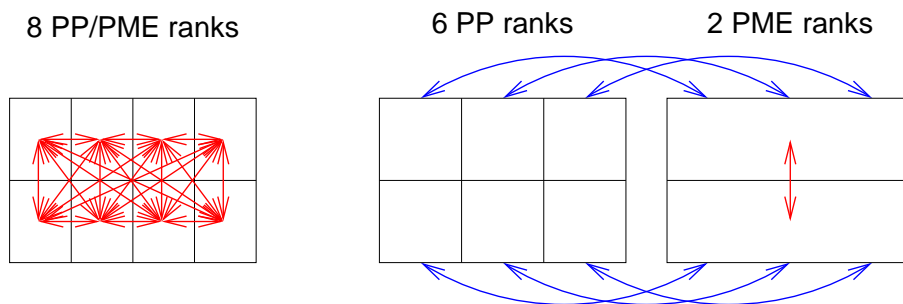


Figure 3.14: Example of 8 ranks without (left) and with (right) MPMD. The PME communication (red arrows) is much higher on the left than on the right. For MPMD additional PP - PME coordinate and force communication (blue arrows) is required, but the total communication complexity is lower.

particles interact with each other, global communication is required. This will usually be the limiting factor for scaling with domain decomposition. To reduce the effect of this problem, we have come up with a Multiple-Program, Multiple-Data approach [5]. Here, some ranks are selected to do only the PME mesh calculation, while the other ranks, called particle-particle (PP) ranks, do all the rest of the work. For rectangular boxes the optimal PP to PME rank ratio is usually 3:1, for rhombic dodecahedra usually 2:1. When the number of PME ranks is reduced by a factor of 4, the number of communication calls is reduced by about a factor of 16. Or put differently, we can now scale to 4 times more ranks. In addition, for modern 4 or 8 core machines in a network, the effective network bandwidth for PME is quadrupled, since only a quarter of the cores will be using the network connection on each machine during the PME calculations.

`mdrun` will by default interleave the PP and PME ranks. If the ranks are not number consecutively inside the machines, one might want to use `mdrun -ddorder pp_pme`. For machines with a real 3-D torus and proper communication software that assigns the ranks accordingly one should use `mdrun -ddorder cartesian`.

To optimize the performance one should usually set up the cut-offs and the PME grid such that the PME load is 25 to 33% of the total calculation load. `grompp` will print an estimate for this load at the end and also `mdrun` calculates the same estimate to determine the optimal number of PME ranks to use. For high parallelization it might be worthwhile to optimize the PME load with the `mdp` settings and/or the number of PME ranks with the `-npme` option of `mdrun`. For changing the electrostatics settings it is useful to know the accuracy of the electrostatics remains nearly constant when the Coulomb cut-off and the PME grid spacing are scaled by the same factor. **Note** that it is usually better to overestimate than to underestimate the number of PME ranks, since the number of PME ranks is smaller than the number of PP ranks, which leads to less total waiting time.

The PME domain decomposition can be 1-D or 2-D along the x and/or y axis. 2-D decomposition is also known as pencil decomposition because of the shape of the domains at high parallelization. 1-D decomposition along the y axis can only be used when the PP decomposition has only 1 domain along x . 2-D PME decomposition has to have the number of domains along x equal to the number of the PP decomposition. `mdrun` automatically chooses 1-D or 2-D PME decomposition (when possible with the total given number of ranks), based on the minimum amount of commu-

nication for the coordinate redistribution in PME plus the communication for the grid overlap and transposes. To avoid superfluous communication of coordinates and forces between the PP and PME ranks, the number of DD cells in the x direction should ideally be the same or a multiple of the number of PME ranks. By default, `mdrun` takes care of this issue.

3.17.6 Domain decomposition flow chart

In Fig. 3.15 a flow chart is shown for domain decomposition with all possible communication for different algorithms. For simpler simulations, the same flow chart applies, without the algorithms and communication for the algorithms that are not used.

3.18 Implicit solvation

Implicit solvent models provide an efficient way of representing the electrostatic effects of solvent molecules, while saving a large piece of the computations involved in an accurate, aqueous description of the surrounding water in molecular dynamics simulations. Implicit solvation models offer several advantages compared with explicit solvation, including eliminating the need for the equilibration of water around the solute, and the absence of viscosity, which allows the protein to more quickly explore conformational space.

Implicit solvent calculations in GROMACS can be done using the generalized Born-formalism, and the Still [71], HCT [72], and OBC [73] models are available for calculating the Born radii.

Here, the free energy G_{solv} of solvation is the sum of three terms, a solvent-solvent cavity term (G_{cav}), a solute-solvent van der Waals term (G_{vdw}), and finally a solvent-solute electrostatics polarization term (G_{pol}).

The sum of G_{cav} and G_{vdw} corresponds to the (non-polar) free energy of solvation for a molecule from which all charges have been removed, and is commonly called G_{np} , calculated from the total solvent accessible surface area multiplied with a surface tension. The total expression for the solvation free energy then becomes:

$$G_{\text{solv}} = G_{\text{np}} + G_{\text{pol}} \quad (3.150)$$

Under the generalized Born model, G_{pol} is calculated from the generalized Born equation [71]:

$$G_{\text{pol}} = \left(1 - \frac{1}{\epsilon}\right) \sum_{i=1}^n \sum_{j>i}^n \frac{q_i q_j}{\sqrt{r_{ij}^2 + b_i b_j} \exp\left(\frac{-r_{ij}^2}{4b_i b_j}\right)} \quad (3.151)$$

In GROMACS, we have introduced the substitution [74]:

$$c_i = \frac{1}{\sqrt{b_i}} \quad (3.152)$$

which makes it possible to introduce a cheap transformation to a new variable x when evaluating each interaction, such that:

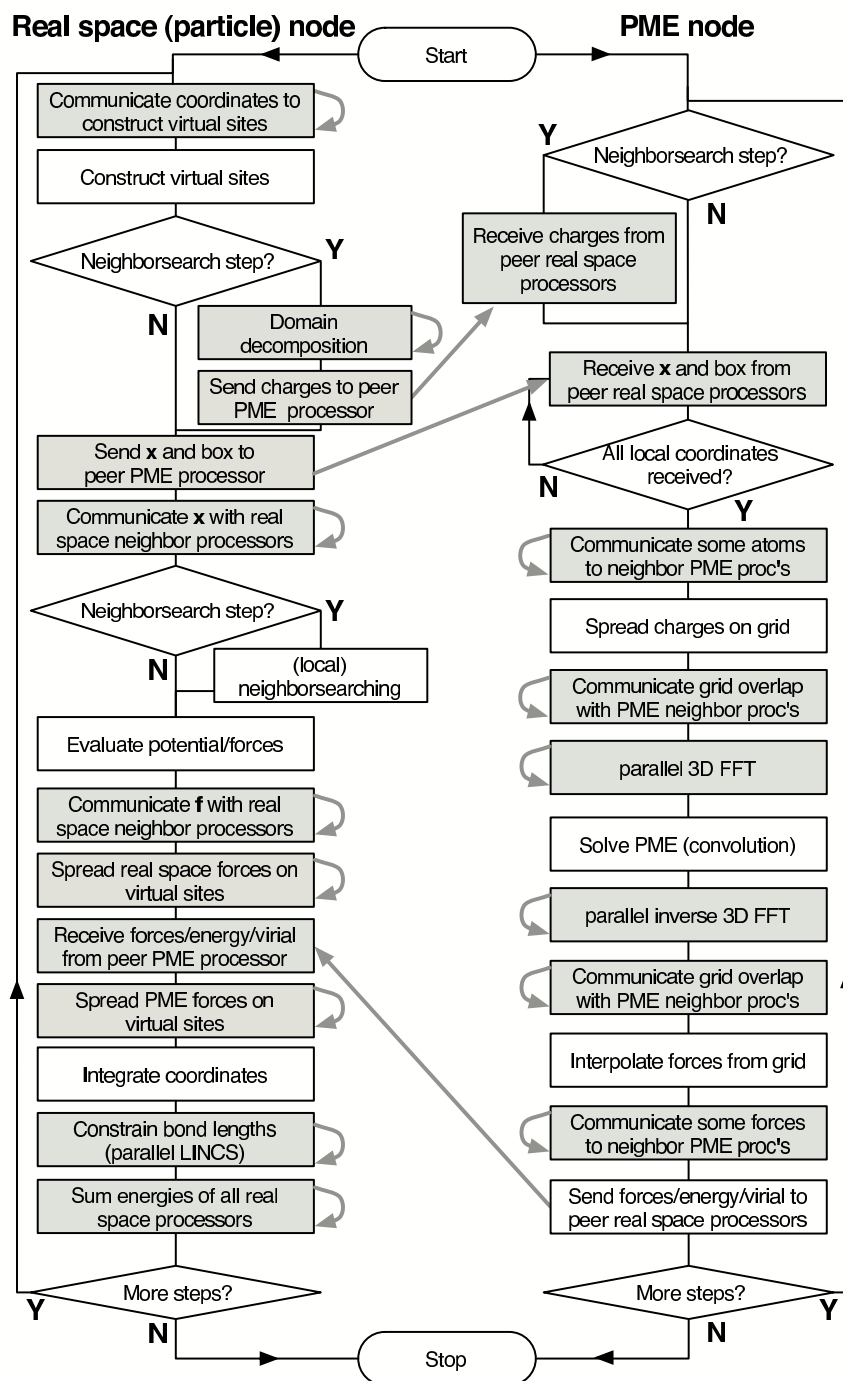


Figure 3.15: Flow chart showing the algorithms and communication (arrows) for a standard MD simulation with virtual sites, constraints and separate PME-mesh ranks.

$$x = \frac{r_{ij}}{\sqrt{b_i b_j}} = r_{ij} c_i c_j \quad (3.153)$$

In the end, the full re-formulation of 3.151 becomes:

$$G_{\text{pol}} = \left(1 - \frac{1}{\epsilon}\right) \sum_{i=1}^n \sum_{j>i}^n \frac{q_i q_j}{\sqrt{b_i b_j}} \xi(x) = \left(1 - \frac{1}{\epsilon}\right) \sum_{i=1}^n q_i c_i \sum_{j>i}^n q_j c_j \xi(x) \quad (3.154)$$

The non-polar part (G_{np}) of Equation 3.150 is calculated directly from the Born radius of each atom using a simple ACE type approximation by Schaefer *et al.* [75], including a simple loop over all atoms. This requires only one extra solvation parameter, independent of atom type, but differing slightly between the three Born radii models.

Chapter 4

Interaction function and force fields

To accommodate the potential functions used in some popular force fields (see 4.10), GROMACS offers a choice of functions, both for non-bonded interaction and for dihedral interactions. They are described in the appropriate subsections.

The potential functions can be subdivided into three parts

1. *Non-bonded*: Lennard-Jones or Buckingham, and Coulomb or modified Coulomb. The non-bonded interactions are computed on the basis of a neighbor list (a list of non-bonded atoms within a certain radius), in which exclusions are already removed.
2. *Bonded*: covalent bond-stretching, angle-bending, improper dihedrals, and proper dihedrals. These are computed on the basis of fixed lists.
3. *Restraints*: position restraints, angle restraints, distance restraints, orientation restraints and dihedral restraints, all based on fixed lists.
4. *Applied Forces*: externally applied forces, see chapter 6.

4.1 Non-bonded interactions

Non-bonded interactions in GROMACS are pair-additive:

$$V(\mathbf{r}_1, \dots, \mathbf{r}_N) = \sum_{i < j} V_{ij}(r_{ij}); \quad (4.1)$$

$$\mathbf{F}_i = - \sum_j \frac{dV_{ij}(r_{ij})}{dr_{ij}} \frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.2)$$

Since the potential only depends on the scalar distance, interactions will be centro-symmetric, i.e. the vectorial partial force on particle i from the pairwise interaction $V_{ij}(r_{ij})$ has the opposite

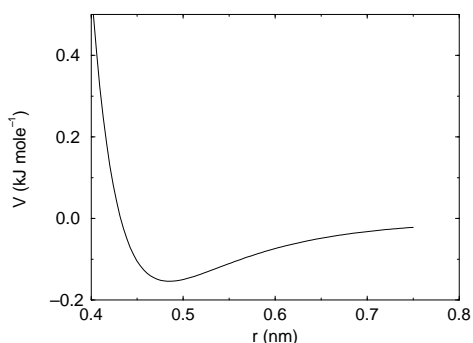


Figure 4.1: The Lennard-Jones interaction.

direction of the partial force on particle j . For efficiency reasons, interactions are calculated by loops over interactions and updating both partial forces rather than summing one complete nonbonded force at a time. The non-bonded interactions contain a repulsion term, a dispersion term, and a Coulomb term. The repulsion and dispersion term are combined in either the Lennard-Jones (or 6-12 interaction), or the Buckingham (or exp-6 potential). In addition, (partially) charged atoms act through the Coulomb term.

4.1.1 The Lennard-Jones interaction

The Lennard-Jones potential V_{LJ} between two atoms equals:

$$V_{LJ}(r_{ij}) = \frac{C_{ij}^{(12)}}{r_{ij}^{12}} - \frac{C_{ij}^{(6)}}{r_{ij}^6} \quad (4.3)$$

See also Fig. 4.1 The parameters $C_{ij}^{(12)}$ and $C_{ij}^{(6)}$ depend on pairs of *atom types*; consequently they are taken from a matrix of LJ-parameters. In the Verlet cut-off scheme, the potential is shifted by a constant such that it is zero at the cut-off distance.

The force derived from this potential is:

$$\mathbf{F}_i(\mathbf{r}_{ij}) = \left(12 \frac{C_{ij}^{(12)}}{r_{ij}^{13}} - 6 \frac{C_{ij}^{(6)}}{r_{ij}^7} \right) \frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.4)$$

The LJ potential may also be written in the following form:

$$V_{LJ}(r_{ij}) = 4\epsilon_{ij} \left(\left(\frac{\sigma_{ij}}{r_{ij}} \right)^{12} - \left(\frac{\sigma_{ij}}{r_{ij}} \right)^6 \right) \quad (4.5)$$

In constructing the parameter matrix for the non-bonded LJ-parameters, two types of combination rules can be used within GROMACS, only geometric averages (type 1 in the input section of the

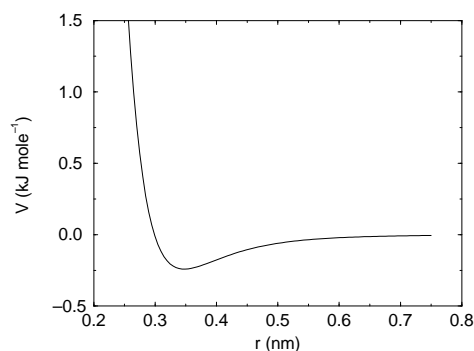


Figure 4.2: The Buckingham interaction.

force-field file):

$$\begin{aligned} C_{ij}^{(6)} &= \left(C_{ii}^{(6)} C_{jj}^{(6)} \right)^{1/2} \\ C_{ij}^{(12)} &= \left(C_{ii}^{(12)} C_{jj}^{(12)} \right)^{1/2} \end{aligned} \quad (4.6)$$

or, alternatively the Lorentz-Berthelot rules can be used. An arithmetic average is used to calculate σ_{ij} , while a geometric average is used to calculate ϵ_{ij} (type 2):

$$\begin{aligned} \sigma_{ij} &= \frac{1}{2}(\sigma_{ii} + \sigma_{jj}) \\ \epsilon_{ij} &= (\epsilon_{ii} \epsilon_{jj})^{1/2} \end{aligned} \quad (4.7)$$

finally an geometric average for both parameters can be used (type 3):

$$\begin{aligned} \sigma_{ij} &= (\sigma_{ii} \sigma_{jj})^{1/2} \\ \epsilon_{ij} &= (\epsilon_{ii} \epsilon_{jj})^{1/2} \end{aligned} \quad (4.8)$$

This last rule is used by the OPLS force field.

4.1.2 Buckingham potential

The Buckingham potential has a more flexible and realistic repulsion term than the Lennard-Jones interaction, but is also more expensive to compute. The potential form is:

$$V_{bh}(r_{ij}) = A_{ij} \exp(-B_{ij}r_{ij}) - \frac{C_{ij}}{r_{ij}^6} \quad (4.9)$$

See also Fig. 4.2. The force derived from this is:

$$\mathbf{F}_i(r_{ij}) = \left[A_{ij} B_{ij} \exp(-B_{ij}r_{ij}) - 6 \frac{C_{ij}}{r_{ij}^7} \right] \frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.10)$$

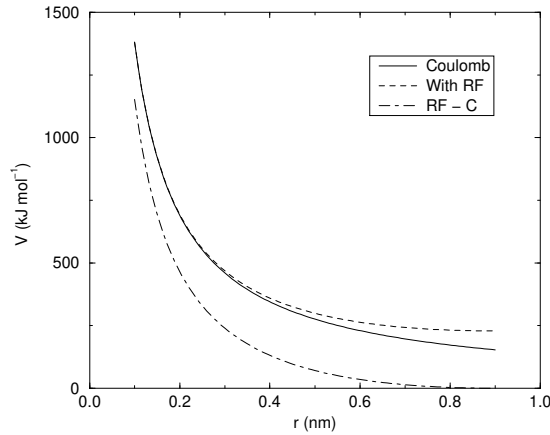


Figure 4.3: The Coulomb interaction (for particles with equal signed charge) with and without reaction field. In the latter case ϵ_r was 1, ϵ_{rf} was 78, and r_c was 0.9 nm. The dot-dashed line is the same as the dashed line, except for a constant.

4.1.3 Coulomb interaction

The Coulomb interaction between two charge particles is given by:

$$V_c(r_{ij}) = f \frac{q_i q_j}{\epsilon_r r_{ij}} \quad (4.11)$$

See also Fig. 4.3, where $f = \frac{1}{4\pi\epsilon_0} = 138.935\,458$ (see chapter 2)

The force derived from this potential is:

$$\mathbf{F}_i(\mathbf{r}_{ij}) = f \frac{q_i q_j}{\epsilon_r r_{ij}^2} \frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.12)$$

A plain Coulomb interaction should only be used without cut-off or when all pairs fall within the cut-off, since there is an abrupt, large change in the force at the cut-off. In case you do want to use a cut-off, the potential can be shifted by a constant to make the potential the integral of the force. With the group cut-off scheme, this shift is only applied to non-excluded pairs. With the Verlet cut-off scheme, the shift is also applied to excluded pairs and self interactions, which makes the potential equivalent to a reaction field with $\epsilon_{rf} = 1$ (see below).

In GROMACS the relative dielectric constant ϵ_r may be set in the in the input for `grompp`.

4.1.4 Coulomb interaction with reaction field

The Coulomb interaction can be modified for homogeneous systems by assuming a constant dielectric environment beyond the cut-off r_c with a dielectric constant of ϵ_{rf} . The interaction then reads:

$$V_{crf} = f \frac{q_i q_j}{\epsilon_r r_{ij}} \left[1 + \frac{\epsilon_{rf} - \epsilon_r}{2\epsilon_{rf} + \epsilon_r} \frac{r_{ij}^3}{r_c^3} \right] - f \frac{q_i q_j}{\epsilon_r r_c} \frac{3\epsilon_{rf}}{2\epsilon_{rf} + \epsilon_r} \quad (4.13)$$

in which the constant expression on the right makes the potential zero at the cut-off r_c . For charged cut-off spheres this corresponds to neutralization with a homogeneous background charge. We can rewrite eqn. 4.13 for simplicity as

$$V_{crf} = f \frac{q_i q_j}{\epsilon_r} \left[\frac{1}{r_{ij}} + k_{rf} r_{ij}^2 - c_{rf} \right] \quad (4.14)$$

with

$$k_{rf} = \frac{1}{r_c^3} \frac{\epsilon_{rf} - \epsilon_r}{(2\epsilon_{rf} + \epsilon_r)} \quad (4.15)$$

$$c_{rf} = \frac{1}{r_c} + k_{rf} r_c^2 = \frac{1}{r_c} \frac{3\epsilon_{rf}}{(2\epsilon_{rf} + \epsilon_r)} \quad (4.16)$$

For large ϵ_{rf} the k_{rf} goes to $r_c^{-3}/2$, while for $\epsilon_{rf} = \epsilon_r$ the correction vanishes. In Fig. 4.3 the modified interaction is plotted, and it is clear that the derivative with respect to r_{ij} (= -force) goes to zero at the cut-off distance. The force derived from this potential reads:

$$\mathbf{F}_i(\mathbf{r}_{ij}) = f \frac{q_i q_j}{\epsilon_r} \left[\frac{1}{r_{ij}^2} - 2k_{rf} r_{ij} \right] \frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.17)$$

The reaction-field correction should also be applied to all excluded atoms pairs, including self pairs, in which case the normal Coulomb term in eqns. 4.13 and 4.17 is absent.

Tironi *et al.* have introduced a generalized reaction field in which the dielectric continuum beyond the cut-off r_c also has an ionic strength I [76]. In this case we can rewrite the constants k_{rf} and c_{rf} using the inverse Debye screening length κ :

$$\kappa^2 = \frac{2I F^2}{\epsilon_0 \epsilon_{rf} RT} = \frac{F^2}{\epsilon_0 \epsilon_{rf} RT} \sum_{i=1}^K c_i z_i^2 \quad (4.18)$$

$$k_{rf} = \frac{1}{r_c^3} \frac{(\epsilon_{rf} - \epsilon_r)(1 + \kappa r_c) + \frac{1}{2}\epsilon_{rf}(\kappa r_c)^2}{(2\epsilon_{rf} + \epsilon_r)(1 + \kappa r_c) + \epsilon_{rf}(\kappa r_c)^2} \quad (4.19)$$

$$c_{rf} = \frac{1}{r_c} \frac{3\epsilon_{rf}(1 + \kappa r_c + \frac{1}{2}(\kappa r_c)^2)}{(2\epsilon_{rf} + \epsilon_r)(1 + \kappa r_c) + \epsilon_{rf}(\kappa r_c)^2} \quad (4.20)$$

where F is Faraday's constant, R is the ideal gas constant, T the absolute temperature, c_i the molar concentration for species i and z_i the charge number of species i where we have K different species. In the limit of zero ionic strength ($\kappa = 0$) eqns. 4.19 and 4.20 reduce to the simple forms of eqns. 4.15 and 4.16 respectively.

4.1.5 Modified non-bonded interactions

In GROMACS, the non-bonded potentials can be modified by a shift function, also called a force-switch function, since it switches the force to zero at the cut-off. The purpose of this is to replace the truncated forces by forces that are continuous and have continuous derivatives at the cut-off radius. With such forces the time integration produces smaller errors. But note that for Lennard-Jones interactions these errors are usually smaller than other errors, such as integration errors at

the repulsive part of the potential. For Coulomb interactions we advise against using a shifted potential and for use of a reaction field or a proper long-range method such as PME.

There is *no* fundamental difference between a switch function (which multiplies the potential with a function) and a shift function (which adds a function to the force or potential) [77]. The switch function is a special case of the shift function, which we apply to the *force function* $F(r)$, related to the electrostatic or van der Waals force acting on particle i by particle j as:

$$\mathbf{F}_i = c F(r_{ij}) \frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.21)$$

For pure Coulomb or Lennard-Jones interactions $F(r) = F_\alpha(r) = \alpha r^{-(\alpha+1)}$. The switched force $F_s(r)$ can generally be written as:

$$\begin{aligned} F_s(r) &= F_\alpha(r) & r < r_1 \\ F_s(r) &= F_\alpha(r) + S(r) & r_1 \leq r < r_c \\ F_s(r) &= 0 & r_c \leq r \end{aligned} \quad (4.22)$$

When $r_1 = 0$ this is a traditional shift function, otherwise it acts as a switch function. The corresponding shifted potential function then reads:

$$V_s(r) = \int_r^\infty F_s(x) dx \quad (4.23)$$

The GROMACS **force switch** function $S_F(r)$ should be smooth at the boundaries, therefore the following boundary conditions are imposed on the switch function:

$$\begin{aligned} S_F(r_1) &= 0 \\ S'_F(r_1) &= 0 \\ S_F(r_c) &= -F_\alpha(r_c) \\ S'_F(r_c) &= -F'_\alpha(r_c) \end{aligned} \quad (4.24)$$

A 3^{rd} degree polynomial of the form

$$S_F(r) = A(r - r_1)^2 + B(r - r_1)^3 \quad (4.25)$$

fulfills these requirements. The constants A and B are given by the boundary condition at r_c :

$$\begin{aligned} A &= -\alpha \frac{(\alpha + 4)r_c - (\alpha + 1)r_1}{r_c^{\alpha+2} (r_c - r_1)^2} \\ B &= \alpha \frac{(\alpha + 3)r_c - (\alpha + 1)r_1}{r_c^{\alpha+2} (r_c - r_1)^3} \end{aligned} \quad (4.26)$$

Thus the total force function is:

$$F_s(r) = \frac{\alpha}{r^{\alpha+1}} + A(r - r_1)^2 + B(r - r_1)^3 \quad (4.27)$$

and the potential function reads:

$$V_s(r) = \frac{1}{r^\alpha} - \frac{A}{3}(r - r_1)^3 - \frac{B}{4}(r - r_1)^4 - C \quad (4.28)$$

where

$$C = \frac{1}{r_c^\alpha} - \frac{A}{3}(r_c - r_1)^3 - \frac{B}{4}(r_c - r_1)^4 \quad (4.29)$$

The GROMACS **potential-switch** function $S_V(r)$ scales the potential between r_1 and r_c , and has similar boundary conditions, intended to produce smoothly-varying potential and forces:

$$\begin{aligned} S_V(r_1) &= 1 \\ S'_V(r_1) &= 0 \\ S''_V(r_1) &= 0 \\ S_V(r_c) &= 0 \\ S'_V(r_c) &= 0 \\ S''_V(r_c) &= 0 \end{aligned} \quad (4.30)$$

The fifth-degree polynomial that has these properties is

$$S_V(r; r_1, r_c) = \frac{1 - 10(r - r_1)^3(r_c - r_1)^2 + 15(r - r_1)^4(r_c - r_1) - 6(r - r_1)}{(r_c - r_1)^5} \quad (4.31)$$

This implementation is found in several other simulation packages,[78, 79, 80] but differs from that in CHARMM.[81] Switching the potential leads to artificially large forces in the switching region, therefore it is not recommended to switch Coulomb interactions using this function,[77] but switching Lennard-Jones interactions using this function produces acceptable results.

4.1.6 Modified short-range interactions with Ewald summation

When Ewald summation or particle-mesh Ewald is used to calculate the long-range interactions, the short-range Coulomb potential must also be modified. Here the potential is switched to (nearly) zero at the cut-off, instead of the force. In this case the short range potential is given by:

$$V(r) = f \frac{\operatorname{erfc}(\beta r_{ij})}{r_{ij}} q_i q_j, \quad (4.32)$$

where β is a parameter that determines the relative weight between the direct space sum and the reciprocal space sum and $\operatorname{erfc}(x)$ is the complementary error function. For further details on long-range electrostatics, see sec. 4.8.

4.2 Bonded interactions

Bonded interactions are based on a fixed list of atoms. They are not exclusively pair interactions, but include 3- and 4-body interactions as well. There are *bond stretching* (2-body), *bond angle* (3-body), and *dihedral angle* (4-body) interactions. A special type of dihedral interaction (called *improper dihedral*) is used to force atoms to remain in a plane or to prevent transition to a configuration of opposite chirality (a mirror image).

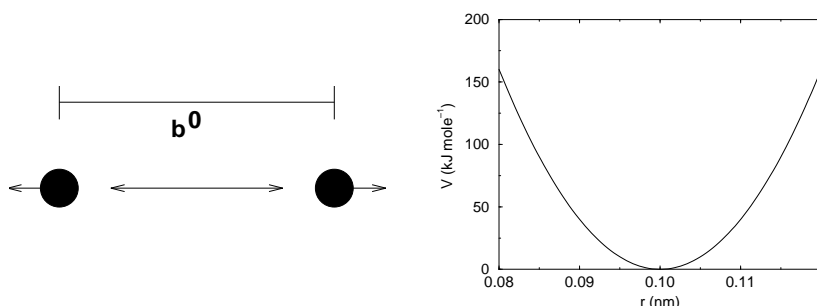


Figure 4.4: Principle of bond stretching (left), and the bond stretching potential (right).

4.2.1 Bond stretching

Harmonic potential

The bond stretching between two covalently bonded atoms i and j is represented by a harmonic potential:

$$V_b(r_{ij}) = \frac{1}{2}k_{ij}^b(r_{ij} - b_{ij})^2 \quad (4.33)$$

See also Fig. 4.4, with the force given by:

$$\mathbf{F}_i(\mathbf{r}_{ij}) = k_{ij}^b(r_{ij} - b_{ij})\frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.34)$$

Fourth power potential

In the GROMOS-96 force field [82], the covalent bond potential is, for reasons of computational efficiency, written as:

$$V_b(r_{ij}) = \frac{1}{4}k_{ij}^b(r_{ij}^2 - b_{ij}^2)^2 \quad (4.35)$$

The corresponding force is:

$$\mathbf{F}_i(\mathbf{r}_{ij}) = k_{ij}^b(r_{ij}^2 - b_{ij}^2)\mathbf{r}_{ij} \quad (4.36)$$

The force constants for this form of the potential are related to the usual harmonic force constant $k^{b,\text{harm}}$ (sec. 4.2.1) as

$$2k^b b_{ij}^2 = k^{b,\text{harm}} \quad (4.37)$$

The force constants are mostly derived from the harmonic ones used in GROMOS-87 [83]. Although this form is computationally more efficient (because no square root has to be evaluated), it is conceptually more complex. One particular disadvantage is that since the form is not harmonic, the average energy of a single bond is not equal to $\frac{1}{2}kT$ as it is for the normal harmonic potential.

4.2.2 Morse potential bond stretching

For some systems that require an anharmonic bond stretching potential, the Morse potential [84] between two atoms i and j is available in GROMACS. This potential differs from the harmonic potential in that it has an asymmetric potential well and a zero force at infinite distance. The functional form is:

$$V_{morse}(r_{ij}) = D_{ij}[1 - \exp(-\beta_{ij}(r_{ij} - b_{ij}))]^2, \quad (4.38)$$

See also Fig. 4.5, and the corresponding force is:

$$\mathbf{F}_{morse}(\mathbf{r}_{ij}) = 2D_{ij}\beta_{ij} \exp(-\beta_{ij}(r_{ij} - b_{ij})) * [1 - \exp(-\beta_{ij}(r_{ij} - b_{ij}))] \frac{\mathbf{r}_{ij}}{r_{ij}}, \quad (4.39)$$

where D_{ij} is the depth of the well in kJ/mol, β_{ij} defines the steepness of the well (in nm^{-1}), and b_{ij} is the equilibrium distance in nm. The steepness parameter β_{ij} can be expressed in terms of the reduced mass of the atoms i and j , the fundamental vibration frequency ω_{ij} and the well depth D_{ij} :

$$\beta_{ij} = \omega_{ij} \sqrt{\frac{\mu_{ij}}{2D_{ij}}} \quad (4.40)$$

and because $\omega = \sqrt{k/\mu}$, one can rewrite β_{ij} in terms of the harmonic force constant k_{ij} :

$$\beta_{ij} = \sqrt{\frac{k_{ij}}{2D_{ij}}} \quad (4.41)$$

For small deviations ($r_{ij} - b_{ij}$), one can approximate the exp-term to first-order using a Taylor expansion:

$$\exp(-x) \approx 1 - x \quad (4.42)$$

and substituting eqn. 4.41 and eqn. 4.42 in the functional form:

$$\begin{aligned} V_{morse}(r_{ij}) &= D_{ij}[1 - \exp(-\beta_{ij}(r_{ij} - b_{ij}))]^2 \\ &= D_{ij}[1 - (1 - \sqrt{\frac{k_{ij}}{2D_{ij}}}(r_{ij} - b_{ij}))]^2 \\ &= \frac{1}{2}k_{ij}(r_{ij} - b_{ij})^2 \end{aligned} \quad (4.43)$$

we recover the harmonic bond stretching potential.

4.2.3 Cubic bond stretching potential

Another anharmonic bond stretching potential that is slightly simpler than the Morse potential adds a cubic term in the distance to the simple harmonic form:

$$V_b(r_{ij}) = k_{ij}^b(r_{ij} - b_{ij})^2 + k_{ij}^b k_{ij}^{cub}(r_{ij} - b_{ij})^3 \quad (4.44)$$

A flexible water model (based on the SPC water model [85]) including a cubic bond stretching potential for the O-H bond was developed by Ferguson [86]. This model was found to yield a reasonable infrared spectrum. The Ferguson water model is available in the GROMACS library

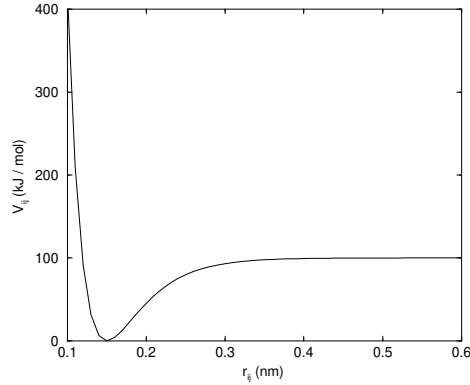


Figure 4.5: The Morse potential well, with bond length 0.15 nm.

(flexwat-ferguson.itp). It should be noted that the potential is asymmetric: overstretching leads to infinitely low energies. The integration timestep is therefore limited to 1 fs.

The force corresponding to this potential is:

$$\mathbf{F}_i(\mathbf{r}_{ij}) = 2k_{ij}^b(r_{ij} - b_{ij}) \frac{\mathbf{r}_{ij}}{r_{ij}} + 3k_{ij}^b k_{ij}^{cub} (r_{ij} - b_{ij})^2 \frac{\mathbf{r}_{ij}}{r_{ij}} \quad (4.45)$$

4.2.4 FENE bond stretching potential

In coarse-grained polymer simulations the beads are often connected by a FENE (finitely extensible nonlinear elastic) potential [87]:

$$V_{\text{FENE}}(r_{ij}) = -\frac{1}{2} k_{ij}^b b_{ij}^2 \log \left(1 - \frac{r_{ij}^2}{b_{ij}^2} \right) \quad (4.46)$$

The potential looks complicated, but the expression for the force is simpler:

$$\mathbf{F}_{\text{FENE}}(\mathbf{r}_{ij}) = -k_{ij}^b \left(1 - \frac{r_{ij}^2}{b_{ij}^2} \right)^{-1} \mathbf{r}_{ij} \quad (4.47)$$

At short distances the potential asymptotically goes to a harmonic potential with force constant k^b , while it diverges at distance b .

4.2.5 Harmonic angle potential

The bond-angle vibration between a triplet of atoms $i - j - k$ is also represented by a harmonic potential on the angle θ_{ijk}

$$V_a(\theta_{ijk}) = \frac{1}{2} k_{ijk}^\theta (\theta_{ijk} - \theta_{ijk}^0)^2 \quad (4.48)$$

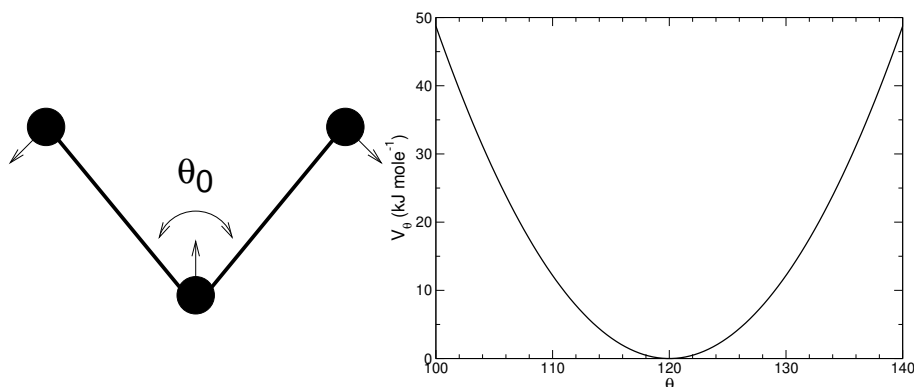


Figure 4.6: Principle of angle vibration (left) and the bond angle potential (right).

As the bond-angle vibration is represented by a harmonic potential, the form is the same as the bond stretching (Fig. 4.4).

The force equations are given by the chain rule:

$$\begin{aligned} \mathbf{F}_i &= -\frac{dV_a(\theta_{ijk})}{d\mathbf{r}_i} \\ \mathbf{F}_k &= -\frac{dV_a(\theta_{ijk})}{d\mathbf{r}_k} \quad \text{where} \quad \theta_{ijk} = \arccos \frac{\mathbf{r}_{ij} \cdot \mathbf{r}_{kj}}{r_{ij}r_{kj}} \\ \mathbf{F}_j &= -\mathbf{F}_i - \mathbf{F}_k \end{aligned} \quad (4.49)$$

The numbering i, j, k is in sequence of covalently bonded atoms. Atom j is in the middle; atoms i and k are at the ends (see Fig. 4.6). **Note** that in the input in topology files, angles are given in degrees and force constants in kJ/mol/rad^2 .

4.2.6 Cosine based angle potential

In the GROMOS-96 force field a simplified function is used to represent angle vibrations:

$$V_a(\theta_{ijk}) = \frac{1}{2} k_{ijk}^{\theta} \left(\cos(\theta_{ijk}) - \cos(\theta_{ijk}^0) \right)^2 \quad (4.50)$$

where

$$\cos(\theta_{ijk}) = \frac{\mathbf{r}_{ij} \cdot \mathbf{r}_{kj}}{r_{ij}r_{kj}} \quad (4.51)$$

The corresponding force can be derived by partial differentiation with respect to the atomic positions. The force constants in this function are related to the force constants in the harmonic form $k^{\theta, \text{harm}}$ (4.2.5) by:

$$k^{\theta} \sin^2(\theta_{ijk}^0) = k^{\theta, \text{harm}} \quad (4.52)$$

In the GROMOS-96 manual there is a much more complicated conversion formula which is temperature dependent. The formulas are equivalent at 0 K and the differences at 300 K are on the order of 0.1 to 0.2%. **Note** that in the input in topology files, angles are given in degrees and force constants in kJ/mol .

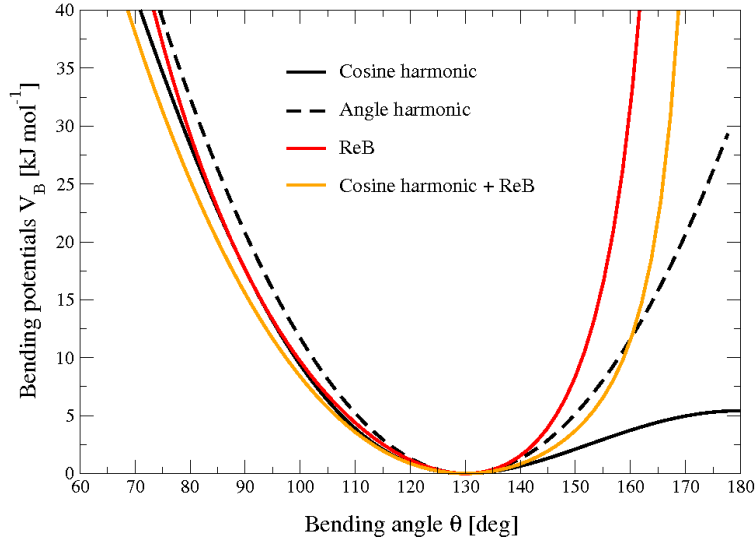


Figure 4.7: Bending angle potentials: cosine harmonic (solid black line), angle harmonic (dashed black line) and restricted bending (red) with the same bending constant $k_\theta = 85 \text{ kJ mol}^{-1}$ and equilibrium angle $\theta_0 = 130^\circ$. The orange line represents the sum of a cosine harmonic ($k = 50 \text{ kJ mol}^{-1}$) with a restricted bending ($k = 25 \text{ kJ mol}^{-1}$) potential, both with $\theta_0 = 130^\circ$.

4.2.7 Restricted bending potential

The restricted bending (ReB) potential [88] prevents the bending angle θ from reaching the 180° value. In this way, the numerical instabilities due to the calculation of the torsion angle and potential are eliminated when performing coarse-grained molecular dynamics simulations.

To systematically hinder the bending angles from reaching the 180° value, the bending potential 4.50 is divided by a $\sin^2 \theta$ factor:

$$V_{\text{ReB}}(\theta_i) = \frac{1}{2} k_\theta \frac{(\cos \theta_i - \cos \theta_0)^2}{\sin^2 \theta_i}. \quad (4.53)$$

Figure Fig. 4.7 shows the comparison between the ReB potential, 4.53, and the standard one 4.50. The wall of the ReB potential is very repulsive in the region close to 180° and, as a result, the bending angles are kept within a safe interval, far from instabilities. The power 2 of $\sin \theta_i$ in the denominator has been chosen to guarantee this behavior and allows an elegant differentiation:

$$F_{\text{ReB}}(\theta_i) = \frac{2k_\theta}{\sin^4 \theta_i} (\cos \theta_i - \cos \theta_0) (1 - \cos \theta_i \cos \theta_0) \frac{\partial \cos \theta_i}{\partial \vec{r}_k}. \quad (4.54)$$

Due to its construction, the restricted bending potential cannot be used for equilibrium θ_0 values too close to 0° or 180° (from experience, at least 10° difference is recommended). It is very important that, in the starting configuration, all the bending angles have to be in the safe interval to avoid initial instabilities. This bending potential can be used in combination with any form of torsion potential. It will always prevent three consecutive particles from becoming collinear and, as a result, any torsion potential will remain free of singularities. It can be also added to a

standard bending potential to affect the angle around 180° , but to keep its original form around the minimum (see the orange curve in Fig. 4.7).

4.2.8 Urey-Bradley potential

The Urey-Bradley bond-angle vibration between a triplet of atoms $i - j - k$ is represented by a harmonic potential on the angle θ_{ijk} and a harmonic correction term on the distance between the atoms i and k . Although this can be easily written as a simple sum of two terms, it is convenient to have it as a single entry in the topology file and in the output as a separate energy term. It is used mainly in the CHARMM force field [89]. The energy is given by:

$$V_a(\theta_{ijk}) = \frac{1}{2}k_{ijk}^\theta(\theta_{ijk} - \theta_{ijk}^0)^2 + \frac{1}{2}k_{ijk}^{UB}(r_{ik} - r_{ik}^0)^2 \quad (4.55)$$

The force equations can be deduced from sections 4.2.1 and 4.2.5.

4.2.9 Bond-Bond cross term

The bond-bond cross term for three particles i, j, k forming bonds $i - j$ and $k - j$ is given by [90]:

$$V_{rr'} = k_{rr'} (|\mathbf{r}_i - \mathbf{r}_j| - r_{1e}) (|\mathbf{r}_k - \mathbf{r}_j| - r_{2e}) \quad (4.56)$$

where $k_{rr'}$ is the force constant, and r_{1e} and r_{2e} are the equilibrium bond lengths of the $i - j$ and $k - j$ bonds respectively. The force associated with this potential on particle i is:

$$\mathbf{F}_i = -k_{rr'} (|\mathbf{r}_k - \mathbf{r}_j| - r_{2e}) \frac{\mathbf{r}_i - \mathbf{r}_j}{|\mathbf{r}_i - \mathbf{r}_j|} \quad (4.57)$$

The force on atom k can be obtained by swapping i and k in the above equation. Finally, the force on atom j follows from the fact that the sum of internal forces should be zero: $\mathbf{F}_j = -\mathbf{F}_i - \mathbf{F}_k$.

4.2.10 Bond-Angle cross term

The bond-angle cross term for three particles i, j, k forming bonds $i - j$ and $k - j$ is given by [90]:

$$V_{r\theta} = k_{r\theta} (|\mathbf{r}_i - \mathbf{r}_k| - r_{3e}) (|\mathbf{r}_i - \mathbf{r}_j| - r_{1e} + |\mathbf{r}_k - \mathbf{r}_j| - r_{2e}) \quad (4.58)$$

where $k_{r\theta}$ is the force constant, r_{3e} is the $i - k$ distance, and the other constants are the same as in Equation 4.56. The force associated with the potential on atom i is:

$$\mathbf{F}_i = -k_{r\theta} \left[(|\mathbf{r}_i - \mathbf{r}_k| - r_{3e}) \frac{\mathbf{r}_i - \mathbf{r}_j}{|\mathbf{r}_i - \mathbf{r}_j|} + (|\mathbf{r}_i - \mathbf{r}_j| - r_{1e} + |\mathbf{r}_k - \mathbf{r}_j| - r_{2e}) \frac{\mathbf{r}_i - \mathbf{r}_k}{|\mathbf{r}_i - \mathbf{r}_k|} \right] \quad (4.59)$$

4.2.11 Quartic angle potential

For special purposes there is an angle potential that uses a fourth order polynomial:

$$V_q(\theta_{ijk}) = \sum_{n=0}^5 C_n (\theta_{ijk} - \theta_{ijk}^0)^n \quad (4.60)$$

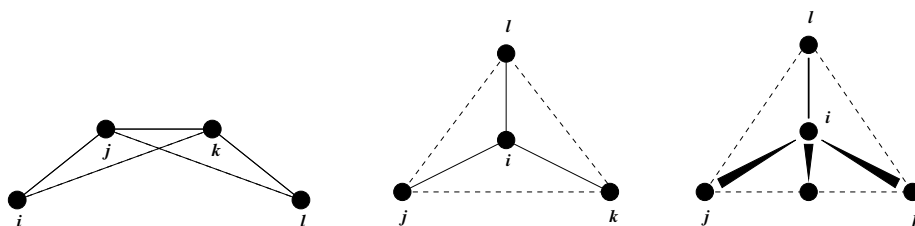


Figure 4.8: Principle of improper dihedral angles. Out of plane bending for rings (left), substituents of rings (middle), out of tetrahedral (right). The improper dihedral angle ξ is defined as the angle between planes (i,j,k) and (j,k,l) in all cases.

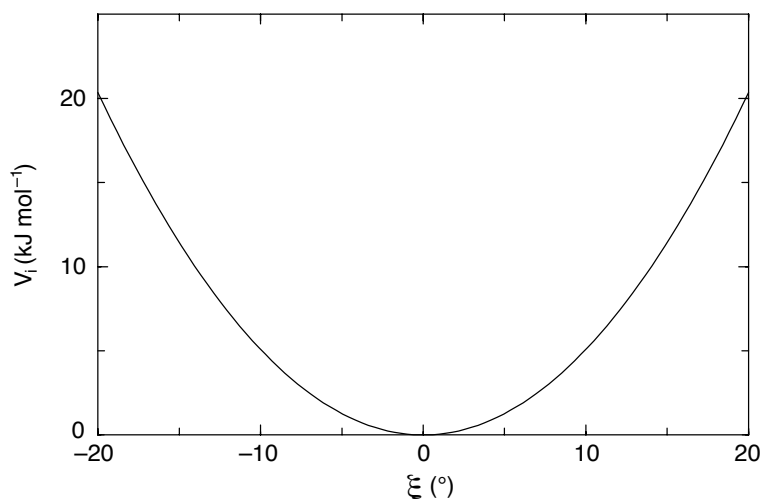


Figure 4.9: Improper dihedral potential.

4.2.12 Improper dihedrals

Improper dihedrals are meant to keep planar groups (*e.g.* aromatic rings) planar, or to prevent molecules from flipping over to their mirror images, see Fig. 4.8.

Improper dihedrals: harmonic type

The simplest improper dihedral potential is a harmonic potential; it is plotted in Fig. 4.9.

$$V_{id}(\xi_{ijkl}) = \frac{1}{2}k_{\xi}(\xi_{ijkl} - \xi_0)^2 \quad (4.61)$$

Since the potential is harmonic it is discontinuous, but since the discontinuity is chosen at 180° distance from ξ_0 this will never cause problems. **Note** that in the input in topology files, angles are given in degrees and force constants in kJ/mol/rad².

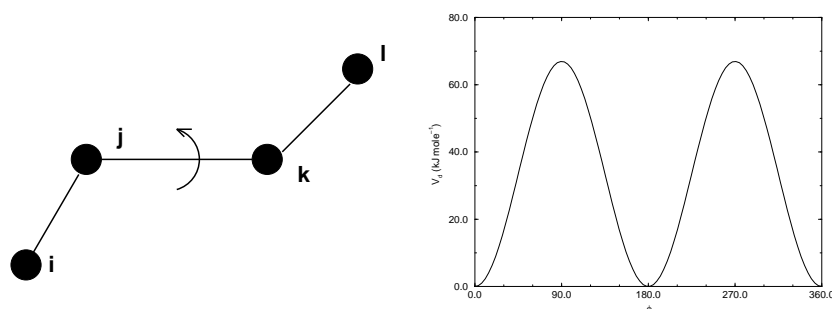


Figure 4.10: Principle of proper dihedral angle (left, in *trans* form) and the dihedral angle potential (right).

Improper dihedrals: periodic type

This potential is identical to the periodic proper dihedral (see below). There is a separate dihedral type for this (type 4) only to be able to distinguish improper from proper dihedrals in the parameter section and the output.

4.2.13 Proper dihedrals

For the normal dihedral interaction there is a choice of either the GROMOS periodic function or a function based on expansion in powers of $\cos \phi$ (the so-called Ryckaert-Bellemans potential). This choice has consequences for the inclusion of special interactions between the first and the fourth atom of the dihedral quadruple. With the periodic GROMOS potential a special 1-4 LJ-interaction must be included; with the Ryckaert-Bellemans potential *for alkanes* the 1-4 interactions must be excluded from the non-bonded list. **Note:** Ryckaert-Bellemans potentials are also used in *e.g.* the OPLS force field in combination with 1-4 interactions. You should therefore not modify topologies generated by `pdb2gmx` in this case.

Proper dihedrals: periodic type

Proper dihedral angles are defined according to the IUPAC/IUB convention, where ϕ is the angle between the ijk and the jkl planes, with **zero** corresponding to the *cis* configuration (i and l on the same side). There are two dihedral function types in GROMACS topology files. There is the standard type 1 which behaves like any other bonded interactions. For certain force fields, type 9 is useful. Type 9 allows multiple potential functions to be applied automatically to a single dihedral in the [`dihedral`] section when multiple parameters are defined for the same atomtypes in the [`dihedraltypes`] section.

$$V_d(\phi_{ijkl}) = k_\phi(1 + \cos(n\phi - \phi_s)) \quad (4.62)$$

C_0	9.28	C_2	-13.12	C_4	26.24
C_1	12.16	C_3	-3.06	C_5	-31.5

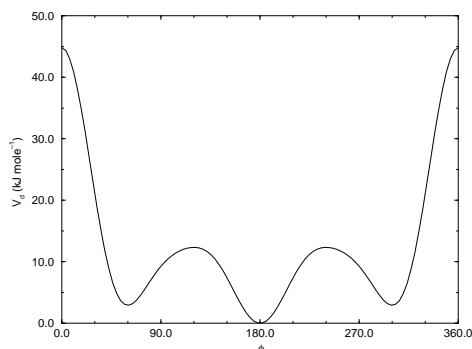
Table 4.1: Constants for Ryckaert-Bellemans potential (kJ mol^{-1}).

Figure 4.11: Ryckaert-Bellemans dihedral potential.

Proper dihedrals: Ryckaert-Bellemans function

For alkanes, the following proper dihedral potential is often used (see Fig. 4.11):

$$V_{rb}(\phi_{ijkl}) = \sum_{n=0}^5 C_n (\cos(\psi))^n, \quad (4.63)$$

where $\psi = \phi - 180^\circ$.

Note: A conversion from one convention to another can be achieved by multiplying every coefficient C_n by $(-1)^n$.

An example of constants for C is given in Table 4.1.

(Note: The use of this potential implies exclusion of LJ interactions between the first and the last atom of the dihedral, and ψ is defined according to the “polymer convention” ($\psi_{trans} = 0$.)

The RB dihedral function can also be used to include Fourier dihedrals (see below):

$$V_{rb}(\phi_{ijkl}) = \frac{1}{2} [F_1(1 + \cos(\phi)) + F_2(1 - \cos(2\phi)) + F_3(1 + \cos(3\phi)) + F_4(1 - \cos(4\phi))] \quad (4.64)$$

Because of the equalities $\cos(2\phi) = 2\cos^2(\phi) - 1$, $\cos(3\phi) = 4\cos^3(\phi) - 3\cos(\phi)$ and $\cos(4\phi) = 8\cos^4(\phi) - 8\cos^2(\phi) + 1$ one can translate the OPLS parameters to Ryckaert-Bellemans param-

eters as follows:

$$\begin{aligned}
 C_0 &= F_2 + \frac{1}{2}(F_1 + F_3) \\
 C_1 &= \frac{1}{2}(-F_1 + 3F_3) \\
 C_2 &= -F_2 + 4F_4 \\
 C_3 &= -2F_3 \\
 C_4 &= -4F_4 \\
 C_5 &= 0
 \end{aligned}
 \tag{4.65}$$

with OPLS parameters in protein convention and RB parameters in polymer convention (this yields a minus sign for the odd powers of $\cos(\phi)$).

Note: Mind the conversion from kcal mol^{-1} for literature OPLS and RB parameters to kJ mol^{-1} in GROMACS.

Proper dihedrals: Fourier function

The OPLS potential function is given as the first three [91] or four [92] cosine terms of a Fourier series. In GROMACS the four term function is implemented:

$$V_F(\phi_{ijkl}) = \frac{1}{2} [C_1(1 + \cos(\phi)) + C_2(1 - \cos(2\phi)) + C_3(1 + \cos(3\phi)) + C_4(1 - \cos(4\phi))],
 \tag{4.66}$$

Internally, GROMACS uses the Ryckaert-Bellemans code to compute Fourier dihedrals (see above), because this is more efficient.

Note: Mind the conversion from kcal mol^{-1} for literature OPLS parameters to kJ mol^{-1} in GROMACS.

Proper dihedrals: Restricted torsion potential

In a manner very similar to the restricted bending potential (see 4.2.7), a restricted torsion/dihedral potential is introduced:

$$V_{\text{ReT}}(\phi_i) = \frac{1}{2} k_\phi \frac{(\cos \phi_i - \cos \phi_0)^2}{\sin^2 \phi_i}
 \tag{4.67}$$

with the advantages of being a function of $\cos \phi$ (no problems taking the derivative of $\sin \phi$) and of keeping the torsion angle at only one minimum value. In this case, the factor $\sin^2 \phi$ does not allow the dihedral angle to move from the $[-180^\circ; 0]$ to $[0; 180^\circ]$ interval, i.e. it cannot have maxima both at $-\phi_0$ and $+\phi_0$ maxima, but only one of them. For this reason, all the dihedral angles of the starting configuration should have their values in the desired angles interval and the the equilibrium ϕ_0 value should not be too close to the interval limits (as for the restricted bending potential, described in 4.2.7, at least 10° difference is recommended).

Proper dihedrals: Combined bending-torsion potential

When the four particles forming the dihedral angle become collinear (this situation will never happen in atomistic simulations, but it can occur in coarse-grained simulations) the calculation of

the torsion angle and potential leads to numerical instabilities. One way to avoid this is to use the restricted bending potential (see 4.2.7) that prevents the dihedral from reaching the 180° value.

Another way is to disregard any effects of the dihedral becoming ill-defined, keeping the dihedral force and potential calculation continuous in entire angle range by coupling the torsion potential (in a cosine form) with the bending potentials of the adjacent bending angles in a unique expression:

$$V_{\text{CBT}}(\theta_{i-1}, \theta_i, \phi_i) = k_\phi \sin^3 \theta_{i-1} \sin^3 \theta_i \sum_{n=0}^4 a_n \cos^n \phi_i. \quad (4.68)$$

This combined bending-torsion (CBT) potential has been proposed by [93] for polymer melt simulations and is extensively described in [88].

This potential has two main advantages:

- it does not only depend on the dihedral angle ϕ_i (between the $i - 2$, $i - 1$, i and $i + 1$ beads) but also on the bending angles θ_{i-1} and θ_i defined from three adjacent beads ($i - 2$, $i - 1$ and i , and $i - 1$, i and $i + 1$, respectively). The two $\sin^3 \theta$ pre-factors, tentatively suggested by [94] and theoretically discussed by [95], cancel the torsion potential and force when either of the two bending angles approaches the value of 180° .
- its dependence on ϕ_i is expressed through a polynomial in $\cos \phi_i$ that avoids the singularities in $\phi = 0^\circ$ or 180° in calculating the torsional force.

These two properties make the CBT potential well-behaved for MD simulations with weak constraints on the bending angles or even for steered / non-equilibrium MD in which the bending and torsion angles suffer major modifications. When using the CBT potential, the bending potentials for the adjacent θ_{i-1} and θ_i may have any form. It is also possible to leave out the two angle bending terms (θ_{i-1} and θ_i) completely. Fig. 4.12 illustrates the difference between a torsion potential with and without the $\sin^3 \theta$ factors (blue and gray curves, respectively). Additionally, the derivative of V_{CBT} with respect to the Cartesian variables is straightforward:

$$\frac{\partial V_{\text{CBT}}(\theta_{i-1}, \theta_i, \phi_i)}{\partial \vec{r}_i} = \frac{\partial V_{\text{CBT}}}{\partial \theta_{i-1}} \frac{\partial \theta_{i-1}}{\partial \vec{r}_i} + \frac{\partial V_{\text{CBT}}}{\partial \theta_i} \frac{\partial \theta_i}{\partial \vec{r}_i} + \frac{\partial V_{\text{CBT}}}{\partial \phi_i} \frac{\partial \phi_i}{\partial \vec{r}_i} \quad (4.69)$$

The CBT is based on a cosine form without multiplicity, so it can only be symmetrical around 0° . To obtain an asymmetrical dihedral angle distribution (e.g. only one maximum in $[-180^\circ:180^\circ]$ interval), a standard torsion potential such as harmonic angle or periodic cosine potentials should be used instead of a CBT potential. However, these two forms have the inconveniences of the force derivation ($1/\sin \phi$) and of the alignment of beads (θ_i or $\theta_{i-1} = 0^\circ, 180^\circ$). Coupling such non- $\cos \phi$ potentials with $\sin^3 \theta$ factors does not improve simulation stability since there are cases in which θ and ϕ are simultaneously 180° . The integration at this step would be possible (due to the cancelling of the torsion potential) but the next step would be singular (θ is not 180° and ϕ is very close to 180°).

4.2.14 Tabulated bonded interaction functions

For full flexibility, any functional shape can be used for bonds, angles and dihedrals through user-supplied tabulated functions. The functional shapes are:

$$V_b(r_{ij}) = k f_n^b(r_{ij}) \quad (4.70)$$

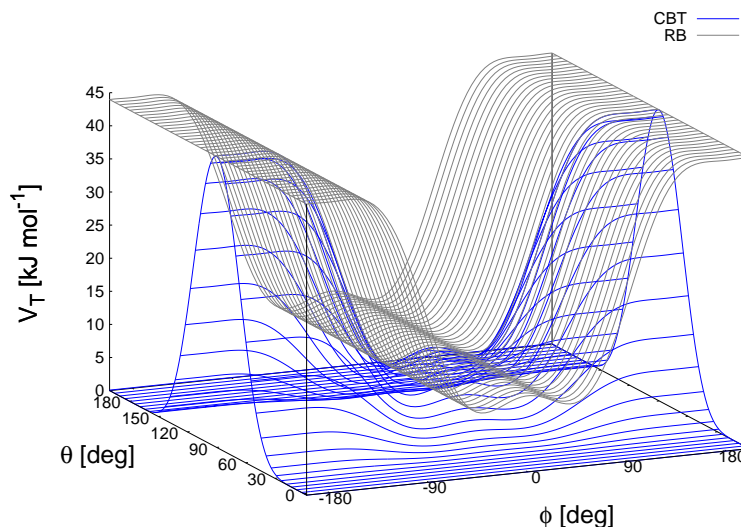


Figure 4.12: Blue: surface plot of the combined bending-torsion potential (4.68 with $k = 10$ kJ mol^{-1} , $a_0 = 2.41$, $a_1 = -2.95$, $a_2 = 0.36$, $a_3 = 1.33$) when, for simplicity, the bending angles behave the same ($\theta_1 = \theta_2 = \theta$). Gray: the same torsion potential without the $\sin^3 \theta$ terms (Ryckaert-Bellemans type). ϕ is the dihedral angle.

$$V_a(\theta_{ijk}) = k f_n^a(\theta_{ijk}) \quad (4.71)$$

$$V_d(\phi_{ijkl}) = k f_n^d(\phi_{ijkl}) \quad (4.72)$$

where k is a force constant in units of energy and f is a cubic spline function; for details see 6.12.1. For each interaction, the force constant k and the table number n are specified in the topology. There are two different types of bonds, one that generates exclusions (type 8) and one that does not (type 9). For details see Table 5.5. The table files are supplied to the `mdrun` program. After the table file name an underscore, the letter “b” for bonds, “a” for angles or “d” for dihedrals and the table number must be appended. For example, a tabulated bond with $n = 0$ can be read from the file `table_b0.xvg`. Multiple tables can be supplied simply by adding files with different values of n , and are applied to the appropriate bonds, as specified in the topology (Table 5.5). The format for the table files is three fixed-format columns of any suitable width. These columns must contain x , $f(x)$, $-f'(x)$, and the values of x should be uniformly spaced. Requirements for entries in the topology are given in Table 5.5. The setup of the tables is as follows:

bonds: x is the distance in nm. For distances beyond the table length, `mdrun` will quit with an error message.

angles: x is the angle in degrees. The table should go from 0 up to and including 180 degrees; the derivative is taken in degrees.

dihedrals: x is the dihedral angle in degrees. The table should go from -180 up to and including 180 degrees; the IUPAC/IUB convention is used, *i.e.* zero is cis, the derivative is taken in degrees.

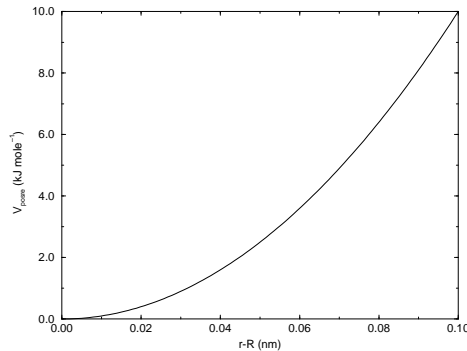


Figure 4.13: Position restraint potential.

4.3 Restraints

Special potentials are used for imposing restraints on the motion of the system, either to avoid disastrous deviations, or to include knowledge from experimental data. In either case they are not really part of the force field and the reliability of the parameters is not important. The potential forms, as implemented in GROMACS, are mentioned just for the sake of completeness. Restraints and constraints refer to quite different algorithms in GROMACS.

4.3.1 Position restraints

These are used to restrain particles to fixed reference positions \mathbf{R}_i . They can be used during equilibration in order to avoid drastic rearrangements of critical parts (*e.g.* to restrain motion in a protein that is subjected to large solvent forces when the solvent is not yet equilibrated). Another application is the restraining of particles in a shell around a region that is simulated in detail, while the shell is only approximated because it lacks proper interaction from missing particles outside the shell. Restraining will then maintain the integrity of the inner part. For spherical shells, it is a wise procedure to make the force constant depend on the radius, increasing from zero at the inner boundary to a large value at the outer boundary. This feature has not, however, been implemented in GROMACS.

The following form is used:

$$V_{pr}(\mathbf{r}_i) = \frac{1}{2} k_{pr} |\mathbf{r}_i - \mathbf{R}_i|^2 \quad (4.73)$$

The potential is plotted in Fig. 4.13.

The potential form can be rewritten without loss of generality as:

$$V_{pr}(\mathbf{r}_i) = \frac{1}{2} \left[k_{pr}^x (x_i - X_i)^2 \hat{\mathbf{x}} + k_{pr}^y (y_i - Y_i)^2 \hat{\mathbf{y}} + k_{pr}^z (z_i - Z_i)^2 \hat{\mathbf{z}} \right] \quad (4.74)$$

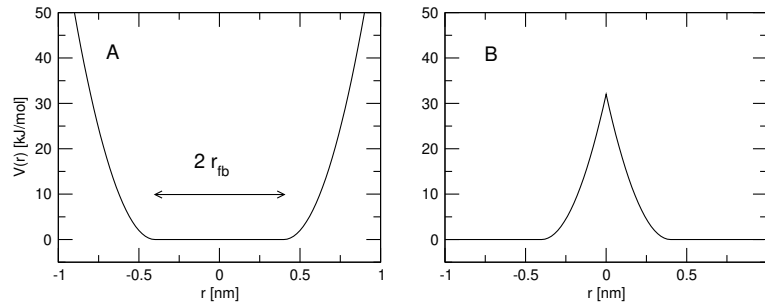


Figure 4.14: Flat-bottomed position restraint potential. (A) Not inverted, (B) inverted.

Now the forces are:

$$\begin{aligned} F_i^x &= -k_{pr}^x (x_i - X_i) \\ F_i^y &= -k_{pr}^y (y_i - Y_i) \\ F_i^z &= -k_{pr}^z (z_i - Z_i) \end{aligned} \quad (4.75)$$

Using three different force constants the position restraints can be turned on or off in each spatial dimension; this means that atoms can be harmonically restrained to a plane or a line. Position restraints are applied to a special fixed list of atoms. Such a list is usually generated by the `pdb2gmx` program.

4.3.2 Flat-bottomed position restraints

Flat-bottomed position restraints can be used to restrain particles to part of the simulation volume. No force acts on the restrained particle within the flat-bottomed region of the potential, however a harmonic force acts to move the particle to the flat-bottomed region if it is outside it. It is possible to apply normal and flat-bottomed position restraints on the same particle (however, only with the same reference position \mathbf{R}_i). The following general potential is used (Figure 4.14A):

$$V_{fb}(\mathbf{r}_i) = \frac{1}{2} k_{fb} [d_g(\mathbf{r}_i; \mathbf{R}_i) - r_{fb}]^2 H[d_g(\mathbf{r}_i; \mathbf{R}_i) - r_{fb}], \quad (4.76)$$

where \mathbf{R}_i is the reference position, r_{fb} is the distance from the center with a flat potential, k_{fb} the force constant, and H is the Heaviside step function. The distance $d_g(\mathbf{r}_i; \mathbf{R}_i)$ from the reference position depends on the geometry g of the flat-bottomed potential.

The following geometries for the flat-bottomed potential are supported:

Sphere ($g = 1$): The particle is kept in a sphere of given radius. The force acts towards the center of the sphere. The following distance calculation is used:

$$d_g(\mathbf{r}_i; \mathbf{R}_i) = |\mathbf{r}_i - \mathbf{R}_i| \quad (4.77)$$

Cylinder ($g = 6, 7, 8$): The particle is kept in a cylinder of given radius parallel to the x ($g = 6$), y ($g = 7$), or z -axis ($g = 8$). For backwards compatibility, setting $g = 2$ is mapped to $g = 8$ in the code so that old `.top` files and topologies work. The force from the flat-bottomed potential acts towards the axis of the cylinder. The component of the force parallel to the cylinder axis is zero. For a cylinder aligned along the z -axis:

$$d_g(\mathbf{r}_i; \mathbf{R}_i) = \sqrt{(x_i - X_i)^2 + (y_i - Y_i)^2} \quad (4.78)$$

Layer ($g = 3, 4, 5$): The particle is kept in a layer defined by the thickness and the normal of the layer. The layer normal can be parallel to the x , y , or z -axis. The force acts parallel to the layer normal.

$$d_g(\mathbf{r}_i; \mathbf{R}_i) = |x_i - X_i|, \quad \text{or} \quad d_g(\mathbf{r}_i; \mathbf{R}_i) = |y_i - Y_i|, \quad \text{or} \quad d_g(\mathbf{r}_i; \mathbf{R}_i) = |z_i - Z_i|. \quad (4.79)$$

It is possible to apply multiple independent flat-bottomed position restraints of different geometry on one particle. For example, applying a cylinder and a layer in z keeps a particle within a disk. Applying three layers in x , y , and z keeps the particle within a cuboid.

In addition, it is possible to invert the restrained region with the unrestrained region, leading to a potential that acts to keep the particle *outside* of the volume defined by \mathbf{R}_i , g , and r_{fb} . That feature is switched on by defining a negative r_{fb} in the topology. The following potential is used (Figure 4.14B):

$$V_{fb}^{\text{inv}}(\mathbf{r}_i) = \frac{1}{2}k_{fb}[d_g(\mathbf{r}_i; \mathbf{R}_i) - |r_{fb}|]^2 H[-(d_g(\mathbf{r}_i; \mathbf{R}_i) - |r_{fb}|)]. \quad (4.80)$$

4.3.3 Angle restraints

These are used to restrain the angle between two pairs of particles or between one pair of particles and the z -axis. The functional form is similar to that of a proper dihedral. For two pairs of atoms:

$$V_{ar}(\mathbf{r}_i, \mathbf{r}_j, \mathbf{r}_k, \mathbf{r}_l) = k_{ar}(1 - \cos(n(\theta - \theta_0))), \quad \text{where} \quad \theta = \arccos\left(\frac{\mathbf{r}_j - \mathbf{r}_i}{\|\mathbf{r}_j - \mathbf{r}_i\|} \cdot \frac{\mathbf{r}_l - \mathbf{r}_k}{\|\mathbf{r}_l - \mathbf{r}_k\|}\right) \quad (4.81)$$

For one pair of atoms and the z -axis:

$$V_{ar}(\mathbf{r}_i, \mathbf{r}_j) = k_{ar}(1 - \cos(n(\theta - \theta_0))), \quad \text{where} \quad \theta = \arccos\left(\frac{\mathbf{r}_j - \mathbf{r}_i}{\|\mathbf{r}_j - \mathbf{r}_i\|} \cdot \begin{pmatrix} 0 \\ 0 \\ 1 \end{pmatrix}\right) \quad (4.82)$$

A multiplicity (n) of 2 is useful when you do not want to distinguish between parallel and anti-parallel vectors. The equilibrium angle θ should be between 0 and 180 degrees for multiplicity 1 and between 0 and 90 degrees for multiplicity 2.

4.3.4 Dihedral restraints

These are used to restrain the dihedral angle ϕ defined by four particles as in an improper dihedral (sec. 4.2.12) but with a slightly modified potential. Using:

$$\phi' = (\phi - \phi_0) \text{ MOD } 2\pi \quad (4.83)$$

where ϕ_0 is the reference angle, the potential is defined as:

$$V_{dih}(\phi') = \begin{cases} \frac{1}{2}k_{dih}(\phi' - \phi_0 - \Delta\phi)^2 & \text{for } \phi' > \Delta\phi \\ 0 & \text{for } \phi' \leq \Delta\phi \end{cases} \quad (4.84)$$

where $\Delta\phi$ is a user defined angle and k_{dih} is the force constant. **Note** that in the input in topology files, angles are given in degrees and force constants in kJ/mol/rad².

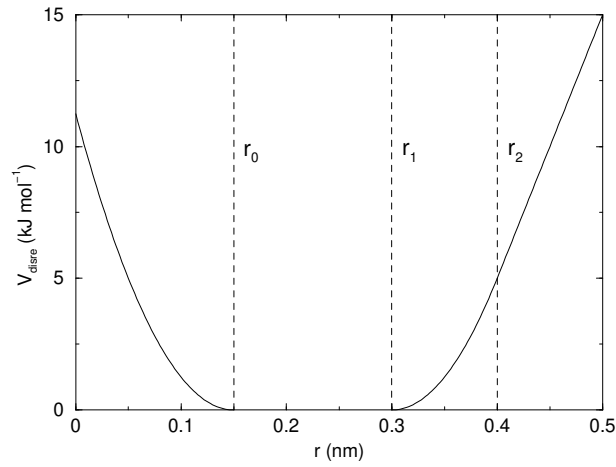


Figure 4.15: Distance Restraint potential.

4.3.5 Distance restraints

Distance restraints add a penalty to the potential when the distance between specified pairs of atoms exceeds a threshold value. They are normally used to impose experimental restraints from, for instance, experiments in nuclear magnetic resonance (NMR), on the motion of the system. Thus, MD can be used for structure refinement using NMR data. In GROMACS there are three ways to impose restraints on pairs of atoms:

- Simple harmonic restraints: use [bonds] type 6 (see sec. 5.4.4).
- Piecewise linear/harmonic restraints: [bonds] type 10.
- Complex NMR distance restraints, optionally with pair, time and/or ensemble averaging.

The last two options will be detailed now.

The potential form for distance restraints is quadratic below a specified lower bound and between two specified upper bounds, and linear beyond the largest bound (see Fig. 4.15).

$$V_{dr}(r_{ij}) = \begin{cases} \frac{1}{2}k_{dr}(r_{ij} - r_0)^2 & \text{for } r_{ij} < r_0 \\ 0 & \text{for } r_0 \leq r_{ij} < r_1 \\ \frac{1}{2}k_{dr}(r_{ij} - r_1)^2 & \text{for } r_1 \leq r_{ij} < r_2 \\ \frac{1}{2}k_{dr}(r_2 - r_1)(2r_{ij} - r_2 - r_1) & \text{for } r_2 \leq r_{ij} \end{cases} \quad (4.85)$$

The forces are

$$\mathbf{F}_i = \begin{cases} -k_{dr}(r_{ij} - r_0)\frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } r_{ij} < r_0 \\ 0 & \text{for } r_0 \leq r_{ij} < r_1 \\ -k_{dr}(r_{ij} - r_1)\frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } r_1 \leq r_{ij} < r_2 \\ -k_{dr}(r_2 - r_1)\frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } r_2 \leq r_{ij} \end{cases} \quad (4.86)$$

For restraints not derived from NMR data, this functionality will usually suffice and a section of [`bonds`] type 10 can be used to apply individual restraints between pairs of atoms, see 5.8.1. For applying restraints derived from NMR measurements, more complex functionality might be required, which is provided through the [`distance_restraints`] section and is described below.

Time averaging

Distance restraints based on instantaneous distances can potentially reduce the fluctuations in a molecule significantly. This problem can be overcome by restraining to a *time averaged* distance [96]. The forces with time averaging are:

$$\mathbf{F}_i = \begin{cases} -k_{dr}^a (\bar{r}_{ij} - r_0) \frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } \bar{r}_{ij} < r_0 \\ 0 & \text{for } r_0 \leq \bar{r}_{ij} < r_1 \\ -k_{dr}^a (\bar{r}_{ij} - r_1) \frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } r_1 \leq \bar{r}_{ij} < r_2 \\ -k_{dr}^a (r_2 - r_1) \frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } r_2 \leq \bar{r}_{ij} \end{cases} \quad (4.87)$$

where \bar{r}_{ij} is given by an exponential running average with decay time τ :

$$\bar{r}_{ij} = \langle r_{ij}^{-3} \rangle^{-1/3} \quad (4.88)$$

The force constant k_{dr}^a is switched on slowly to compensate for the lack of history at the beginning of the simulation:

$$k_{dr}^a = k_{dr} \left(1 - \exp\left(-\frac{t}{\tau}\right) \right) \quad (4.89)$$

Because of the time averaging, we can no longer speak of a distance restraint potential.

This way an atom can satisfy two incompatible distance restraints *on average* by moving between two positions. An example would be an amino acid side-chain that is rotating around its χ dihedral angle, thereby coming close to various other groups. Such a mobile side chain can give rise to multiple NOEs that can not be fulfilled by a single structure.

The computation of the time averaged distance in the `mdrun` program is done in the following fashion:

$$\begin{aligned} \bar{r}_{ij}^{-3}(0) &= r_{ij}(0)^{-3} \\ \bar{r}_{ij}^{-3}(t) &= \bar{r}_{ij}^{-3}(t - \Delta t) \exp\left(-\frac{\Delta t}{\tau}\right) + r_{ij}(t)^{-3} \left[1 - \exp\left(-\frac{\Delta t}{\tau}\right) \right] \end{aligned} \quad (4.90)$$

When a pair is within the bounds, it can still feel a force because the time averaged distance can still be beyond a bound. To prevent the protons from being pulled too close together, a mixed approach can be used. In this approach, the penalty is zero when the instantaneous distance is within the bounds, otherwise the violation is the square root of the product of the instantaneous violation and the time averaged violation:

$$\mathbf{F}_i = \begin{cases} k_{dr}^a \sqrt{(r_{ij} - r_0)(\bar{r}_{ij} - r_0)} \frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } r_{ij} < r_0 \text{ and } \bar{r}_{ij} < r_0 \\ -k_{dr}^a \min\left(\sqrt{(r_{ij} - r_1)(\bar{r}_{ij} - r_1)}, r_2 - r_1\right) \frac{\mathbf{r}_{ij}}{r_{ij}} & \text{for } r_{ij} > r_1 \text{ and } \bar{r}_{ij} > r_1 \\ 0 & \text{otherwise} \end{cases} \quad (4.91)$$

Averaging over multiple pairs

Sometimes it is unclear from experimental data which atom pair gives rise to a single NOE, in other occasions it can be obvious that more than one pair contributes due to the symmetry of the system, *e.g.* a methyl group with three protons. For such a group, it is not possible to distinguish between the protons, therefore they should all be taken into account when calculating the distance between this methyl group and another proton (or group of protons). Due to the physical nature of magnetic resonance, the intensity of the NOE signal is inversely proportional to the sixth power of the inter-atomic distance. Thus, when combining atom pairs, a fixed list of N restraints may be taken together, where the apparent “distance” is given by:

$$r_N(t) = \left[\sum_{n=1}^N \bar{r}_n(t)^{-6} \right]^{-1/6} \quad (4.92)$$

where we use r_{ij} or eqn. 4.88 for the \bar{r}_n . The r_N of the instantaneous and time-averaged distances can be combined to do a mixed restraining, as indicated above. As more pairs of protons contribute to the same NOE signal, the intensity will increase, and the summed “distance” will be shorter than any of its components due to the reciprocal summation.

There are two options for distributing the forces over the atom pairs. In the conservative option, the force is defined as the derivative of the restraint potential with respect to the coordinates. This results in a conservative potential when time averaging is not used. The force distribution over the pairs is proportional to r^{-6} . This means that a close pair feels a much larger force than a distant pair, which might lead to a molecule that is “too rigid.” The other option is an equal force distribution. In this case each pair feels $1/N$ of the derivative of the restraint potential with respect to r_N . The advantage of this method is that more conformations might be sampled, but the non-conservative nature of the forces can lead to local heating of the protons.

It is also possible to use *ensemble averaging* using multiple (protein) molecules. In this case the bounds should be lowered as in:

$$\begin{aligned} r_1 &= r_1 * M^{-1/6} \\ r_2 &= r_2 * M^{-1/6} \end{aligned} \quad (4.93)$$

where M is the number of molecules. The GROMACS preprocessor `grompp` can do this automatically when the appropriate option is given. The resulting “distance” is then used to calculate the scalar force according to:

$$\mathbf{F}_i = \begin{cases} 0 & r_N < r_1 \\ k_{dr}(r_N - r_1) \frac{\mathbf{r}_{ij}}{r_{ij}} & r_1 \leq r_N < r_2 \\ k_{dr}(r_2 - r_1) \frac{\mathbf{r}_{ij}}{r_{ij}} & r_N \geq r_2 \end{cases} \quad (4.94)$$

where i and j denote the atoms of all the pairs that contribute to the NOE signal.

Using distance restraints

A list of distance restraints based on NOE data can be added to a molecule definition in your topology file, like in the following example:

```
[ distance_restraints ]
; ai   aj   type   index   type'   low   up1   up2   fac
10    16    1      0      1      0.0  0.3  0.4  1.0
10    28    1      1      1      0.0  0.3  0.4  1.0
10    46    1      1      1      0.0  0.3  0.4  1.0
16    22    1      2      1      0.0  0.3  0.4  2.5
16    34    1      3      1      0.0  0.5  0.6  1.0
```

In this example a number of features can be found. In columns `ai` and `aj` you find the atom numbers of the particles to be restrained. The `type` column should always be 1. As explained in 4.3.5, multiple distances can contribute to a single NOE signal. In the topology this can be set using the `index` column. In our example, the restraints 10-28 and 10-46 both have index 1, therefore they are treated simultaneously. An extra requirement for treating restraints together is that the restraints must be on successive lines, without any other intervening restraint. The `type'` column will usually be 1, but can be set to 2 to obtain a distance restraint that will never be time- and ensemble-averaged; this can be useful for restraining hydrogen bonds. The columns `low`, `up1`, and `up2` hold the values of r_0 , r_1 , and r_2 from eqn. 4.85. In some cases it can be useful to have different force constants for some restraints; this is controlled by the column `fac`. The force constant in the parameter file is multiplied by the value in the column `fac` for each restraint. Information for each restraint is stored in the energy file and can be processed and plotted with `gmx nmr`.

4.3.6 Orientation restraints

This section describes how orientations between vectors, as measured in certain NMR experiments, can be calculated and restrained in MD simulations. The presented refinement methodology and a comparison of results with and without time and ensemble averaging have been published [97].

Theory

In an NMR experiment, orientations of vectors can be measured when a molecule does not tumble completely isotropically in the solvent. Two examples of such orientation measurements are residual dipolar couplings (between two nuclei) or chemical shift anisotropies. An observable for a vector \mathbf{r}_i can be written as follows:

$$\delta_i = \frac{2}{3} \text{tr}(\mathbf{S}\mathbf{D}_i) \quad (4.95)$$

where \mathbf{S} is the dimensionless order tensor of the molecule. The tensor \mathbf{D}_i is given by:

$$\mathbf{D}_i = \frac{c_i}{\|\mathbf{r}_i\|^\alpha} \begin{pmatrix} 3xx - 1 & 3xy & 3xz \\ 3xy & 3yy - 1 & 3yz \\ 3xz & 3yz & 3zz - 1 \end{pmatrix} \quad (4.96)$$

$$\text{with: } x = \frac{r_{i,x}}{\|\mathbf{r}_i\|}, \quad y = \frac{r_{i,y}}{\|\mathbf{r}_i\|}, \quad z = \frac{r_{i,z}}{\|\mathbf{r}_i\|} \quad (4.97)$$

For a dipolar coupling r_i is the vector connecting the two nuclei, $\alpha = 3$ and the constant c_i is given by:

$$c_i = \frac{\mu_0}{4\pi} \gamma_1^i \gamma_2^i \frac{\hbar}{4\pi} \quad (4.98)$$

where γ_1^i and γ_2^i are the gyromagnetic ratios of the two nuclei.

The order tensor is symmetric and has trace zero. Using a rotation matrix \mathbf{T} it can be transformed into the following form:

$$\mathbf{T}^T \mathbf{S} \mathbf{T} = s \begin{pmatrix} -\frac{1}{2}(1-\eta) & 0 & 0 \\ 0 & -\frac{1}{2}(1+\eta) & 0 \\ 0 & 0 & 1 \end{pmatrix} \quad (4.99)$$

where $-1 \leq s \leq 1$ and $0 \leq \eta \leq 1$. s is called the order parameter and η the asymmetry of the order tensor \mathbf{S} . When the molecule tumbles isotropically in the solvent, s is zero, and no orientational effects can be observed because all δ_i are zero.

Calculating orientations in a simulation

For reasons which are explained below, the \mathbf{D} matrices are calculated which respect to a reference orientation of the molecule. The orientation is defined by a rotation matrix \mathbf{R} , which is needed to least-squares fit the current coordinates of a selected set of atoms onto a reference conformation. The reference conformation is the starting conformation of the simulation. In case of ensemble averaging, which will be treated later, the structure is taken from the first subsystem. The calculated \mathbf{D}_i^c matrix is given by:

$$\mathbf{D}_i^c(t) = \mathbf{R}(t) \mathbf{D}_i(t) \mathbf{R}^T(t) \quad (4.100)$$

The calculated orientation for vector i is given by:

$$\delta_i^c(t) = \frac{2}{3} \text{tr}(\mathbf{S}(t) \mathbf{D}_i^c(t)) \quad (4.101)$$

The order tensor $\mathbf{S}(t)$ is usually unknown. A reasonable choice for the order tensor is the tensor which minimizes the (weighted) mean square difference between the calculated and the observed orientations:

$$MSD(t) = \left(\sum_{i=1}^N w_i \right)^{-1} \sum_{i=1}^N w_i (\delta_i^c(t) - \delta_i^{exp})^2 \quad (4.102)$$

To properly combine different types of measurements, the unit of w_i should be such that all terms are dimensionless. This means the unit of w_i is the unit of δ_i to the power -2 . **Note** that scaling all w_i with a constant factor does not influence the order tensor.

Time averaging

Since the tensors \mathbf{D}_i fluctuate rapidly in time, much faster than can be observed in an experiment, they should be averaged over time in the simulation. However, in a simulation the time and the number of copies of a molecule are limited. Usually one can not obtain a converged average of the \mathbf{D}_i tensors over all orientations of the molecule. If one assumes that the average orientations of

the \mathbf{r}_i vectors within the molecule converge much faster than the tumbling time of the molecule, the tensor can be averaged in an axis system that rotates with the molecule, as expressed by equation (4.100). The time-averaged tensors are calculated using an exponentially decaying memory function:

$$\mathbf{D}_i^a(t) = \frac{\int_{u=t_0}^t \mathbf{D}_i^c(u) \exp\left(-\frac{t-u}{\tau}\right) du}{\int_{u=t_0}^t \exp\left(-\frac{t-u}{\tau}\right) du} \quad (4.103)$$

Assuming that the order tensor \mathbf{S} fluctuates slower than the \mathbf{D}_i , the time-averaged orientation can be calculated as:

$$\delta_i^a(t) = \frac{2}{3} \text{tr}(\mathbf{S}(t) \mathbf{D}_i^a(t)) \quad (4.104)$$

where the order tensor $\mathbf{S}(t)$ is calculated using expression (4.102) with $\delta_i^c(t)$ replaced by $\delta_i^a(t)$.

Restraining

The simulated structure can be restrained by applying a force proportional to the difference between the calculated and the experimental orientations. When no time averaging is applied, a proper potential can be defined as:

$$V = \frac{1}{2} k \sum_{i=1}^N w_i (\delta_i^c(t) - \delta_i^{exp})^2 \quad (4.105)$$

where the unit of k is the unit of energy. Thus the effective force constant for restraint i is $k w_i$. The forces are given by minus the gradient of V . The force \mathbf{F}_i working on vector \mathbf{r}_i is:

$$\begin{aligned} \mathbf{F}_i(t) &= -\frac{dV}{d\mathbf{r}_i} \\ &= -k w_i (\delta_i^c(t) - \delta_i^{exp}) \frac{d\delta_i^c(t)}{d\mathbf{r}_i} \\ &= -k w_i (\delta_i^c(t) - \delta_i^{exp}) \frac{2c_i}{\|\mathbf{r}_i\|^{2+\alpha}} \left(2\mathbf{R}^T \mathbf{S} \mathbf{R} \mathbf{r}_i - \frac{2+\alpha}{\|\mathbf{r}_i\|^2} \text{tr}(\mathbf{R}^T \mathbf{S} \mathbf{R} \mathbf{r}_i \mathbf{r}_i^T) \mathbf{r}_i \right) \end{aligned}$$

Ensemble averaging

Ensemble averaging can be applied by simulating a system of M subsystems that each contain an identical set of orientation restraints. The systems only interact via the orientation restraint potential which is defined as:

$$V = M \frac{1}{2} k \sum_{i=1}^N w_i (\delta_i^c(t) - \delta_i^{exp})^2 \quad (4.106)$$

The force on vector $\mathbf{r}_{i,m}$ in subsystem m is given by:

$$\mathbf{F}_{i,m}(t) = -\frac{dV}{d\mathbf{r}_{i,m}} = -k w_i (\delta_i^c(t) - \delta_i^{exp}) \frac{d\delta_{i,m}^c(t)}{d\mathbf{r}_{i,m}} \quad (4.107)$$

Time averaging

When using time averaging it is not possible to define a potential. We can still define a quantity that gives a rough idea of the energy stored in the restraints:

$$V = M \frac{1}{2} k^a \sum_{i=1}^N w_i \langle \delta_i^a(t) - \delta_i^{exp} \rangle^2 \quad (4.108)$$

The force constant k_a is switched on slowly to compensate for the lack of history at times close to t_0 . It is exactly proportional to the amount of average that has been accumulated:

$$k^a = k \frac{1}{\tau} \int_{u=t_0}^t \exp\left(-\frac{t-u}{\tau}\right) du \quad (4.109)$$

What really matters is the definition of the force. It is chosen to be proportional to the square root of the product of the time-averaged and the instantaneous deviation. Using only the time-averaged deviation induces large oscillations. The force is given by:

$$\mathbf{F}_{i,m}(t) = \begin{cases} 0 & \text{for } a b \leq 0 \\ k^a w_i \frac{a}{|a|} \sqrt{a b} \frac{d\delta_{i,m}^c(t)}{d\mathbf{r}_{i,m}} & \text{for } a b > 0 \end{cases} \quad (4.110)$$

$$a = \langle \delta_i^a(t) - \delta_i^{exp} \rangle$$

$$b = \langle \delta_i^c(t) - \delta_i^{exp} \rangle$$

Using orientation restraints

Orientation restraints can be added to a molecule definition in the topology file in the section [orientation_restraints]. Here we give an example section containing five N-H residual dipolar coupling restraints:

```
[ orientation_restraints ]
; ai  aj  type  exp.  label  alpha  const.  obs.  weight
;                                Hz      nm^3    Hz      Hz^-2
  31  32    1    1     3       3     6.083  -6.73  1.0
  43  44    1    1     4       3     6.083  -7.87  1.0
  55  56    1    1     5       3     6.083  -7.13  1.0
  65  66    1    1     6       3     6.083  -2.57  1.0
  73  74    1    1     7       3     6.083  -2.10  1.0
```

The unit of the observable is Hz, but one can choose any other unit. In columns `ai` and `aj` you find the atom numbers of the particles to be restrained. The `type` column should always be 1. The `exp.` column denotes the experiment number, starting at 1. For each experiment a separate order tensor \mathbf{S} is optimized. The label should be a unique number larger than zero for each restraint. The `alpha` column contains the power α that is used in equation (4.96) to calculate the orientation. The `const.` column contains the constant c_i used in the same equation. The constant should

have the unit of the observable times nm^α . The column `obs.` contains the observable, in any unit you like. The last column contains the weights w_i ; the unit should be the inverse of the square of the unit of the observable.

Some parameters for orientation restraints can be specified in the `grompp.mdp` file, for a study of the effect of different force constants and averaging times and ensemble averaging see [97]. Information for each restraint is stored in the energy file and can be processed and plotted with `gmx nmr`.

4.4 Polarization

Polarization can be treated by GROMACS by attaching shell (Drude) particles to atoms and/or virtual sites. The energy of the shell particle is then minimized at each time step in order to remain on the Born-Oppenheimer surface.

4.4.1 Simple polarization

This is implemented as a harmonic potential with equilibrium distance 0. The input given in the topology file is the polarizability α (in GROMACS units) as follows:

```
[ polarization ]
; Atom i j type alpha
1      2 1   0.001
```

in this case the polarizability volume is 0.001 nm^3 (or 1 \AA^3). In order to compute the harmonic force constant k_{cs} (where *cs* stands for core-shell), the following is used [45]:

$$k_{cs} = \frac{q_s^2}{\alpha} \quad (4.111)$$

where q_s is the charge on the shell particle.

4.4.2 Anharmonic polarization

For the development of the Drude force field by Roux and McKerell [98] it was found that some particles can overpolarize and this was fixed by introducing a higher order term in the polarization energy:

$$V_{pol} = \frac{k_{cs}}{2} r_{cs}^2 \quad r_{cs} \leq \delta \quad (4.112)$$

$$= \frac{k_{cs}}{2} r_{cs}^2 + k_{hyp} (r_{cs} - \delta)^4 \quad r_{cs} > \delta \quad (4.113)$$

where δ is a user-defined constant that is set to 0.02 nm for anions in the Drude force field [99]. Since this original introduction it has also been used in other atom types [98].

```
[ polarization ]
;Atom i j type alpha (nm^3) delta khyp
1      2 2   0.001786 0.02 16.736e8
```

The above force constant k_{hyp} corresponds to $4 \cdot 10^8 \text{ kcal/mol/nm}^4$, hence the strange number.

4.4.3 Water polarization

A special potential for water that allows anisotropic polarization of a single shell particle [45].

4.4.4 Thole polarization

Based on early work by Thole [100], Roux and coworkers have implemented potentials for molecules like ethanol [101, 102, 103]. Within such molecules, there are intra-molecular interactions between shell particles, however these must be screened because full Coulomb would be too strong. The potential between two shell particles i and j is:

$$V_{thole} = \frac{q_i q_j}{r_{ij}} \left[1 - \left(1 + \frac{\bar{r}_{ij}}{2} \right) \exp^{-\bar{r}_{ij}} \right] \quad (4.114)$$

Note that there is a sign error in Equation 1 of Noskov *et al.* [103]:

$$\bar{r}_{ij} = a \frac{r_{ij}}{(\alpha_i \alpha_j)^{1/6}} \quad (4.115)$$

where a is a magic (dimensionless) constant, usually chosen to be 2.6 [103]; α_i and α_j are the polarizabilities of the respective shell particles.

4.5 Free energy interactions

This section describes the λ -dependence of the potentials used for free energy calculations (see sec. 3.12). All common types of potentials and constraints can be interpolated smoothly from state A ($\lambda = 0$) to state B ($\lambda = 1$) and vice versa. All bonded interactions are interpolated by linear interpolation of the interaction parameters. Non-bonded interactions can be interpolated linearly or via soft-core interactions.

Starting in GROMACS 4.6, λ is a vector, allowing different components of the free energy transformation to be carried out at different rates. Coulomb, Lennard-Jones, bonded, and restraint terms can all be controlled independently, as described in the `.mdp` options.

Harmonic potentials

The example given here is for the bond potential, which is harmonic in GROMACS. However, these equations apply to the angle potential and the improper dihedral potential as well.

$$V_b = \frac{1}{2} \left[(1 - \lambda) k_b^A + \lambda k_b^B \right] \left[b - (1 - \lambda) b_0^A - \lambda b_0^B \right]^2 \quad (4.116)$$

$$\begin{aligned} \frac{\partial V_b}{\partial \lambda} &= \frac{1}{2} (k_b^B - k_b^A) \left[b - (1 - \lambda) b_0^A + \lambda b_0^B \right]^2 + \\ &\quad (b_0^A - b_0^B) \left[b - (1 - \lambda) b_0^A - \lambda b_0^B \right] \left[(1 - \lambda) k_b^A + \lambda k_b^B \right] \end{aligned} \quad (4.117)$$

GROMOS-96 bonds and angles

Fourth-power bond stretching and cosine-based angle potentials are interpolated by linear interpolation of the force constant and the equilibrium position. Formulas are not given here.

Proper dihedrals

For the proper dihedrals, the equations are somewhat more complicated:

$$V_d = [(1 - \lambda)k_d^A + \lambda k_d^B] \left(1 + \cos [n_\phi \phi - (1 - \lambda)\phi_s^A - \lambda\phi_s^B]\right) \quad (4.118)$$

$$\begin{aligned} \frac{\partial V_d}{\partial \lambda} &= (k_d^B - k_d^A) \left(1 + \cos [n_\phi \phi - (1 - \lambda)\phi_s^A - \lambda\phi_s^B]\right) + \\ &(\phi_s^B - \phi_s^A) [(1 - \lambda)k_d^A - \lambda k_d^B] \sin [n_\phi \phi - (1 - \lambda)\phi_s^A - \lambda\phi_s^B] \end{aligned} \quad (4.119)$$

Note: that the multiplicity n_ϕ can not be parameterized because the function should remain periodic on the interval $[0, 2\pi]$.

Tabulated bonded interactions

For tabulated bonded interactions only the force constant can be interpolated:

$$V = ((1 - \lambda)k^A + \lambda k^B) f \quad (4.120)$$

$$\frac{\partial V}{\partial \lambda} = (k^B - k^A) f \quad (4.121)$$

Coulomb interaction

The Coulomb interaction between two particles of which the charge varies with λ is:

$$V_c = \frac{f}{\varepsilon_{rf} r_{ij}} [(1 - \lambda)q_i^A q_j^A + \lambda q_i^B q_j^B] \quad (4.122)$$

$$\frac{\partial V_c}{\partial \lambda} = \frac{f}{\varepsilon_{rf} r_{ij}} [-q_i^A q_j^A + q_i^B q_j^B] \quad (4.123)$$

where $f = \frac{1}{4\pi\varepsilon_0} = 138.935\,458$ (see chapter 2).

Coulomb interaction with reaction field

The Coulomb interaction including a reaction field, between two particles of which the charge varies with λ is:

$$V_c = f \left[\frac{1}{r_{ij}} + k_{rf} r_{ij}^2 - c_{rf} \right] [(1 - \lambda)q_i^A q_j^A + \lambda q_i^B q_j^B] \quad (4.124)$$

$$\frac{\partial V_c}{\partial \lambda} = f \left[\frac{1}{r_{ij}} + k_{rf} r_{ij}^2 - c_{rf} \right] [-q_i^A q_j^A + q_i^B q_j^B] \quad (4.125)$$

Note that the constants k_{rf} and c_{rf} are defined using the dielectric constant ε_{rf} of the medium (see sec. 4.1.4).

Lennard-Jones interaction

For the Lennard-Jones interaction between two particles of which the *atom type* varies with λ we can write:

$$V_{LJ} = \frac{(1-\lambda)C_{12}^A + \lambda C_{12}^B}{r_{ij}^{12}} - \frac{(1-\lambda)C_6^A + \lambda C_6^B}{r_{ij}^6} \quad (4.126)$$

$$\frac{\partial V_{LJ}}{\partial \lambda} = \frac{C_{12}^B - C_{12}^A}{r_{ij}^{12}} - \frac{C_6^B - C_6^A}{r_{ij}^6} \quad (4.127)$$

It should be noted that it is also possible to express a pathway from state A to state B using σ and ϵ (see eqn. 4.5). It may seem to make sense physically to vary the force field parameters σ and ϵ rather than the derived parameters C_{12} and C_6 . However, the difference between the pathways in parameter space is not large, and the free energy itself does not depend on the pathway, so we use the simple formulation presented above.

Kinetic Energy

When the mass of a particle changes, there is also a contribution of the kinetic energy to the free energy (note that we can not write the momentum \mathbf{p} as $m\mathbf{v}$, since that would result in the sign of $\frac{\partial E_k}{\partial \lambda}$ being incorrect [104]):

$$E_k = \frac{1}{2} \frac{\mathbf{p}^2}{(1-\lambda)m^A + \lambda m^B} \quad (4.128)$$

$$\frac{\partial E_k}{\partial \lambda} = -\frac{1}{2} \frac{\mathbf{p}^2(m^B - m^A)}{((1-\lambda)m^A + \lambda m^B)^2} \quad (4.129)$$

after taking the derivative, we *can* insert $\mathbf{p} = m\mathbf{v}$, such that:

$$\frac{\partial E_k}{\partial \lambda} = -\frac{1}{2} \mathbf{v}^2 (m^B - m^A) \quad (4.130)$$

Constraints

The constraints are formally part of the Hamiltonian, and therefore they give a contribution to the free energy. In GROMACS this can be calculated using the LINCS or the SHAKE algorithm. If we have $k = 1 \dots K$ constraint equations g_k for LINCS, then

$$g_k = |\mathbf{r}_k| - d_k \quad (4.131)$$

where \mathbf{r}_k is the displacement vector between two particles and d_k is the constraint distance between the two particles. We can express the fact that the constraint distance has a λ dependency by

$$d_k = (1-\lambda)d_k^A + \lambda d_k^B \quad (4.132)$$

Thus the λ -dependent constraint equation is

$$g_k = |\mathbf{r}_k| - \left((1-\lambda)d_k^A + \lambda d_k^B \right). \quad (4.133)$$

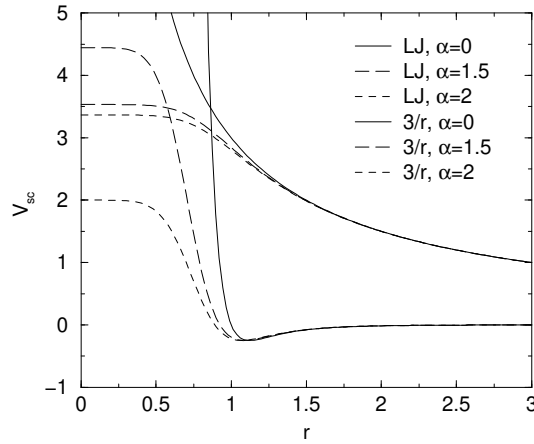


Figure 4.16: Soft-core interactions at $\lambda = 0.5$, with $p = 2$ and $C_6^A = C_{12}^A = C_6^B = C_{12}^B = 1$.

The (zero) contribution G to the Hamiltonian from the constraints (using Lagrange multipliers λ_k , which are logically distinct from the free-energy λ) is

$$G = \sum_k^K \lambda_k g_k \quad (4.134)$$

$$\frac{\partial G}{\partial \lambda} = \frac{\partial G}{\partial d_k} \frac{\partial d_k}{\partial \lambda} \quad (4.135)$$

$$= - \sum_k^K \lambda_k (d_k^B - d_k^A) \quad (4.136)$$

For SHAKE, the constraint equations are

$$g_k = r_k^2 - d_k^2 \quad (4.137)$$

with d_k as before, so

$$\frac{\partial G}{\partial \lambda} = -2 \sum_k^K \lambda_k (d_k^B - d_k^A) \quad (4.138)$$

4.5.1 Soft-core interactions

In a free-energy calculation where particles grow out of nothing, or particles disappear, using the simple linear interpolation of the Lennard-Jones and Coulomb potentials as described in Equations 4.127 and 4.125 may lead to poor convergence. When the particles have nearly disappeared, or are close to appearing (at λ close to 0 or 1), the interaction energy will be weak enough for particles to get very close to each other, leading to large fluctuations in the measured values of $\partial V / \partial \lambda$ (which, because of the simple linear interpolation, depends on the potentials at both the endpoints of λ).

To circumvent these problems, the singularities in the potentials need to be removed. This can be done by modifying the regular Lennard-Jones and Coulomb potentials with “soft-core” potentials that limit the energies and forces involved at λ values between 0 and 1, but not at $\lambda = 0$ or 1.

In GROMACS the soft-core potentials V_{sc} are shifted versions of the regular potentials, so that the singularity in the potential and its derivatives at $r = 0$ is never reached:

$$V_{sc}(r) = (1 - \lambda)V^A(r_A) + \lambda V^B(r_B) \quad (4.139)$$

$$r_A = \left(\alpha \sigma_A^6 \lambda^p + r^6 \right)^{\frac{1}{6}} \quad (4.140)$$

$$r_B = \left(\alpha \sigma_B^6 (1 - \lambda)^p + r^6 \right)^{\frac{1}{6}} \quad (4.141)$$

where V^A and V^B are the normal “hard core” Van der Waals or electrostatic potentials in state A ($\lambda = 0$) and state B ($\lambda = 1$) respectively, α is the soft-core parameter (set with `sc_alpha` in the `.mdp` file), p is the soft-core λ power (set with `sc_power`), σ is the radius of the interaction, which is $(C_{12}/C_6)^{1/6}$ or an input parameter (`sc_sigma`) when C_6 or C_{12} is zero.

For intermediate λ , r_A and r_B alter the interactions very little for $r > \alpha^{1/6}\sigma$ and quickly switch the soft-core interaction to an almost constant value for smaller r (Fig. 4.16). The force is:

$$F_{sc}(r) = -\frac{\partial V_{sc}(r)}{\partial r} = (1 - \lambda)F^A(r_A) \left(\frac{r}{r_A} \right)^5 + \lambda F^B(r_B) \left(\frac{r}{r_B} \right)^5 \quad (4.142)$$

where F^A and F^B are the “hard core” forces. The contribution to the derivative of the free energy is:

$$\begin{aligned} \frac{\partial V_{sc}(r)}{\partial \lambda} &= V^B(r_B) - V^A(r_A) + (1 - \lambda) \frac{\partial V^A(r_A)}{\partial r_A} \frac{\partial r_A}{\partial \lambda} + \lambda \frac{\partial V^B(r_B)}{\partial r_B} \frac{\partial r_B}{\partial \lambda} \\ &= V^B(r_B) - V^A(r_A) + \\ &\quad \frac{p\alpha}{6} \left[\lambda F^B(r_B) r_B^{-5} \sigma_B^6 (1 - \lambda)^{p-1} - (1 - \lambda) F^A(r_A) r_A^{-5} \sigma_A^6 \lambda^{p-1} \right] \end{aligned} \quad (4.143)$$

The original GROMOS Lennard-Jones soft-core function [105] uses $p = 2$, but $p = 1$ gives a smoother $\partial H/\partial \lambda$ curve.

Another issue that should be considered is the soft-core effect of hydrogens without Lennard-Jones interaction. Their soft-core σ is set with `sc-sigma` in the `.mdp` file. These hydrogens produce peaks in $\partial H/\partial \lambda$ at λ is 0 and/or 1 for $p = 1$ and close to 0 and/or 1 with $p = 2$. Lowering `sc-sigma` will decrease this effect, but it will also increase the interactions with hydrogens relative to the other interactions in the soft-core state.

When soft-core potentials are selected (by setting `sc-alpha > 0`), and the Coulomb and Lennard-Jones potentials are turned on or off sequentially, then the Coulombic interaction is turned off linearly, rather than using soft-core interactions, which should be less statistically noisy in most cases. This behavior can be overwritten by using the `mdp` option `sc-coul` to `yes`. Note that the `sc-coul` is only taken into account when `lambda` states are used, not with `couple-lambda0` / `couple-lambda1`, and you can still turn off soft-core interactions by setting `sc-alpha=0`. Additionally, the soft-core interaction potential is only applied when either the A or B state has zero interaction potential. If both A and B states have nonzero interaction potential, default linear scaling described above is used. When both Coulombic and Lennard-Jones interactions are turned

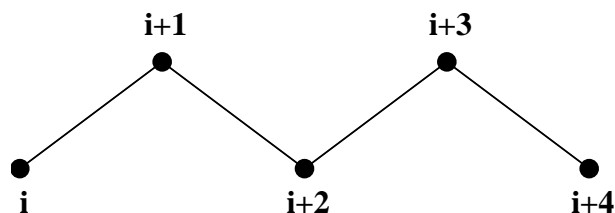


Figure 4.17: Atoms along an alkane chain.

off simultaneously, a soft-core potential is used, and a hydrogen is being introduced or deleted, the sigma is set to `sc-sigma-min`, which itself defaults to `sc-sigma-default`.

Recently, a new formulation of the soft-core approach has been derived that in most cases gives lower and more even statistical variance than the standard soft-core path described above. [106, 107] Specifically, we have:

$$V_{sc}(r) = (1 - \lambda)V^A(r_A) + \lambda V^B(r_B) \quad (4.144)$$

$$r_A = \left(\alpha \sigma_A^{48} \lambda^p + r^{48} \right)^{\frac{1}{48}} \quad (4.145)$$

$$r_B = \left(\alpha \sigma_B^{48} (1 - \lambda)^p + r^{48} \right)^{\frac{1}{48}} \quad (4.146)$$

This “1-1-48” path is also implemented in GROMACS. Note that for this path the soft core α should satisfy $0.001 < \alpha < 0.003$, rather than $\alpha \approx 0.5$.

4.6 Methods

4.6.1 Exclusions and 1-4 Interactions.

Atoms within a molecule that are close by in the chain, *i.e.* atoms that are covalently bonded, or linked by one or two atoms are called *first neighbors*, *second neighbors* and *third neighbors*, respectively (see Fig. 4.17). Since the interactions of atom **i** with atoms **i+1** and **i+2** are mainly quantum mechanical, they can not be modeled by a Lennard-Jones potential. Instead it is assumed that these interactions are adequately modeled by a harmonic bond term or constraint (**i**, **i+1**) and a harmonic angle term (**i**, **i+2**). The first and second neighbors (atoms **i+1** and **i+2**) are therefore *excluded* from the Lennard-Jones interaction list of atom **i**; atoms **i+1** and **i+2** are called *exclusions* of atom **i**.

For third neighbors, the normal Lennard-Jones repulsion is sometimes still too strong, which means that when applied to a molecule, the molecule would deform or break due to the internal strain. This is especially the case for carbon-carbon interactions in a *cis*-conformation (*e.g.* *cis*-butane). Therefore, for some of these interactions, the Lennard-Jones repulsion has been reduced in the GROMOS force field, which is implemented by keeping a separate list of 1-4 and normal Lennard-Jones parameters. In other force fields, such as OPLS [108], the standard Lennard-Jones parameters are reduced by a factor of two, but in that case also the dispersion (r^{-6}) and the Coulomb interaction are scaled. GROMACS can use either of these methods.

4.6.2 Charge Groups

In principle, the force calculation in MD is an $O(N^2)$ problem. Therefore, we apply a cut-off for non-bonded force (NBF) calculations; only the particles within a certain distance of each other are interacting. This reduces the cost to $O(N)$ (typically $100N$ to $200N$) of the NBF. It also introduces an error, which is, in most cases, acceptable, except when applying the cut-off implies the creation of charges, in which case you should consider using the lattice sum methods provided by GROMACS.

Consider a water molecule interacting with another atom. If we would apply a plain cut-off on an atom-atom basis we might include the atom-oxygen interaction (with a charge of -0.82) without the compensating charge of the protons, and as a result, induce a large dipole moment over the system. Therefore, we have to keep groups of atoms with total charge 0 together. These groups are called *charge groups*. Note that with a proper treatment of long-range electrostatics (e.g. particle-mesh Ewald (sec. 4.8.2), keeping charge groups together is not required.

4.6.3 Treatment of Cut-offs in the group scheme

GROMACS is quite flexible in treating cut-offs, which implies there can be quite a number of parameters to set. These parameters are set in the input file for `grompp`. There are two sort of parameters that affect the cut-off interactions; you can select which type of interaction to use in each case, and which cut-offs should be used in the neighbor searching.

For both Coulomb and van der Waals interactions there are interaction type selectors (termed `vdwtype` and `coulombtype`) and two parameters, for a total of six non-bonded interaction parameters. See the User Guide for a complete description of these parameters.

In the group cut-off scheme, all of the interaction functions in Table 4.2 require that neighbor searching be done with a radius at least as large as the r_c specified for the functional form, because of the use of charge groups. The extra radius is typically of the order of 0.25 nm (roughly the largest distance between two atoms in a charge group plus the distance a charge group can diffuse within neighbor list updates).

	Type	Parameters
Coulomb	Plain cut-off	r_c, ϵ_r
	Reaction field	r_c, ϵ_{rf}
	Shift function	r_1, r_c, ϵ_r
	Switch function	r_1, r_c, ϵ_r
VdW	Plain cut-off	r_c
	Shift function	r_1, r_c
	Switch function	r_1, r_c

Table 4.2: Parameters for the different functional forms of the non-bonded interactions.

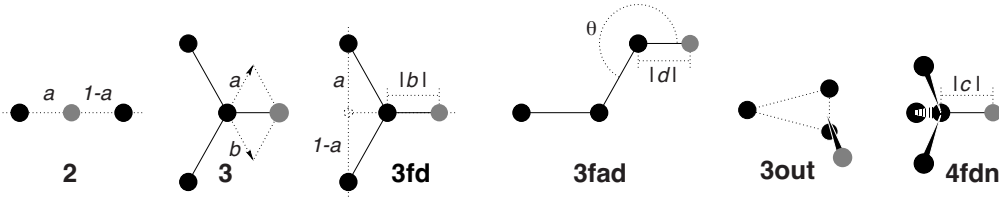


Figure 4.18: The six different types of virtual site construction in GROMACS. The constructing atoms are shown as black circles, the virtual sites in gray.

4.7 Virtual interaction sites

Virtual interaction sites (called dummy atoms in GROMACS versions before 3.3) can be used in GROMACS in a number of ways. We write the position of the virtual site \mathbf{r}_s as a function of the positions of other particles \mathbf{r}_i : $\mathbf{r}_s = f(\mathbf{r}_1, \dots, \mathbf{r}_n)$. The virtual site, which may carry charge or be involved in other interactions, can now be used in the force calculation. The force acting on the virtual site must be redistributed over the particles with mass in a consistent way. A good way to do this can be found in ref. [109]. We can write the potential energy as:

$$V = V(\mathbf{r}_s, \mathbf{r}_1, \dots, \mathbf{r}_n) = V^*(\mathbf{r}_1, \dots, \mathbf{r}_n) \quad (4.147)$$

The force on the particle i is then:

$$\mathbf{F}_i = -\frac{\partial V^*}{\partial \mathbf{r}_i} = -\frac{\partial V}{\partial \mathbf{r}_i} - \frac{\partial V}{\partial \mathbf{r}_s} \frac{\partial \mathbf{r}_s}{\partial \mathbf{r}_i} = \mathbf{F}_i^{direct} + \mathbf{F}'_i \quad (4.148)$$

The first term is the normal force. The second term is the force on particle i due to the virtual site, which can be written in tensor notation:

$$\mathbf{F}'_i = \begin{bmatrix} \frac{\partial x_s}{\partial x_i} & \frac{\partial y_s}{\partial x_i} & \frac{\partial z_s}{\partial x_i} \\ \frac{\partial x_s}{\partial y_i} & \frac{\partial y_s}{\partial y_i} & \frac{\partial z_s}{\partial y_i} \\ \frac{\partial x_s}{\partial z_i} & \frac{\partial y_s}{\partial z_i} & \frac{\partial z_s}{\partial z_i} \end{bmatrix} \mathbf{F}_s \quad (4.149)$$

where \mathbf{F}_s is the force on the virtual site and x_s , y_s and z_s are the coordinates of the virtual site. In this way, the total force and the total torque are conserved [109].

The computation of the virial (eqn. 3.24) is non-trivial when virtual sites are used. Since the virial involves a summation over all the atoms (rather than virtual sites), the forces must be redistributed from the virtual sites to the atoms (using eqn. 4.149) *before* computation of the virial. In some special cases where the forces on the atoms can be written as a linear combination of the forces on the virtual sites (types 2 and 3 below) there is no difference between computing the virial before and after the redistribution of forces. However, in the general case redistribution should be done first.

There are six ways to construct virtual sites from surrounding atoms in GROMACS, which we classify by the number of constructing atoms. **Note** that all site types mentioned can be constructed

from types 3fd (normalized, in-plane) and 3out (non-normalized, out of plane). However, the amount of computation involved increases sharply along this list, so we strongly recommended using the first adequate virtual site type that will be sufficient for a certain purpose. Fig. 4.18 depicts 6 of the available virtual site constructions. The conceptually simplest construction types are linear combinations:

$$\mathbf{r}_s = \sum_{i=1}^N w_i \mathbf{r}_i \quad (4.150)$$

The force is then redistributed using the same weights:

$$\mathbf{F}'_i = w_i \mathbf{F}_s \quad (4.151)$$

The types of virtual sites supported in GROMACS are given in the list below. Constructing atoms in virtual sites can be virtual sites themselves, but only if they are higher in the list, i.e. virtual sites can be constructed from “particles” that are simpler virtual sites.

2. As a linear combination of two atoms (Fig. 4.18 2):

$$w_i = 1 - a, \quad w_j = a \quad (4.152)$$

In this case the virtual site is on the line through atoms i and j .

3. As a linear combination of three atoms (Fig. 4.18 3):

$$w_i = 1 - a - b, \quad w_j = a, \quad w_k = b \quad (4.153)$$

In this case the virtual site is in the plane of the other three particles.

- 3fd. In the plane of three atoms, with a fixed distance (Fig. 4.18 3fd):

$$\mathbf{r}_s = \mathbf{r}_i + b \frac{\mathbf{r}_{ij} + a\mathbf{r}_{jk}}{|\mathbf{r}_{ij} + a\mathbf{r}_{jk}|} \quad (4.154)$$

In this case the virtual site is in the plane of the other three particles at a distance of $|b|$ from i . The force on particles i , j and k due to the force on the virtual site can be computed as:

$$\begin{aligned} \mathbf{F}'_i &= \mathbf{F}_s - \gamma(\mathbf{F}_s - \mathbf{p}) \\ \mathbf{F}'_j &= (1 - a)\gamma(\mathbf{F}_s - \mathbf{p}) \quad \text{where} \quad \gamma = \frac{b}{|\mathbf{r}_{ij} + a\mathbf{r}_{jk}|} \\ \mathbf{F}'_k &= a\gamma(\mathbf{F}_s - \mathbf{p}) \quad \mathbf{p} = \frac{\mathbf{r}_{is} \cdot \mathbf{F}_s}{\mathbf{r}_{is} \cdot \mathbf{r}_{is}} \mathbf{r}_{is} \end{aligned} \quad (4.155)$$

- 3fad. In the plane of three atoms, with a fixed angle and distance (Fig. 4.18 3fad):

$$\mathbf{r}_s = \mathbf{r}_i + d \cos \theta \frac{\mathbf{r}_{ij}}{|\mathbf{r}_{ij}|} + d \sin \theta \frac{\mathbf{r}_\perp}{|\mathbf{r}_\perp|} \quad \text{where} \quad \mathbf{r}_\perp = \mathbf{r}_{jk} - \frac{\mathbf{r}_{ij} \cdot \mathbf{r}_{jk}}{\mathbf{r}_{ij} \cdot \mathbf{r}_{ij}} \mathbf{r}_{ij} \quad (4.156)$$

In this case the virtual site is in the plane of the other three particles at a distance of $|d|$ from i at an angle of α with \mathbf{r}_{ij} . Atom k defines the plane and the direction of the angle. **Note** that in this case b and α must be specified, instead of a and b (see also sec. 5.2.2). The force

on particles i, j and k due to the force on the virtual site can be computed as (with \mathbf{r}_\perp as defined in eqn. 4.156):

$$\begin{aligned} \mathbf{F}'_i &= \mathbf{F}_s - \frac{d \cos \theta}{|\mathbf{r}_{ij}|} \mathbf{F}_1 + \frac{d \sin \theta}{|\mathbf{r}_\perp|} \left(\frac{\mathbf{r}_{ij} \cdot \mathbf{r}_{jk}}{\mathbf{r}_{ij} \cdot \mathbf{r}_{ij}} \mathbf{F}_2 + \mathbf{F}_3 \right) \\ \mathbf{F}'_j &= \frac{d \cos \theta}{|\mathbf{r}_{ij}|} \mathbf{F}_1 - \frac{d \sin \theta}{|\mathbf{r}_\perp|} \left(\mathbf{F}_2 + \frac{\mathbf{r}_{ij} \cdot \mathbf{r}_{jk}}{\mathbf{r}_{ij} \cdot \mathbf{r}_{ij}} \mathbf{F}_2 + \mathbf{F}_3 \right) \\ \mathbf{F}'_k &= \frac{d \sin \theta}{|\mathbf{r}_\perp|} \mathbf{F}_2 \end{aligned}$$

where $\mathbf{F}_1 = \mathbf{F}_s - \frac{\mathbf{r}_{ij} \cdot \mathbf{F}_s}{\mathbf{r}_{ij} \cdot \mathbf{r}_{ij}} \mathbf{r}_{ij}$, $\mathbf{F}_2 = \mathbf{F}_1 - \frac{\mathbf{r}_\perp \cdot \mathbf{F}_s}{\mathbf{r}_\perp \cdot \mathbf{r}_\perp} \mathbf{r}_\perp$ and $\mathbf{F}_3 = \frac{\mathbf{r}_{ij} \cdot \mathbf{F}_s}{\mathbf{r}_{ij} \cdot \mathbf{r}_{ij}} \mathbf{r}_\perp$

(4.157)

3out. As a non-linear combination of three atoms, out of plane (Fig. 4.18 3out):

$$\mathbf{r}_s = \mathbf{r}_i + a\mathbf{r}_{ij} + b\mathbf{r}_{ik} + c(\mathbf{r}_{ij} \times \mathbf{r}_{ik}) \quad (4.158)$$

This enables the construction of virtual sites out of the plane of the other atoms. The force on particles i, j and k due to the force on the virtual site can be computed as:

$$\begin{aligned} \mathbf{F}'_j &= \begin{bmatrix} a & -c z_{ik} & c y_{ik} \\ c z_{ik} & a & -c x_{ik} \\ -c y_{ik} & c x_{ik} & a \end{bmatrix} \mathbf{F}_s \\ \mathbf{F}'_k &= \begin{bmatrix} b & c z_{ij} & -c y_{ij} \\ -c z_{ij} & b & c x_{ij} \\ c y_{ij} & -c x_{ij} & b \end{bmatrix} \mathbf{F}_s \\ \mathbf{F}'_i &= \mathbf{F}_s - \mathbf{F}'_j - \mathbf{F}'_k \end{aligned} \quad (4.159)$$

4fdn. From four atoms, with a fixed distance, see separate Fig. 4.19. This construction is a bit complex, in particular since the previous type (4fd) could be unstable which forced us to introduce a more elaborate construction:

$$\begin{aligned} \mathbf{r}_{ja} &= a \mathbf{r}_{ik} - \mathbf{r}_{ij} = a(\mathbf{x}_k - \mathbf{x}_i) - (\mathbf{x}_j - \mathbf{x}_i) \\ \mathbf{r}_{jb} &= b \mathbf{r}_{il} - \mathbf{r}_{ij} = b(\mathbf{x}_l - \mathbf{x}_i) - (\mathbf{x}_j - \mathbf{x}_i) \\ \mathbf{r}_m &= \mathbf{r}_{ja} \times \mathbf{r}_{jb} \\ \mathbf{x}_s &= \mathbf{x}_i + c \frac{\mathbf{r}_m}{|\mathbf{r}_m|} \end{aligned} \quad (4.160)$$

In this case the virtual site is at a distance of $|c|$ from i , while a and b are parameters. **Note** that the vectors \mathbf{r}_{ik} and \mathbf{r}_{ij} are not normalized to save floating-point operations. The force on particles i, j, k and l due to the force on the virtual site are computed through chain rule derivatives of the construction expression. This is exact and conserves energy, but it does

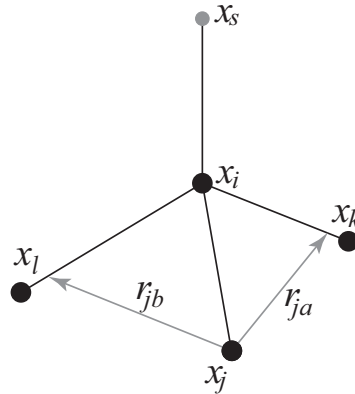


Figure 4.19: The new 4fdn virtual site construction, which is stable even when all constructing atoms are in the same plane.

lead to relatively lengthy expressions that we do not include here (over 200 floating-point operations). The interested reader can look at the source code in `vsite.c`. Fortunately, this `vsite` type is normally only used for chiral centers such as C_α atoms in proteins.

The new 4fdn construct is identified with a ‘type’ value of 2 in the topology. The earlier 4fd type is still supported internally (‘type’ value 1), but it should not be used for new simulations. All current GROMACS tools will automatically generate type 4fdn instead.

N. A linear combination of N atoms with relative weights a_i . The weight for atom i is:

$$w_i = a_i \left(\sum_{j=1}^N a_j \right)^{-1} \quad (4.161)$$

There are three options for setting the weights:

COG center of geometry: equal weights

COM center of mass: a_i is the mass of atom i ; when in free-energy simulations the mass of the atom is changed, only the mass of the A-state is used for the weight

COW center of weights: a_i is defined by the user

4.8 Long Range Electrostatics

4.8.1 Ewald summation

The total electrostatic energy of N particles and their periodic images is given by

$$V = \frac{f}{2} \sum_{n_x} \sum_{n_y} \sum_{n_z^*} \sum_i^N \sum_j^N \frac{q_i q_j}{\mathbf{r}_{ij,\mathbf{n}}}. \quad (4.162)$$

$(n_x, n_y, n_z) = \mathbf{n}$ is the box index vector, and the star indicates that terms with $i = j$ should be omitted when $(n_x, n_y, n_z) = (0, 0, 0)$. The distance $r_{ij,\mathbf{n}}$ is the real distance between the charges and not the minimum-image. This sum is conditionally convergent, but very slow.

Ewald summation was first introduced as a method to calculate long-range interactions of the periodic images in crystals [110]. The idea is to convert the single slowly-converging sum eqn. 4.162 into two quickly-converging terms and a constant term:

$$V = V_{\text{dir}} + V_{\text{rec}} + V_0 \quad (4.163)$$

$$V_{\text{dir}} = \frac{f}{2} \sum_{i,j}^N \sum_{n_x} \sum_{n_y} \sum_{n_z^*} q_i q_j \frac{\text{erfc}(\beta r_{ij,\mathbf{n}})}{r_{ij,\mathbf{n}}} \quad (4.164)$$

$$V_{\text{rec}} = \frac{f}{2\pi V} \sum_{i,j}^N q_i q_j \sum_{m_x} \sum_{m_y} \sum_{m_z^*} \frac{\exp(-(\pi \mathbf{m}/\beta)^2 + 2\pi i \mathbf{m} \cdot (\mathbf{r}_i - \mathbf{r}_j))}{\mathbf{m}^2} \quad (4.165)$$

$$V_0 = -\frac{f\beta}{\sqrt{\pi}} \sum_i^N q_i^2, \quad (4.166)$$

where β is a parameter that determines the relative weight of the direct and reciprocal sums and $\mathbf{m} = (m_x, m_y, m_z)$. In this way we can use a short cut-off (of the order of 1 nm) in the direct space sum and a short cut-off in the reciprocal space sum (e.g. 10 wave vectors in each direction). Unfortunately, the computational cost of the reciprocal part of the sum increases as N^2 (or $N^{3/2}$ with a slightly better algorithm) and it is therefore not realistic for use in large systems.

Using Ewald

Don't use Ewald unless you are absolutely sure this is what you want - for almost all cases the PME method below will perform much better. If you still want to employ classical Ewald summation enter this in your `.mdp` file, if the side of your box is about 3 nm:

```
coulombtype      = Ewald
rvdw             = 0.9
rlist            = 0.9
rcoulomb         = 0.9
fourierspacing   = 0.6
ewald-rtol       = 1e-5
```

The ratio of the box dimensions and the `fourierspacing` parameter determines the highest magnitude of wave vectors m_x, m_y, m_z to use in each direction. With a 3-nm cubic box this example would use 11 wave vectors (from -5 to 5) in each direction. The `ewald-rtol` parameter is the relative strength of the electrostatic interaction at the cut-off. Decreasing this gives you a more accurate direct sum, but a less accurate reciprocal sum.

4.8.2 PME

Particle-mesh Ewald is a method proposed by Tom Darden [14] to improve the performance of the reciprocal sum. Instead of directly summing wave vectors, the charges are assigned to a grid

using interpolation. The implementation in GROMACS uses cardinal B-spline interpolation [15], which is referred to as smooth PME (SPME). The grid is then Fourier transformed with a 3D FFT algorithm and the reciprocal energy term obtained by a single sum over the grid in k-space.

The potential at the grid points is calculated by inverse transformation, and by using the interpolation factors we get the forces on each atom.

The PME algorithm scales as $N \log(N)$, and is substantially faster than ordinary Ewald summation on medium to large systems. On very small systems it might still be better to use Ewald to avoid the overhead in setting up grids and transforms. For the parallelization of PME see the section on MPMD PME (3.17.5).

With the Verlet cut-off scheme, the PME direct space potential is shifted by a constant such that the potential is zero at the cut-off. This shift is small and since the net system charge is close to zero, the total shift is very small, unlike in the case of the Lennard-Jones potential where all shifts add up. We apply the shift anyhow, such that the potential is the exact integral of the force.

Using PME

As an example for using Particle-mesh Ewald summation in GROMACS, specify the following lines in your `.mdp` file:

```
coulombtype      = PME
rvdw             = 0.9
rlist            = 0.9
rcoulomb         = 0.9
fourierspacing  = 0.12
pme-order        = 4
ewald-rtol       = 1e-5
```

In this case the `fourierspacing` parameter determines the maximum spacing for the FFT grid (i.e. minimum number of grid points), and `pme-order` controls the interpolation order. Using fourth-order (cubic) interpolation and this spacing should give electrostatic energies accurate to about $5 \cdot 10^{-3}$. Since the Lennard-Jones energies are not this accurate it might even be possible to increase this spacing slightly.

Pressure scaling works with PME, but be aware of the fact that anisotropic scaling can introduce artificial ordering in some systems.

4.8.3 P3M-AD

The Particle-Particle Particle-Mesh methods of Hockney & Eastwood can also be applied in GROMACS for the treatment of long range electrostatic interactions [111]. Although the P3M method was the first efficient long-range electrostatics method for molecular simulation, the smooth PME (SPME) method has largely replaced P3M as the method of choice in atomistic simulations. One performance disadvantage of the original P3M method was that it required 3 3D-FFT back transforms to obtain the forces on the particles. But this is not required for P3M and the forces can be

derived through analytical differentiation of the potential, as done in PME. The resulting method is termed P3M-AD. The only remaining difference between P3M-AD and PME is the optimization of the lattice Green influence function for error minimization that P3M uses. However, in 2012 it has been shown that the SPME influence function can be modified to obtain P3M [112]. This means that the advantage of error minimization in P3M-AD can be used at the same computational cost and with the same code as PME, just by adding a few lines to modify the influence function. However, at optimal parameter setting the effect of error minimization in P3M-AD is less than 10%. P3M-AD does show large accuracy gains with interlaced (also known as staggered) grids, but that is not supported in GROMACS (yet).

P3M is used in GROMACS with exactly the same options as used with PME by selecting the electrostatics type:

```
coulombtype = P3M-AD
```

4.8.4 Optimizing Fourier transforms and PME calculations

It is recommended to optimize the parameters for calculation of electrostatic interaction such as PME grid dimensions and cut-off radii. This is particularly relevant to do before launching long production runs.

`gmx mdrun` will automatically do a lot of PME optimization, and GROMACS also includes a special tool, `gmx tune_pme`, which automates the process of selecting the optimal number of PME-only ranks.

4.9 Long Range Van der Waals interactions

4.9.1 Dispersion correction

In this section, we derive long-range corrections due to the use of a cut-off for Lennard-Jones or Buckingham interactions. We assume that the cut-off is so long that the repulsion term can safely be neglected, and therefore only the dispersion term is taken into account. Due to the nature of the dispersion interaction (we are truncating a potential proportional to $-r^{-6}$), energy and pressure corrections are both negative. While the energy correction is usually small, it may be important for free energy calculations where differences between two different Hamiltonians are considered. In contrast, the pressure correction is very large and can not be neglected under any circumstances where a correct pressure is required, especially for any NPT simulations. Although it is, in principle, possible to parameterize a force field such that the pressure is close to the desired experimental value without correction, such a method makes the parameterization dependent on the cut-off and is therefore undesirable.

Energy

The long-range contribution of the dispersion interaction to the virial can be derived analytically, if we assume a homogeneous system beyond the cut-off distance r_c . The dispersion energy between

two particles is written as:

$$V(r_{ij}) = -C_6 r_{ij}^{-6} \quad (4.167)$$

and the corresponding force is:

$$\mathbf{F}_{ij} = -6 C_6 r_{ij}^{-8} \mathbf{r}_{ij} \quad (4.168)$$

In a periodic system it is not easy to calculate the full potentials, so usually a cut-off is applied, which can be abrupt or smooth. We will call the potential and force with cut-off V_c and \mathbf{F}_c . The long-range contribution to the dispersion energy in a system with N particles and particle density $\rho = N/V$ is:

$$V_{lr} = \frac{1}{2} N \rho \int_0^\infty 4\pi r^2 g(r) (V(r) - V_c(r)) dr \quad (4.169)$$

We will integrate this for the shift function, which is the most general form of van der Waals interaction available in GROMACS. The shift function has a constant difference S from 0 to r_1 and is 0 beyond the cut-off distance r_c . We can integrate eqn. 4.169, assuming that the density in the sphere within r_1 is equal to the global density and the radial distribution function $g(r)$ is 1 beyond r_1 :

$$\begin{aligned} V_{lr} &= \frac{1}{2} N \left(\rho \int_0^{r_1} 4\pi r^2 g(r) C_6 S dr + \rho \int_{r_1}^{r_c} 4\pi r^2 (V(r) - V_c(r)) dr + \rho \int_{r_c}^\infty 4\pi r^2 V(r) dr \right) \\ &= \frac{1}{2} N \left(\left(\frac{4}{3} \pi \rho r_1^3 - 1 \right) C_6 S + \rho \int_{r_1}^{r_c} 4\pi r^2 (V(r) - V_c(r)) dr - \frac{4}{3} \pi N \rho C_6 r_c^{-3} \right) \quad (4.170) \end{aligned}$$

where the term -1 corrects for the self-interaction. For a plain cut-off we only need to assume that $g(r)$ is 1 beyond r_c and the correction reduces to [113]:

$$V_{lr} = -\frac{2}{3} \pi N \rho C_6 r_c^{-3} \quad (4.171)$$

If we consider, for example, a box of pure water, simulated with a cut-off of 0.9 nm and a density of 1 g cm^{-3} this correction is $-0.75 \text{ kJ mol}^{-1}$ per molecule.

For a homogeneous mixture we need to define an *average dispersion constant*:

$$\langle C_6 \rangle = \frac{2}{N(N-1)} \sum_i^N \sum_{j>i}^N C_6(i, j) \quad (4.172)$$

In GROMACS, excluded pairs of atoms do not contribute to the average.

In the case of inhomogeneous simulation systems, *e.g.* a system with a lipid interface, the energy correction can be applied if $\langle C_6 \rangle$ for both components is comparable.

Virial and pressure

The scalar virial of the system due to the dispersion interaction between two particles i and j is given by:

$$\Xi = -\frac{1}{2} \mathbf{r}_{ij} \cdot \mathbf{F}_{ij} = 3 C_6 r_{ij}^{-6} \quad (4.173)$$

The pressure is given by:

$$P = \frac{2}{3V} (E_{kin} - \Xi) \quad (4.174)$$

The long-range correction to the virial is given by:

$$\Xi_{lr} = \frac{1}{2}N\rho \int_0^\infty 4\pi r^2 g(r)(\Xi - \Xi_c) dr \quad (4.175)$$

We can again integrate the long-range contribution to the virial assuming $g(r)$ is 1 beyond r_1 :

$$\begin{aligned} \Xi_{lr} &= \frac{1}{2}N\rho \left(\int_{r_1}^{r_c} 4\pi r^2 (\Xi - \Xi_c) dr + \int_{r_c}^\infty 4\pi r^2 3 C_6 r_{ij}^{-6} dr \right) \\ &= \frac{1}{2}N\rho \left(\int_{r_1}^{r_c} 4\pi r^2 (\Xi - \Xi_c) dr + 4\pi C_6 r_c^{-3} \right) \end{aligned} \quad (4.176)$$

For a plain cut-off the correction to the pressure is [113]:

$$P_{lr} = -\frac{4}{3}\pi C_6 \rho^2 r_c^{-3} \quad (4.177)$$

Using the same example of a water box, the correction to the virial is 0.75 kJ mol^{-1} per molecule, the corresponding correction to the pressure for SPC water is approximately -280 bar .

For homogeneous mixtures, we can again use the average dispersion constant $\langle C_6 \rangle$ (eqn. 4.172):

$$P_{lr} = -\frac{4}{3}\pi \langle C_6 \rangle \rho^2 r_c^{-3} \quad (4.178)$$

For inhomogeneous systems, eqn. 4.178 can be applied under the same restriction as holds for the energy (see sec. 4.9.1).

4.9.2 Lennard-Jones PME

In order to treat systems, using Lennard-Jones potentials, that are non-homogeneous outside of the cut-off distance, we can instead use the Particle-mesh Ewald method as discussed for electrostatics above. In this case the modified Ewald equations become

$$V = V_{\text{dir}} + V_{\text{rec}} + V_0 \quad (4.179)$$

$$V_{\text{dir}} = -\frac{1}{2} \sum_{i,j}^N \sum_{n_x} \sum_{n_y} \sum_{n_z^*} \frac{C_6^{ij} g(\beta r_{ij,\mathbf{n}})}{r_{ij,\mathbf{n}}^6} \quad (4.180)$$

$$V_{\text{rec}} = \frac{\pi^{\frac{3}{2}} \beta^3}{2V} \sum_{m_x} \sum_{m_y} \sum_{m_z^*} f(\pi|\mathbf{m}|/\beta) \times \sum_{i,j}^N C_6^{ij} \exp[-2\pi i \mathbf{m} \cdot (\mathbf{r}_i - \mathbf{r}_j)] \quad (4.181)$$

$$V_0 = -\frac{\beta^6}{12} \sum_i^N C_6^{ii} \quad (4.182)$$

where $\mathbf{m} = (m_x, m_y, m_z)$, β is the parameter determining the weight between direct and reciprocal space, and C_6^{ij} is the combined dispersion parameter for particle i and j . The star indicates that terms with $i = j$ should be omitted when $((n_x, n_y, n_z) = (0, 0, 0))$, and $\mathbf{r}_{ij,\mathbf{n}}$ is the real distance

between the particles. Following the derivation by Essmann [15], the functions f and g introduced above are defined as

$$f(x) = 1/3 \left[(1 - 2x^2)\exp(-x^2) + 2x^3\sqrt{\pi} \operatorname{erfc}(x) \right] \quad (4.183)$$

$$g(x) = \exp(-x^2)\left(1 + x^2 + \frac{x^4}{2}\right). \quad (4.184)$$

The above methodology works fine as long as the dispersion parameters can be combined geometrically (eqn. 4.6) in the same way as the charges for electrostatics

$$C_{6,\text{geom}}^{ij} = \left(C_6^{ii} C_6^{jj} \right)^{1/2} \quad (4.185)$$

For Lorentz-Berthelot combination rules (eqn. 4.7), the reciprocal part of this sum has to be calculated seven times due to the splitting of the dispersion parameter according to

$$C_{6,\text{L-B}}^{ij} = (\sigma_i + \sigma_j)^6 = \sum_{n=0}^6 P_n \sigma_i^n \sigma_j^{(6-n)}, \quad (4.186)$$

for P_n the Pascal triangle coefficients. This introduces a non-negligible cost to the reciprocal part, requiring seven separate FFTs, and therefore this has been the limiting factor in previous attempts to implement LJ-PME. A solution to this problem is to use geometrical combination rules in order to calculate an approximate interaction parameter for the reciprocal part of the potential, yielding a total interaction of

$$\begin{aligned} V(r < r_c) &= \underbrace{C_6^{\text{dir}} g(\beta r) r^{-6}}_{\text{Direct space}} + \underbrace{C_{6,\text{geom}}^{\text{recip}} [1 - g(\beta r)] r^{-6}}_{\text{Reciprocal space}} \\ &= C_{6,\text{geom}}^{\text{recip}} r^{-6} + \left(C_6^{\text{dir}} - C_{6,\text{geom}}^{\text{recip}} \right) g(\beta r) r^{-6} \end{aligned} \quad (4.187)$$

$$V(r > r_c) = \underbrace{C_{6,\text{geom}}^{\text{recip}} [1 - g(\beta r)] r^{-6}}_{\text{Reciprocal space}}. \quad (4.188)$$

This will preserve a well-defined Hamiltonian and significantly increase the performance of the simulations. The approximation does introduce some errors, but since the difference is located in the interactions calculated in reciprocal space, the effect will be very small compared to the total interaction energy. In a simulation of a lipid bilayer, using a cut-off of 1.0 nm, the relative error in total dispersion energy was below 0.5%. A more thorough discussion of this can be found in [114].

In GROMACS we now perform the proper calculation of this interaction by subtracting, from the direct-space interactions, the contribution made by the approximate potential that is used in the reciprocal part

$$V_{\text{dir}} = C_6^{\text{dir}} r^{-6} - C_{6,\text{geom}}^{\text{recip}} [1 - g(\beta r)] r^{-6}. \quad (4.189)$$

This potential will reduce to the expression in eqn. 4.180 when $C_6^{\text{dir}} = C_{6,\text{geom}}^{\text{recip}}$, and the total interaction is given by

$$\begin{aligned} V(r < r_c) &= \underbrace{C_6^{\text{dir}} r^{-6} - C_{6,\text{geom}}^{\text{recip}} [1 - g(\beta r)] r^{-6}}_{\text{Direct space}} + \underbrace{C_{6,\text{geom}}^{\text{recip}} [1 - g(\beta r)] r^{-6}}_{\text{Reciprocal space}} \\ &= C_6^{\text{dir}} r^{-6} \end{aligned} \quad (4.190)$$

$$V(r > r_c) = C_{6,\text{geom}}^{\text{recip}} [1 - g(\beta r)] r^{-6}. \quad (4.191)$$

For the case when $C_6^{\text{dir}} \neq C_6^{\text{recip}}$ this will retain an unmodified LJ force up to the cut-off, and the error is an order of magnitude smaller than in simulations where the direct-space interactions do not account for the approximation used in reciprocal space. When using a VdW interaction modifier of potential-shift, the constant

$$\left(-C_6^{\text{dir}} + C_6^{\text{recip}}[1 - g(\beta r_c)]\right) r_c^{-6} \quad (4.192)$$

is added to eqn. 4.190 in order to ensure that the potential is continuous at the cutoff. Note that, in the same way as eqn. 4.189, this degenerates into the expected $-C_6 g(\beta r_c) r_c^{-6}$ when $C_6^{\text{dir}} = C_6^{\text{recip}}$. In addition to this, a long-range dispersion correction can be applied to correct for the approximation using a combination rule in reciprocal space. This correction assumes, as for the cut-off LJ potential, a uniform particle distribution. But since the error of the combination rule approximation is very small this long-range correction is not necessary in most cases. Also note that this homogenous correction does not correct the surface tension, which is an inhomogeneous property.

Using LJ-PME

As an example for using Particle-mesh Ewald summation for Lennard-Jones interactions in GRO-MACS, specify the following lines in your `.mdp` file:

```
vdwtype           = PME
rvdw              = 0.9
vdw-modifier      = Potential-Shift
rlist             = 0.9
rcoulomb          = 0.9
fourierspacing   = 0.12
pme-order         = 4
ewald-rtol-lj    = 0.001
lj-pme-comb-rule = geometric
```

The same Fourier grid and interpolation order are used if both LJ-PME and electrostatic PME are active, so the settings for `fourierspacing` and `pme-order` are common to both. `ewald-rtol-lj` controls the splitting between direct and reciprocal space in the same way as `ewald-rtol`. In addition to this, the combination rule to be used in reciprocal space is determined by `lj-pme-comb-rule`. If the current force field uses Lorentz-Berthelot combination rules, it is possible to set `lj-pme-comb-rule = geometric` in order to gain a significant increase in performance for a small loss in accuracy. The details of this approximation can be found in the section above.

Note that the use of a complete long-range dispersion correction means that as with Coulomb PME, `rvdw` is now a free parameter in the method, rather than being necessarily restricted by the force-field parameterization scheme. Thus it is now possible to optimize the cutoff, spacing, order and tolerance terms for accuracy and best performance.

Naturally, the use of LJ-PME rather than LJ cut-off adds computation and communication done for the reciprocal-space part, so for best performance in balancing the load of parallel simulations using PME-only ranks, more such ranks should be used. It may be possible to improve upon the automatic load-balancing used by `mdrun`.

4.10 Force field

A force field is built up from two distinct components:

- The set of equations (called the *potential functions*) used to generate the potential energies and their derivatives, the forces. These are described in detail in the previous chapter.
- The parameters used in this set of equations. These are not given in this manual, but in the data files corresponding to your GROMACS distribution.

Within one set of equations various sets of parameters can be used. Care must be taken that the combination of equations and parameters form a consistent set. It is in general dangerous to make *ad hoc* changes in a subset of parameters, because the various contributions to the total force are usually interdependent. This means in principle that every change should be documented, verified by comparison to experimental data and published in a peer-reviewed journal before it can be used.

GROMACS 2018.4 includes several force fields, and additional ones are available on the website. If you do not know which one to select we recommend GROMOS-96 for united-atom setups and OPLS-AA/L for all-atom parameters. That said, we describe the available options in some detail.

All-hydrogen force field

The GROMOS-87-based all-hydrogen force field is almost identical to the normal GROMOS-87 force field, since the extra hydrogens have no Lennard-Jones interaction and zero charge. The only differences are in the bond angle and improper dihedral angle terms. This force field is only useful when you need the exact hydrogen positions, for instance for distance restraints derived from NMR measurements. When citing this force field please read the previous paragraph.

4.10.1 GROMOS-96

GROMACS supports the GROMOS-96 force fields [82]. All parameters for the 43A1, 43A2 (development, improved alkane dihedrals), 45A3, 53A5, and 53A6 parameter sets are included. All standard building blocks are included and topologies can be built automatically by `pdb2gmx`.

The GROMOS-96 force field is a further development of the GROMOS-87 force field. It has improvements over the GROMOS-87 force field for proteins and small molecules. **Note** that the sugar parameters present in 53A6 do correspond to those published in 2004[115], which are different from those present in 45A4, which is not included in GROMACS at this time. The 45A4 parameter set corresponds to a later revision of these parameters. The GROMOS-96 force field is not, however, recommended for use with long alkanes and lipids. The GROMOS-96 force field differs from the GROMOS-87 force field in a few respects:

- the force field parameters
- the parameters for the bonded interactions are not linked to atom types
- a fourth power bond stretching potential (4.2.1)

- an angle potential based on the cosine of the angle (4.2.6)

There are two differences in implementation between GROMACS and GROMOS-96 which can lead to slightly different results when simulating the same system with both packages:

- in GROMOS-96 neighbor searching for solvents is performed on the first atom of the solvent molecule. This is not implemented in GROMACS, but the difference with searching by centers of charge groups is very small
- the virial in GROMOS-96 is molecule-based. This is not implemented in GROMACS, which uses atomic virials

The GROMOS-96 force field was parameterized with a Lennard-Jones cut-off of 1.4 nm, so be sure to use a Lennard-Jones cut-off (`rvdw`) of at least 1.4. A larger cut-off is possible because the Lennard-Jones potential and forces are almost zero beyond 1.4 nm.

GROMOS-96 files

GROMACS can read and write GROMOS-96 coordinate and trajectory files. These files should have the extension `.g96`. Such a file can be a GROMOS-96 initial/final configuration file, a coordinate trajectory file, or a combination of both. The file is fixed format; all floats are written as 15.9, and as such, files can get huge. GROMACS supports the following data blocks in the given order:

- Header block:

```
TITLE (mandatory)
```

- Frame blocks:

```
TIMESTEP (optional)  
POSITION/POSITIONRED (mandatory)  
VELOCITY/VELOCITYRED (optional)  
BOX (optional)
```

See the GROMOS-96 manual [82] for a complete description of the blocks. **Note** that all GROMACS programs can read compressed (`.Z`) or gzipped (`.gz`) files.

4.10.2 OPLS/AA

4.10.3 AMBER

GROMACS provides native support for the following AMBER force fields:

- AMBER94 [116]

- AMBER96 [117]
- AMBER99 [118]
- AMBER99SB [119]
- AMBER99SB-ILDN [120]
- AMBER03 [121]
- AMBERGS [122]

4.10.4 CHARMM

GROMACS supports the CHARMM force field for proteins [123, 124], lipids [125] and nucleic acids [126, 127]. The protein parameters (and to some extent the lipid and nucleic acid parameters) were thoroughly tested – both by comparing potential energies between the port and the standard parameter set in the CHARMM molecular simulation package, as well by how the protein force field behaves together with GROMACS-specific techniques such as virtual sites (enabling long time steps) and a fast implicit solvent recently implemented [74] – and the details and results are presented in the paper by Bjelkmar et al. [128]. The nucleic acid parameters, as well as the ones for HEME, were converted and tested by Michel Cuendet.

When selecting the CHARMM force field in `pdb2gmx` the default option is to use CMAP (for torsional correction map). To exclude CMAP, use `-nocmap`. The basic form of the CMAP term implemented in GROMACS is a function of the ϕ and ψ backbone torsion angles. This term is defined in the `.rtp` file by a `[cmap]` statement at the end of each residue supporting CMAP. The following five atom names define the two torsional angles. Atoms 1-4 define ϕ , and atoms 2-5 define ψ . The corresponding atom types are then matched to the correct CMAP type in the `cmap.itp` file that contains the correction maps.

A port of the CHARMM36 force field for use with GROMACS is also available at http://mackerell.umaryland.edu/charmm_ff.shtml#gromacs.

For branched polymers or other topologies not supported by `pdb2gmx`, it is possible to use TopoTools [129] to generate a GROMACS top file.

4.10.5 Coarse-grained force fields

Coarse-graining is a systematic way of reducing the number of degrees of freedom representing a system of interest. To achieve this, typically whole groups of atoms are represented by single beads and the coarse-grained force fields describes their effective interactions. Depending on the choice of parameterization, the functional form of such an interaction can be complicated and often tabulated potentials are used.

Coarse-grained models are designed to reproduce certain properties of a reference system. This can be either a full atomistic model or even experimental data. Depending on the properties to reproduce there are different methods to derive such force fields. An incomplete list of methods is given below:

- Conserving free energies
 - Simplex method
 - MARTINI force field (see next section)
- Conserving distributions (like the radial distribution function), so-called structure-based coarse-graining
 - (iterative) Boltzmann inversion
 - Inverse Monte Carlo
- Conserving forces
 - Force matching

Note that coarse-grained potentials are state dependent (e.g. temperature, density,...) and should be re-parametrized depending on the system of interest and the simulation conditions. This can for example be done using the Versatile Object-oriented Toolkit for Coarse-Graining Applications (VOTCA) [130]. The package was designed to assist in systematic coarse-graining, provides implementations for most of the algorithms mentioned above and has a well tested interface to GROMACS. It is available as open source and further information can be found at www.votca.org.

4.10.6 MARTINI

The MARTINI force field is a coarse-grain parameter set that allows for the construction of many systems, including proteins and membranes.

4.10.7 PLUM

The PLUM force field [131] is an example of a solvent-free protein-membrane model for which the membrane was derived from structure-based coarse-graining [132]. A GROMACS implementation can be found at code.google.com/p/plumx.

Chapter 5

Topologies

5.1 Introduction

GROMACS must know on which atoms and combinations of atoms the various contributions to the potential functions (see chapter 4) must act. It must also know what parameters must be applied to the various functions. All this is described in the *topology* file `*.top`, which lists the *constant attributes* of each atom. There are many more atom types than elements, but only atom types present in biological systems are parameterized in the force field, plus some metals, ions and silicon. The bonded and special interactions are determined by fixed lists that are included in the topology file. Certain non-bonded interactions must be excluded (first and second neighbors), as these are already treated in bonded interactions. In addition, there are *dynamic attributes* of atoms - their positions, velocities and forces. These do not strictly belong to the molecular topology, and are stored in the coordinate file `*.gro` (positions and velocities), or trajectory file `*.trr` (positions, velocities, forces).

This chapter describes the setup of the topology file, the `*.top` file and the database files: what the parameters stand for and how/where to change them if needed. First, all file formats are explained. Section 5.9.1 describes the organization of the files in each force field.

Note: if you construct your own topologies, we encourage you to upload them to our topology archive at www.gromacs.org! Just imagine how thankful you'd have been if your topology had been available there before you started. The same goes for new force fields or modified versions of the standard force fields - contribute them to the force field archive!

5.2 Particle type

In GROMACS, there are three types of particles, see Table 5.1. Only regular atoms and virtual interaction sites are used in GROMACS; shells are necessary for polarizable models like the Shell-Water models [45].

Particle	Symbol
atoms	A
shells	S
virtual interaction sites	V (or D)

Table 5.1: Particle types in GROMACS

5.2.1 Atom types

Each force field defines a set of atom types, which have a characteristic name or number, and mass (in a.m.u.). These listings are found in the `atomtypes.atp` file (`.atp` = **atom type parameter** file). Therefore, it is in this file that you can begin to change and/or add an atom type. A sample from the `gromos43a1.ff` force field is listed below.

```

O 15.99940 ; carbonyl oxygen (C=O)
OM 15.99940 ; carboxyl oxygen (CO-)
OA 15.99940 ; hydroxyl, sugar or ester oxygen
OW 15.99940 ; water oxygen
N 14.00670 ; peptide nitrogen (N or NH)
NT 14.00670 ; terminal nitrogen (NH2)
NL 14.00670 ; terminal nitrogen (NH3)
NR 14.00670 ; aromatic nitrogen
NZ 14.00670 ; Arg NH (NH2)
NE 14.00670 ; Arg NE (NH)
C 12.01100 ; bare carbon
CH1 13.01900 ; aliphatic or sugar CH-group
CH2 14.02700 ; aliphatic or sugar CH2-group
CH3 15.03500 ; aliphatic CH3-group

```

Note: GROMACS makes use of the atom types as a name, *not* as a number (as *e.g.* in GROMOS).

5.2.2 Virtual sites

Some force fields use virtual interaction sites (interaction sites that are constructed from other particle positions) on which certain interactions are located (*e.g.* on benzene rings, to reproduce the correct quadrupole). This is described in sec. 4.7.

To make virtual sites in your system, you should include a section `[virtual_sites?]` (for backward compatibility the old name `[dummies?]` can also be used) in your topology file, where the ‘?’ stands for the number constructing particles for the virtual site. This will be ‘2’ for type 2, ‘3’ for types 3, 3fd, 3fad and 3out and ‘4’ for type 4fdn. The last of these replace an older 4fd type (with the ‘type’ value 1) that could occasionally be unstable; while it is still supported internally in the code, the old 4fd type should not be used in new input files. The different types are explained in sec. 4.7.

Parameters for type 2 should look like this:

```
[ virtual_sites2 ]
```



```
; Site from      funct a
5      1      2      1      0.7439756
```

for type 3 like this:

```
[ virtual_sites3 ]
; Site from      funct a      b
5      1      2      3      1      0.7439756  0.128012
```

for type 3fd like this:

```
[ virtual_sites3 ]
; Site from      funct a      d
5      1      2      3      2      0.5      -0.105
```

for type 3fad like this:

```
[ virtual_sites3 ]
; Site from      funct theta  d
5      1      2      3      3      120      0.5
```

for type 3out like this:

```
[ virtual_sites3 ]
; Site from      funct a      b      c
5      1      2      3      4      -0.4      -0.4      6.9281
```

for type 4fdn like this:

```
[ virtual_sites4 ]
; Site from      funct a      b      c
5      1      2      3      4      2      1.0      0.9      0.105
```

This will result in the construction of a virtual site, number 5 (first column ‘Site’), based on the positions of the atoms whose indices are 1 and 2 or 1, 2 and 3 or 1, 2, 3 and 4 (next two, three or four columns ‘from’) following the rules determined by the function number (next column ‘funct’) with the parameters specified (last one, two or three columns ‘a b . .’). Obviously, the atom numbers (including virtual site number) depend on the molecule. It may be instructive to study the topologies for TIP4P or TIP5P water models that are included with the GROMACS distribution.

Note that if any constant bonded interactions are defined between virtual sites and/or normal atoms, they will be removed by `grompp` (unless the option `tt -normvsbds` is used). This removal of bonded interactions is done after generating exclusions, as the generation of exclusions is based on “chemically” bonded interactions.

Virtual sites can be constructed in a more generic way using basic geometric parameters. The directive that can be used is `[virtual_sitesn]`. Required parameters are listed in Table 5.5. An example entry for defining a virtual site at the center of geometry of a given set of atoms might be:

```
[ virtual_sitesn ]
; Site  funct  from
5      1      1      2      3      4
```

Property	Symbol	Unit
Type	-	-
Mass	m	a.m.u.
Charge	q	electron
epsilon	ϵ	kJ/mol
sigma	σ	nm

Table 5.2: Static atom type properties in GROMACS

5.3 Parameter files

5.3.1 Atoms

The *static* properties (see Table 5.2) assigned to the atom types are assigned based on data in several places. The mass is listed in `atomtypes.atp` (see 5.2.1), whereas the charge is listed in `*.rtp` (`.rtp` = **r**esidue **t**opology **p**arameter file, see 5.7.1). This implies that the charges are only defined in the building blocks of amino acids, nucleic acids or otherwise, as defined by the user. When generating a topology (`*.top`) using the `pdb2gmx` program, the information from these files is combined.

5.3.2 Non-bonded parameters

The non-bonded parameters consist of the van der Waals parameters V (`c6` or σ , depending on the combination rule) and W (`c12` or ϵ), as listed in the file `ffnonbonded.itp`, where `pctype` is the particle type (see Table 5.1). As with the bonded parameters, entries in `[*type]` directives are applied to their counterparts in the topology file. Missing parameters generate warnings, except as noted below in section 5.4.3.

```
[ atomtypes ]
;name  at.num      mass      charge  ptype      V(c6)      W(c12)
  O      8  15.99940    0.000    A    0.22617E-02  0.74158E-06
  OM     8  15.99940    0.000    A    0.22617E-02  0.74158E-06
  .....

[ nonbond_params ]
; i    j  func      V(c6)      W(c12)
  O    O    1  0.22617E-02  0.74158E-06
  O    OA   1  0.22617E-02  0.13807E-05
  .....
```

Note that most of the included force fields also include the `at.num.` column, but this same information is implied in the OPLS-AA `bond_type` column. The interpretation of the parameters V and W depends on the combination rule that was chosen in the `[defaults]` section of the topology file (see 5.8.1):

$$\text{for combination rule 1 : } \begin{aligned} V_{ii} &= C_i^{(6)} = 4 \epsilon_i \sigma_i^6 \quad [\text{kJ mol}^{-1} \text{ nm}^6] \\ W_{ii} &= C_i^{(12)} = 4 \epsilon_i \sigma_i^{12} \quad [\text{kJ mol}^{-1} \text{ nm}^{12}] \end{aligned} \quad (5.1)$$

$$\begin{aligned} \text{for combination rules 2 and 3 :} \quad V_{ii} &= \sigma_i \text{ [nm]} \\ W_{ii} &= \epsilon_i \text{ [kJ mol}^{-1} \text{]} \end{aligned} \quad (5.2)$$

Some or all combinations for different atom types can be given in the [nonbond_params] section, again with parameters V and W as defined above. Any combination that is not given will be computed from the parameters for the corresponding atom types, according to the combination rule:

$$\begin{aligned} \text{for combination rules 1 and 3 :} \quad C_{ij}^{(6)} &= \left(C_i^{(6)} C_j^{(6)} \right)^{\frac{1}{2}} \\ C_{ij}^{(12)} &= \left(C_i^{(12)} C_j^{(12)} \right)^{\frac{1}{2}} \end{aligned} \quad (5.3)$$

$$\begin{aligned} \text{for combination rule 2 :} \quad \sigma_{ij} &= \frac{1}{2}(\sigma_i + \sigma_j) \\ \epsilon_{ij} &= \sqrt{\epsilon_i \epsilon_j} \end{aligned} \quad (5.4)$$

When σ and ϵ need to be supplied (rules 2 and 3), it would seem it is impossible to have a non-zero C^{12} combined with a zero C^6 parameter. However, providing a negative σ will do exactly that, such that C^6 is set to zero and C^{12} is calculated normally. This situation represents a special case in reading the value of σ , and nothing more.

There is only one set of combination rules for Buckingham potentials:

$$\begin{aligned} A_{ij} &= (A_{ii} A_{jj})^{1/2} \\ B_{ij} &= 2 / \left(\frac{1}{B_{ii}} + \frac{1}{B_{jj}} \right) \\ C_{ij} &= (C_{ii} C_{jj})^{1/2} \end{aligned} \quad (5.5)$$

5.3.3 Bonded parameters

The bonded parameters (*i.e.* bonds, bond angles, improper and proper dihedrals) are listed in `ffbonded.itp`. The entries in this database describe, respectively, the atom types in the interactions, the type of the interaction, and the parameters associated with that interaction. These parameters are then read by `grompp` when processing a topology and applied to the relevant bonded parameters, *i.e.* `bondtypes` are applied to entries in the [bonds] directive, etc. Any bonded parameter that is missing from the relevant [*type] directive generates a fatal error. The types of interactions are listed in Table 5.5. Example excerpts from such files follow:

```
[ bondtypes ]
; i      j func          b0          kb
  C      O      1      0.12300      502080.
  C      OM     1      0.12500      418400.
  .....

[ angletypes ]
; i      j      k func          th0          cth
  HO     OA     C      1      109.500      397.480
  HO     OA     CH1    1      109.500      397.480
  .....

[ dihedraltypes ]
```

```

; i      l func          q0          cq
NR5*  NR5      2      0.000      167.360
NR5*  NR5*     2      0.000      167.360
.....

[ dihedraltypes ]
; j      k func          phi0          cp      mult
  C     OA      1     180.000          16.736      2
  C     N       1     180.000          33.472      2
.....

[ dihedraltypes ]
;
; Ryckaert-Bellemans Dihedrals
;
; aj      ak      funct
CP2      CP2      3          9.2789  12.156  -13.120  -3.0597  26.240  -31.495

```

In the `ffbonded.itp` file, you can add bonded parameters. If you want to include parameters for new atom types, make sure you define them in `atomtypes.atp` as well.

For most interaction types, bonded parameters are searched and assigned using an exact match for all type names and allowing only a single set of parameters. The exception to this rule are dihedral parameters. For `[dihedraltypes]` wildcard atom type names can be specified with the letter `X` in one or more of the four positions. Thus one can for example assign proper dihedral parameters based on the types of the middle two atoms. The parameters for the entry with the most exact matches, i.e. the least wildcard matches, will be used. Note that GROMACS versions older than 5.1.3 used the first match, which means that a full match would be ignored if it is preceded by an entry that matches on wildcards. Thus it is suggested to put wildcard entries at the end, in case someone might use a forcefield with older versions of GROMACS. In addition there is a dihedral type 9 which adds the possibility of assigning multiple dihedral potentials, useful for combining terms with different multiplicities. The different dihedral potential parameter sets should be on directly adjacent lines in the `[dihedraltypes]` section.

5.4 Molecule definition

5.4.1 Moleculetype entries

An organizational structure that usually corresponds to molecules is the `[moleculetype]` entry. This entry serves two main purposes. One is to give structure to the topology file(s), usually corresponding to real molecules. This makes the topology easier to read and writing it less labor intensive. A second purpose is computational efficiency. The system definition that is kept in memory is proportional in size of the `moleculetype` definitions. If a molecule is present in 100000 copies, this saves a factor of 100000 in memory, which means the system usually fits in cache, which can improve performance tremendously. Interactions that correspond to chemical bonds, that generate exclusions, can only be defined between atoms within a `moleculetype`. It is allowed to have multiple molecules which are not covalently bonded in one `moleculetype`

definition. Molecules can be made infinitely long by connecting to themselves over periodic boundaries. When such periodic molecules are present, an option in the `mdp` file needs to be set to tell GROMACS not to attempt to make molecules that are broken over periodic boundaries whole again.

5.4.2 Intermolecular interactions

In some cases, one would like atoms in different molecules to also interact with other interactions than the usual non-bonded interactions. This is often the case in binding studies. When the molecules are covalently bound, e.g. a ligand binding covalently to a protein, they are effectively one molecule and they should be defined in one `[moleculetype]` entry. Note that `pdb2gmx` has an option to put two or more molecules in one `[moleculetype]` entry. When molecules are not covalently bound, it is much more convenient to use separate `moleculetype` definitions and specify the intermolecular interactions in the `[intermolecular_interactions]` section. In this section, which is placed at the end of the topology (see Table 5.4), normal bonded interactions can be specified using global atom indices. The only restrictions are that no interactions can be used that generates exclusions and no constraints can be used.

5.4.3 Intramolecular pair interactions

Extra Lennard-Jones and electrostatic interactions between pairs of atoms in a molecule can be added in the `[pairs]` section of a molecule definition. The parameters for these interactions can be set independently from the non-bonded interaction parameters. In the GROMOS force fields, pairs are only used to modify the 1-4 interactions (interactions of atoms separated by three bonds). In these force fields the 1-4 interactions are excluded from the non-bonded interactions (see sec. 5.4.4).

```
[ pairtypes ]
; i      j func          cs6          cs12 ; THESE ARE 1-4 INTERACTIONS
  O      O      1 0.22617E-02  0.74158E-06
  O      OM     1 0.22617E-02  0.74158E-06
  . . . . .
```

The pair interaction parameters for the atom types in `ffnonbonded.itp` are listed in the `[pairtypes]` section. The GROMOS force fields list all these interaction parameters explicitly, but this section might be empty for force fields like OPLS that calculate the 1-4 interactions by uniformly scaling the parameters. Pair parameters that are not present in the `[pairtypes]` section are only generated when `gen-pairs` is set to “yes” in the `[defaults]` directive of `forcefield.itp` (see 5.8.1). When `gen-pairs` is set to “no,” `grompp` will give a warning for each pair type for which no parameters are given.

The normal pair interactions, intended for 1-4 interactions, have function type 1. Function type 2 and the `[pairs_nb]` are intended for free-energy simulations. When determining hydration free energies, the solute needs to be decoupled from the solvent. This can be done by adding a B-state topology (see sec. 3.12) that uses zero for all solute non-bonded parameters, *i.e.* charges

and LJ parameters. However, the free energy difference between the A and B states is not the total hydration free energy. One has to add the free energy for reintroducing the internal Coulomb and LJ interactions in the solute when in vacuum. This second step can be combined with the first step when the Coulomb and LJ interactions within the solute are not modified. For this purpose, there is a pairs function type 2, which is identical to function type 1, except that the B-state parameters are always identical to the A-state parameters. For searching the parameters in the [pairtypes] section, no distinction is made between function type 1 and 2. The pairs section [pairs_nb] is intended to replace the non-bonded interaction. It uses the unscaled charges and the non-bonded LJ parameters; it also only uses the A-state parameters. **Note** that one should add exclusions for all atom pairs listed in [pairs_nb], otherwise such pairs will also end up in the normal neighbor lists.

Alternatively, this same behavior can be achieved without ever touching the topology, by using the `couple-moltype`, `couple-lambda0`, `couple-lambda1`, and `couple-intramol` keywords. See sections [sec. 3.12](#) and [sec. 6.1](#) for more information.

All three pair types always use plain Coulomb interactions, even when Reaction-field, PME, Ewald or shifted Coulomb interactions are selected for the non-bonded interactions. Energies for types 1 and 2 are written to the energy and log file in separate “LJ-14” and “Coulomb-14” entries per energy group pair. Energies for [pairs_nb] are added to the “LJ-(SR)” and “Coulomb-(SR)” terms.

5.4.4 Exclusions

The exclusions for non-bonded interactions are generated by `grompp` for neighboring atoms up to a certain number of bonds away, as defined in the [moleculetype] section in the topology file (see [5.8.1](#)). Particles are considered bonded when they are connected by “chemical” bonds ([bonds] types 1 to 5, 7 or 8) or constraints ([constraints] type 1). Type 5 [bonds] can be used to create a connection between two atoms without creating an interaction. There is a harmonic interaction ([bonds] type 6) that does not connect the atoms by a chemical bond. There is also a second constraint type ([constraints] type 2) that fixes the distance, but does not connect the atoms by a chemical bond. For a complete list of all these interactions, see [Table 5.5](#).

Extra exclusions within a molecule can be added manually in a [exclusions] section. Each line should start with one atom index, followed by one or more atom indices. All non-bonded interactions between the first atom and the other atoms will be excluded.

When all non-bonded interactions within or between groups of atoms need to be excluded, is it more convenient and much more efficient to use energy monitor group exclusions (see [sec. 3.3](#)).

5.5 Implicit solvation parameters

Starting with GROMACS 4.5, implicit solvent is supported. A section in the topology has been introduced to list those parameters:

```
[ implicit_genborn_params ]
```

```

; Atomtype  sar      st      pi      gbr      hct
NH1         0.155    1      1.028   0.17063  0.79 ; N
N           0.155    1      1       0.155    0.79 ; Proline backbone N
H           0.1      1      1       0.115    0.85 ; H
CT1        0.180    1      1.276   0.190    0.72 ; C

```

In this example the atom type is listed first, followed by five numbers, and a comment (following a semicolon).

Values in columns 1-3 are not currently used. They pertain to more elaborate surface area algorithms, the one from Qiu *et al.* [71] in particular. Column 4 contains the atomic van der Waals radii, which are used in computing the Born radii. The dielectric offset is specified in the `*.mdp` file, and gets subtracted from the input van der Waals radii for the different Born radii methods, as described by Onufriev *et al.* [73]. Column 5 is the scale factor for the HCT and OBC models. The values are taken from the Tinker implementation of the HCT pairwise scaling method [72]. This method has been modified such that the scaling factors have been adjusted to minimize differences between analytical surface areas and GB using the HCT algorithm. The scaling is further modified in that it is not applied pairwise as proposed by Hawkins *et al.* [72], but on a per-atom (rather than a per-pair) basis.

5.6 Constraint algorithms

Constraints are defined in the `[constraints]` section. The format is two atom numbers followed by the function type, which can be 1 or 2, and the constraint distance. The only difference between the two types is that type 1 is used for generating exclusions and type 2 is not (see sec. 5.4.4). The distances are constrained using the LINCS or the SHAKE algorithm, which can be selected in the `*.mdp` file. Both types of constraints can be perturbed in free-energy calculations by adding a second constraint distance (see 5.8.5). Several types of bonds and angles (see Table 5.5) can be converted automatically to constraints by `grompp`. There are several options for this in the `*.mdp` file.

We have also implemented the SETTLE algorithm [47], which is an analytical solution of SHAKE, specifically for water. SETTLE can be selected in the topology file. See, for instance, the SPC molecule definition:

```

[ moleculetype ]
; molname      nrexcl
SOL           1

[ atoms ]
; nr      at type res nr  ren nm  at nm  cg nr  charge
1         OW      1      SOL   OW1    1      -0.82
2         HW      1      SOL   HW2    1      0.41
3         HW      1      SOL   HW3    1      0.41

[ settles ]
; OW      funct  doh      dhh
1         1      0.1     0.16333

```

```
[ exclusions ]
1      2      3
2      1      3
3      1      2
```

The `[settles]` directive defines the first atom of the water molecule. The `settle` funct is always 1, and the distance between O-H and H-H distances must be given. **Note** that the algorithm can also be used for TIP3P and TIP4P [133]. TIP3P just has another geometry. TIP4P has a virtual site, but since that is generated it does not need to be shaken (nor stirred).

5.7 pdb2gmx input files

The GROMACS program `pdb2gmx` generates a topology for the input coordinate file. Several formats are supported for that coordinate file, but `*.pdb` is the most commonly-used format (hence the name `pdb2gmx`). `pdb2gmx` searches for force fields in sub-directories of the GROMACS `share/top` directory and your working directory. Force fields are recognized from the file `forcefield.itp` in a directory with the extension `.ff`. The file `forcefield.doc` may be present, and if so, its first line will be used by `pdb2gmx` to present a short description to the user to help in choosing a force field. Otherwise, the user can choose a force field with the `-ff xxx` command-line argument to `pdb2gmx`, which indicates that a force field in a `xxx.ff` directory is desired. `pdb2gmx` will search first in the working directory, then in the GROMACS `share/top` directory, and use the first matching `xxx.ff` directory found.

Two general files are read by `pdb2gmx`: an atom type file (extension `.atp`, see 5.2.1) from the force-field directory, and a file called `residuetypes.dat` from either the working directory, or the GROMACS `share/top` directory. `residuetypes.dat` determines which residue names are considered protein, DNA, RNA, water, and ions.

`pdb2gmx` can read one or multiple databases with topological information for different types of molecules. A set of files belonging to one database should have the same basename, preferably telling something about the type of molecules (*e.g.* aminoacids, rna, dna). The possible files are:

- `<basename>.rtp`
- `<basename>.r2b` (optional)
- `<basename>.arn` (optional)
- `<basename>.hdb` (optional)
- `<basename>.n.tdb` (optional)
- `<basename>.c.tdb` (optional)

Only the `.rtp` file, which contains the topologies of the building blocks, is mandatory. Information from other files will only be used for building blocks that come from an `.rtp` file with the same base name. The user can add building blocks to a force field by having additional files

with the same base name in their working directory. By default, only extra building blocks can be defined, but calling `pdb2gmx` with the `-rtpo` option will allow building blocks in a local file to replace the default ones in the force field.

5.7.1 Residue database

The files holding the residue databases have the extension `.rtp`. Originally this file contained building blocks (amino acids) for proteins, and is the GROMACS interpretation of the `rt37c4.dat` file of GROMOS. So the residue database file contains information (bonds, charges, charge groups, and improper dihedrals) for a frequently-used building block. It is better *not* to change this file because it is standard input for `pdb2gmx`, but if changes are needed make them in the `*.top` file (see 5.8.1), or in a `.rtp` file in the working directory as explained in sec. 5.7. Defining topologies of new small molecules is probably easier by writing an include topology file `*.itp` directly. This will be discussed in section 5.8.2. When adding a new protein residue to the database, don't forget to add the residue name to the `residuetypes.dat` file, so that `grompp`, `make_ndx` and analysis tools can recognize the residue as a protein residue (see 8.1.1).

The `.rtp` files are only used by `pdb2gmx`. As mentioned before, the only extra information this program needs from the `.rtp` database is bonds, charges of atoms, charge groups, and improper dihedrals, because the rest is read from the coordinate input file. Some proteins contain residues that are not standard, but are listed in the coordinate file. You have to construct a building block for this "strange" residue, otherwise you will not obtain a `*.top` file. This also holds for molecules in the coordinate file such as ligands, polyatomic ions, crystallization co-solvents, etc. The residue database is constructed in the following way:

```
[ bondedtypes ] ; mandatory
; bonds angles dihedrals impropers
    1      1      1      2 ; mandatory

[ GLY ] ; mandatory

[ atoms ] ; mandatory
; name type charge chargegroup
    N    N  -0.280    0
    H    H   0.280    0
    CA   CH2  0.000    1
    C    C   0.380    2
    O    O  -0.380    2

[ bonds ] ; optional
;atom1 atom2      b0      kb
    N      H
    N      CA
    CA     C
    C      O
    -C     N

[ exclusions ] ; optional
;atom1 atom2
```

```

[ angles ] ; optional
;atom1 atom2 atom3 th0 cth

[ dihedrals ] ; optional
;atom1 atom2 atom3 atom4 phi0 cp mult

[ impropers ] ; optional
;atom1 atom2 atom3 atom4 q0 cq
  N -C CA H
  -C -CA N -O

[ ZN ]

[ atoms ]
  ZN ZN 2.000 0

```

The file is free format; the only restriction is that there can be at most one entry on a line. The first field in the file is the `[bondedtypes]` field, which is followed by four numbers, indicating the interaction type for bonds, angles, dihedrals, and improper dihedrals. The file contains residue entries, which consist of atoms and (optionally) bonds, angles, dihedrals, and impropers. The charge group codes denote the charge group numbers. Atoms in the same charge group should always be ordered consecutively. When using the hydrogen database with `pdb2gmx` for adding missing hydrogens (see 5.7.4), the atom names defined in the `.rtp` entry should correspond exactly to the naming convention used in the hydrogen database. The atom names in the bonded interaction can be preceded by a minus or a plus, indicating that the atom is in the preceding or following residue respectively. Explicit parameters added to bonds, angles, dihedrals, and impropers override the standard parameters in the `.itp` files. This should only be used in special cases. Instead of parameters, a string can be added for each bonded interaction. This is used in GROMOS-96 `.rtp` files. These strings are copied to the topology file and can be replaced by force-field parameters by the C-preprocessor in `grompp` using `#define` statements.

`pdb2gmx` automatically generates all angles. This means that for most force fields the `[angles]` field is only useful for overriding `.itp` parameters. For the GROMOS-96 force field the interaction number of all angles needs to be specified.

`pdb2gmx` automatically generates one proper dihedral for every rotatable bond, preferably on heavy atoms. When the `[dihedrals]` field is used, no other dihedrals will be generated for the bonds corresponding to the specified dihedrals. It is possible to put more than one dihedral function on a rotatable bond. In the case of CHARMM27 FF `pdb2gmx` can add correction maps to the dihedrals using the default `-cmap` option. Please refer to 4.10.4 for more information.

`pdb2gmx` sets the number of exclusions to 3, which means that interactions between atoms connected by at most 3 bonds are excluded. Pair interactions are generated for all pairs of atoms that are separated by 3 bonds (except pairs of hydrogens). When more interactions need to be excluded, or some pair interactions should not be generated, an `[exclusions]` field can be added, followed by pairs of atom names on separate lines. All non-bonded and pair interactions between these atoms will be excluded.

ARG	protonated arginine
ARGN	neutral arginine
ASP	negatively charged aspartic acid
ASPH	neutral aspartic acid
CYS	neutral cysteine
CYS2	cysteine with sulfur bound to another cysteine or a heme
GLU	negatively charged glutamic acid
GLUH	neutral glutamic acid
HISD	neutral histidine with N _δ protonated
HISE	neutral histidine with N _ε protonated
HISH	positive histidine with both N _δ and N _ε protonated
HIS1	histidine bound to a heme
LYSN	neutral lysine
LYS	protonated lysine
HEME	heme

Table 5.3: Internal GROMACS residue naming convention.

5.7.2 Residue to building block database

Each force field has its own naming convention for residues. Most residues have consistent naming, but some, especially those with different protonation states, can have many different names. The `.r2b` files are used to convert standard residue names to the force-field build block names. If no `.r2b` is present in the force-field directory or a residue is not listed, the building block name is assumed to be identical to the residue name. The `.r2b` can contain 2 or 5 columns. The 2-column format has the residue name in the first column and the building block name in the second. The 5-column format has 3 additional columns with the building block for the residue occurring in the N-terminus, C-terminus and both termini at the same time (single residue molecule). This is useful for, for instance, the AMBER force fields. If one or more of the terminal versions are not present, a dash should be entered in the corresponding column.

There is a GROMACS naming convention for residues which is only apparent (except for the `pdb2gmx` code) through the `.r2b` file and `specbond.dat` files. This convention is only of importance when you are adding residue types to an `.rtp` file. The convention is listed in Table 5.3. For special bonds with, for instance, a heme group, the GROMACS naming convention is introduced through `specbond.dat` (see 5.7.7), which can subsequently be translated by the `.r2b` file, if required.

5.7.3 Atom renaming database

Force fields often use atom names that do not follow IUPAC or PDB convention. The `.arn` database is used to translate the atom names in the coordinate file to the force-field names. Atoms that are not listed keep their names. The file has three columns: the building block name, the old atom name, and the new atom name, respectively. The residue name supports question-mark wildcards that match a single character.

An additional general atom renaming file called `xlateat.dat` is present in the `share/top` directory, which translates common non-standard atom names in the coordinate file to IUPAC/PDB convention. Thus, when writing force-field files, you can assume standard atom names and no further atom name translation is required, except for translating from standard atom names to the force-field ones.

5.7.4 Hydrogen database

The hydrogen database is stored in `.hdb` files. It contains information for the `pdb2gmx` program on how to connect hydrogen atoms to existing atoms. In versions of the database before GRO-MACS 3.3, hydrogen atoms were named after the atom they are connected to: the first letter of the atom name was replaced by an 'H.' In the versions from 3.3 onwards, the H atom has to be listed explicitly, because the old behavior was protein-specific and hence could not be generalized to other molecules. If more than one hydrogen atom is connected to the same atom, a number will be added to the end of the hydrogen atom name. For example, adding two hydrogen atoms to ND2 (in asparagine), the hydrogen atoms will be named HD21 and HD22. This is important since atom naming in the `.rtp` file (see 5.7.1) must be the same. The format of the hydrogen database is as follows:

```
; res      # additions
           # H add type      H      i      j      k
ALA        1
           1      1      H      N      -C      CA
ARG        4
           1      2      H      N      CA      C
           1      1      HE     NE     CD     CZ
           2      3      HH1    NH1    CZ     NE
           2      3      HH2    NH2    CZ     NE
```

On the first line we see the residue name (ALA or ARG) and the number of kinds of hydrogen atoms that may be added to this residue by the hydrogen database. After that follows one line for each addition, on which we see:

- The number of H atoms added
- The method for adding H atoms, which can be any of:
 - 1 *one planar hydrogen, e.g. rings or peptide bond*
One hydrogen atom (n) is generated, lying in the plane of atoms (i,j,k) on the plane bisecting angle (j-i-k) at a distance of 0.1 nm from atom i, such that the angles (n-i-j) and (n-i-k) are $> 90^\circ$.
 - 2 *one single hydrogen, e.g. hydroxyl*
One hydrogen atom (n) is generated at a distance of 0.1 nm from atom i, such that angle (n-i-j)=109.5 degrees and dihedral (n-i-j-k)=trans.
 - 3 *two planar hydrogens, e.g. ethylene -C=CH₂, or amide -C(=O)NH₂*
Two hydrogens (n1,n2) are generated at a distance of 0.1 nm from atom i, such that

angle (n1-i-j)=(n2-i-j)=120 degrees and dihedral (n1-i-j-k)=cis and (n2-i-j-k)=trans, such that names are according to IUPAC standards [134].

4 *two or three tetrahedral hydrogens*, e.g. $-CH_3$

Three (n1,n2,n3) or two (n1,n2) hydrogens are generated at a distance of 0.1 nm from atom i, such that angle (n1-i-j)=(n2-i-j)=(n3-i-j)=109.47°, dihedral (n1-i-j-k)=trans, (n2-i-j-k)=trans+120 and (n3-i-j-k)=trans+240°.

5 *one tetrahedral hydrogen*, e.g. C_3CH

One hydrogen atom (n') is generated at a distance of 0.1 nm from atom i in tetrahedral conformation such that angle (n'-i-j)=(n'-i-k)=(n'-i-l)=109.47°.

6 *two tetrahedral hydrogens*, e.g. $C-CH_2-C$

Two hydrogen atoms (n1,n2) are generated at a distance of 0.1 nm from atom i in tetrahedral conformation on the plane bisecting angle j-i-k with angle (n1-i-n2)=(n1-i-j)=(n1-i-k)=109.47°.

7 *two water hydrogens*

Two hydrogens are generated around atom i according to SPC [85] water geometry. The symmetry axis will alternate between three coordinate axes in both directions.

10 *three water "hydrogens"*

Two hydrogens are generated around atom i according to SPC [85] water geometry. The symmetry axis will alternate between three coordinate axes in both directions. In addition, an extra particle is generated on the position of the oxygen with the first letter of the name replaced by 'M'. This is for use with four-atom water models such as TIP4P [133].

11 *four water "hydrogens"*

Same as above, except that two additional particles are generated on the position of the oxygen, with names 'LP1' and 'LP2.' This is for use with five-atom water models such as TIP5P [135].

- The name of the new H atom (or its prefix, e.g. HD2 for the asparagine example given earlier).
- Three or four control atoms (i,j,k,l), where the first always is the atom to which the H atoms are connected. The other two or three depend on the code selected. For water, there is only one control atom.

Some more exotic cases can be approximately constructed from the above tools, and with suitable use of energy minimization are good enough for beginning MD simulations. For example secondary amine hydrogen, nitrenyl hydrogen (C=NH) and even ethynyl hydrogen could be approximately constructed using method 2 above for hydroxyl hydrogen.

5.7.5 Termini database

The termini databases are stored in `aminoacids.n.tdb` and `aminoacids.c.tdb` for the N- and C-termini respectively. They contain information for the `pdb2gmx` program on how to connect new atoms to existing ones, which atoms should be removed or changed, and which bonded interactions should be added. Their format is as follows (from `gromos43a1.ff/aminoacids.c.tdb`):

```
[ None ]

[ COO- ]
[ replace ]
C C C 12.011 0.27
O O1 OM 15.9994 -0.635
OXT O2 OM 15.9994 -0.635
[ add ]
2 8 O C CA N
OM 15.9994 -0.635
[ bonds ]
C O1 gb_5
C O2 gb_5
[ angles ]
O1 C O2 ga_37
CA C O1 ga_21
CA C O2 ga_21
[ dihedrals ]
N CA C O2 gd_20
[ impropers ]
C CA O2 O1 gi_1
```

The file is organized in blocks, each with a header specifying the name of the block. These blocks correspond to different types of termini that can be added to a molecule. In this example [COO-] is the first block, corresponding to changing the terminal carbon atom into a deprotonated carboxyl group. [None] is the second terminus type, corresponding to a terminus that leaves the molecule as it is. Block names cannot be any of the following: `replace`, `add`, `delete`, `bonds`, `angles`, `dihedrals`, `impropers`. Doing so would interfere with the parameters of the block, and would probably also be very confusing to human readers.

For each block the following options are present:

- [`replace`]
Replace an existing atom by one with a different atom type, atom name, charge, and/or mass. This entry can be used to replace an atom that is present both in the input coordinates and in the `.rtp` database, but also to only rename an atom in the input coordinates such that it matches the name in the force field. In the latter case, there should also be a corresponding [`add`] section present that gives instructions to add the same atom, such that the position in the sequence and the bonding is known. Such an atom can be present in the input coordinates and kept, or not present and constructed by `pdb2gmx`. For each atom to be replaced on line should be entered with the following fields:
 - name of the atom to be replaced
 - new atom name (optional)
 - new atom type
 - new mass
 - new charge

- [`add`]
Add new atoms. For each (group of) added atom(s), a two-line entry is necessary. The first line contains the same fields as an entry in the hydrogen database (name of the new atom, number of atoms, type of addition, control atoms, see 5.7.4), but the possible types of addition are extended by two more, specifically for C-terminal additions:

8 *two carboxyl oxygens, -COO⁻*

Two oxygens (n1,n2) are generated according to rule 3, at a distance of 0.136 nm from atom i and an angle (n1-i-j)=(n2-i-j)=117 degrees

9 *carboxyl oxygens and hydrogen, -COOH*

Two oxygens (n1,n2) are generated according to rule 3, at distances of 0.123 nm and 0.125 nm from atom i for n1 and n2, respectively, and angles (n1-i-j)=121 and (n2-i-j)=115 degrees. One hydrogen (n') is generated around n2 according to rule 2, where n-i-j and n-i-j-k should be read as n'-n2-i and n'-n2-i-j, respectively.

After this line, another line follows that specifies the details of the added atom(s), in the same way as for replacing atoms, *i.e.*:

- atom type
- mass
- charge
- charge group (optional)

Like in the hydrogen database (see 5.7.1), when more than one atom is connected to an existing one, a number will be appended to the end of the atom name. **Note** that, like in the hydrogen database, the atom name is now on the same line as the control atoms, whereas it was at the beginning of the second line prior to GROMACS version 3.3. When the charge group field is left out, the added atom will have the same charge group number as the atom that it is bonded to.

- [`delete`]
Delete existing atoms. One atom name per line.
- [`bonds`], [`angles`], [`dihedrals`] and [`impropers`]
Add additional bonded parameters. The format is identical to that used in the `*.rtp` file, see 5.7.1.

5.7.6 Virtual site database

Since we cannot rely on the positions of hydrogens in input files, we need a special input file to decide the geometries and parameters with which to add virtual site hydrogens. For more complex virtual site constructs (*e.g.* when entire aromatic side chains are made rigid) we also need information about the equilibrium bond lengths and angles for all atoms in the side chain. This information is specified in the `.vsd` file for each force field. Just as for the termini, there is one such file for each class of residues in the `.rtp` file.

The virtual site database is not really a very simple list of information. The first couple of sections specify which mass centers (typically called MCH₃/MNH₃) to use for CH₃, NH₃, and NH₂ groups.

Depending on the equilibrium bond lengths and angles between the hydrogens and heavy atoms we need to apply slightly different constraint distances between these mass centers. **Note** that we do *not* have to specify the actual parameters (that is automatic), just the type of mass center to use. To accomplish this, there are three sections names [CH3], [NH3], and [NH2]. For each of these we expect three columns. The first column is the atom type bound to the 2/3 hydrogens, the second column is the next heavy atom type which this is bound, and the third column the type of mass center to use. As a special case, in the [NH2] section it is also possible to specify `planar` in the second column, which will use a different construction without mass center. There are currently different opinions in some force fields whether an NH₂ group should be planar or not, but we try hard to stick to the default equilibrium parameters of the force field.

The second part of the virtual site database contains explicit equilibrium bond lengths and angles for pairs/triplets of atoms in aromatic side chains. These entries are currently read by specific routines in the virtual site generation code, so if you would like to extend it *e.g.* to nucleic acids you would also need to write new code there. These sections are named after the short amino acid names ([PHE], [TYR], [TRP], [HID], [HIE], [HIP]), and simply contain 2 or 3 columns with atom names, followed by a number specifying the bond length (in nm) or angle (in degrees). **Note** that these are approximations of the equilibrated geometry for the entire molecule, which might not be identical to the equilibrium value for a single bond/angle if the molecule is strained.

5.7.7 Special bonds

The primary mechanism used by `pdb2gmx` to generate inter-residue bonds relies on head-to-tail linking of backbone atoms in different residues to build a macromolecule. In some cases (*e.g.* disulfide bonds, a heme group, branched polymers), it is necessary to create inter-residue bonds that do not lie on the backbone. The file `specbond.dat` takes care of this function. It is necessary that the residues belong to the same [`moleculetype`]. The `-merge` and `-chainsep` functions of `pdb2gmx` can be useful when managing special inter-residue bonds between different chains.

The first line of `specbond.dat` indicates the number of entries that are in the file. If you add a new entry, be sure to increment this number. The remaining lines in the file provide the specifications for creating bonds. The format of the lines is as follows:

```
resA atomA nbondsA resB atomB nbondsB length newresA newresB
```

The columns indicate:

1. `resA` The name of residue A that participates in the bond.
2. `atomA` The name of the atom in residue A that forms the bond.
3. `nbondsA` The total number of bonds `atomA` can form.
4. `resB` The name of residue B that participates in the bond.
5. `atomB` The name of the atom in residue B that forms the bond.
6. `nbondsB` The total number of bonds `atomB` can form.

7. `length` The reference length for the bond. If `atomA` and `atomB` are not within `length` $\pm 10\%$ in the coordinate file supplied to `pdb2gmx`, no bond will be formed.
8. `newresA` The new name of residue A, if necessary. Some force fields use *e.g.* CYS2 for a cysteine in a disulfide or heme linkage.
9. `newresB` The new name of residue B, likewise.

5.8 File formats

5.8.1 Topology file

The topology file is built following the GROMACS specification for a molecular topology. A `*.top` file can be generated by `pdb2gmx`. All possible entries in the topology file are listed in Tables 5.4 and 5.5. Also tabulated are: all the units of the parameters, which interactions can be perturbed for free energy calculations, which bonded interactions are used by `grompp` for generating exclusions, and which bonded interactions can be converted to constraints by `grompp`.

Parameters

interaction type	directive	# at.	f. tp	parameters	F. E.
<i>mandatory</i>	defaults			non-bonded function type; combination rule ^(cr) ; generate pairs (no/yes); fudge LJ (); fudge QQ ()	
<i>mandatory</i>	atomtypes			atom type; m (u); q (e); particle type; $V^{(cr)}$; $W^{(cr)}$	
	bondtypes			(see Table 5.5, directive bonds)	
	pairtypes			(see Table 5.5, directive pairs)	
	angletypes			(see Table 5.5, directive angles)	
	dihedraltypes ^(*)			(see Table 5.5, directive dihedrals)	
	constrainttypes			(see Table 5.5, directive constraints)	
LJ	nonbond_params	2	1	$V^{(cr)}$; $W^{(cr)}$	
Buckingham	nonbond_params	2	2	a (kJ mol ⁻¹); b (nm ⁻¹); c_6 (kJ mol ⁻¹ nm ⁶)	

Molecule definition(s)

<i>mandatory</i>	moleculetype			molecule name; $n_{ex}^{(nrexcl)}$	
<i>mandatory</i>	atoms	1		atom type; residue number; residue name; atom name; charge group number; q (e); m (u)	type q, m
intra-molecular interaction and geometry definitions as described in Table 5.5					

System

<i>mandatory</i>	system			system name	
<i>mandatory</i>	molecules			molecule name; number of molecules	

Inter-molecular interactions

<i>optional</i>	intermolecular_interactions			one or more bonded interactions as described in Table 5.5, with two or more atoms, no interactions that generate exclusions, no constraints, use global atom numbers	
-----------------	-----------------------------	--	--	---	--

'# at' is the required number of atom type indices for this directive

'f. tp' is the value used to select this function type

'F. E.' indicates which of the parameters can be interpolated in free energy calculations

^(cr) the combination rule determines the type of LJ parameters, see 5.3.2

^(*) for dihedraltypes one can specify 4 atoms or the inner (outer for improper) 2 atoms

^(nrexcl) exclude neighbors n_{ex} bonds away for non-bonded interactions

For free energy calculations, type, q and m or no parameters should be added

for topology 'B' ($\lambda = 1$) on the same line, after the normal parameters.

Table 5.4: The topology (*.top) file.

Table 5.5: Details of [moleculetype] directives

Name of interaction	Topology file directive	num. atoms*	func. type [†]	Order of parameters and their units	use in F.E.? [‡]	Cross-references
bond	bonds ^{§,¶}	2	1	b_0 (nm); k_b (kJ mol ⁻¹ nm ⁻²)	all	4.2.1
G96 bond	bonds ^{§,¶}	2	2	b_0 (nm); k_b (kJ mol ⁻¹ nm ⁻⁴)	all	4.2.1
Morse	bonds ^{§,¶}	2	3	b_0 (nm); D (kJ mol ⁻¹); β (nm ⁻¹)	all	4.2.2
cubic bond	bonds ^{§,¶}	2	4	b_0 (nm); $C_{i=2,3}$ (kJ mol ⁻¹ nm ⁻ⁱ)		4.2.3
connection	bonds [§]	2	5			5.4.4
harmonic potential	bonds	2	6	b_0 (nm); k_b (kJ mol ⁻¹ nm ⁻²)	all	4.2.1,5.4.4
FENE bond	bonds [§]	2	7	b_m (nm); k_b (kJ mol ⁻¹ nm ⁻²)		4.2.4
tabulated bond	bonds [§]	2	8	table number (≥ 0); k (kJ mol ⁻¹)	k	4.2.14
tabulated bond	bonds	2	9	table number (≥ 0); k (kJ mol ⁻¹)	k	4.2.14,5.4.4
restraint potential	bonds	2	10	low, up ₁ , up ₂ (nm); k_{dr} (kJ mol ⁻¹ nm ⁻²)	all	4.3.5
extra LJ or Coulomb	pairs	2	1	V^{**} ; W^{**}	all	5.4.3
extra LJ or Coulomb	pairs	2	2	fudge QQ (); q_i, q_j (e), V^{**} ; W^{**}		5.4.3
extra LJ or Coulomb	pairs_nb	2	1	q_i, q_j (e); V^{**} ; W^{**}		5.4.3
angle	angles [¶]	3	1	θ_0 (deg); k_θ (kJ mol ⁻¹ rad ⁻²)	all	4.2.5
G96 angle	angles [¶]	3	2	θ_0 (deg); k_θ (kJ mol ⁻¹)	all	4.2.6
cross bond-bond	angles	3	3	r_{1e}, r_{2e} (nm); $k_{rr'}$ (kJ mol ⁻¹ nm ⁻²)		4.2.9
cross bond-angle	angles	3	4	r_{1e}, r_{2e}, r_{3e} (nm); $k_{r\theta}$ (kJ mol ⁻¹ nm ⁻²)		4.2.10
Urey-Bradley	angles [¶]	3	5	θ_0 (deg); k_θ (kJ mol ⁻¹ rad ⁻²); r_{13} (nm); k_{UB} (kJ mol ⁻¹ nm ⁻²)	all	4.2.8
quartic angle	angles [¶]	3	6	θ_0 (deg); $C_{i=0,1,2,3,4}$ (kJ mol ⁻¹ rad ⁻ⁱ)		4.2.11

*The required number of atom indices for this directive

[†]The index to use to select this function type

[‡]Indicates which of the parameters can be interpolated in free energy calculations

[§]This interaction type will be used by `grompp` for generating exclusions

[¶]This interaction type can be converted to constraints by `grompp`

**The combination rule determines the type of LJ parameters, see 5.3.2

^{||}No connection, and so no exclusions, are generated for this interaction

Table 5.5: Details of [moleculetype] directives

Name of interaction	Topology file directive	num. atoms*	func. type [†]	Order of parameters and their units	use in F.E.? [‡]	Cross-references
tabulated angle	angles	3	8	table number (≥ 0); k (kJ mol ⁻¹)	k	4.2.14
restricted bending potential	angles	3	10	θ_0 (deg); k_θ (kJ mol ⁻¹)		4.2.7
proper dihedral	dihedrals	4	1	ϕ_s (deg); k_ϕ (kJ mol ⁻¹); multiplicity	ϕ, k	4.2.13
improper dihedral	dihedrals	4	2	ξ_0 (deg); k_ξ (kJ mol ⁻¹ rad ⁻²)	all	4.2.12
Ryckaert-Bellemans dihedral	dihedrals	4	3	$C_0, C_1, C_2, C_3, C_4, C_5$ (kJ mol ⁻¹)	all	4.2.13
periodic improper dihedral	dihedrals	4	4	ϕ_s (deg); k_ϕ (kJ mol ⁻¹); multiplicity	ϕ, k	4.2.12
Fourier dihedral	dihedrals	4	5	C_1, C_2, C_3, C_4 (kJ mol ⁻¹)	all	4.2.13
tabulated dihedral	dihedrals	4	8	table number (≥ 0); k (kJ mol ⁻¹)	k	4.2.14
proper dihedral (multiple)	dihedrals	4	9	ϕ_s (deg); k_ϕ (kJ mol ⁻¹); multiplicity	ϕ, k	4.2.13
restricted dihedral	dihedrals	4	10	ϕ_0 (deg); k_ϕ (kJ mol ⁻¹)		4.2.13
combined bending-torsion potential	dihedrals	4	11	a_0, a_1, a_2, a_3, a_4 (kJ mol ⁻¹)		4.2.13
exclusions	exclusions	1		one or more atom indices		5.4.4
constraint	constraints [§]	2	1	b_0 (nm)	all	4.5,5.6
constraint	constraints	2	2	b_0 (nm)	all	4.5,5.6,5.4.4
SETTLE	settles	1	1	d_{OH}, d_{HH} (nm)		3.6.1,5.6
2-body virtual site	virtual_sites2	3	1	a ()		4.7
3-body virtual site	virtual_sites3	4	1	a, b ()		4.7
3-body virtual site (fd)	virtual_sites3	4	2	a (); d (nm)		4.7
3-body virtual site (fad)	virtual_sites3	4	3	θ (deg); d (nm)		4.7
3-body virtual site (out)	virtual_sites3	4	4	a, b (); c (nm ⁻¹)		4.7
4-body virtual site (fdn)	virtual_sites4	5	2	a, b (); c (nm)		4.7
N-body virtual site (COG)	virtual_sitesn	1	1	one or more constructing atom indices		4.7
N-body virtual site (COM)	virtual_sitesn	1	2	one or more constructing atom indices		4.7
N-body virtual site (COW)	virtual_sitesn	1	3	one or more pairs consisting of constructing atom index and weight		4.7
position restraint	position_restraints	1	1	k_x, k_y, k_z (kJ mol ⁻¹ nm ⁻²)	all	4.3.1

Table 5.5: Details of [moleculetype] directives

Name of interaction	Topology file directive	num. atoms*	func. type [†]	Order of parameters and their units	use in F.E.? [‡]	Cross-references
flat-bottomed position restraint	position_restraints	1	2	g, r (nm), k (kJ mol ⁻¹ nm ⁻²)		4.3.2
distance restraint	distance_restraints	2	1	type; label; low, up ₁ , up ₂ (nm); weight ()		4.3.5
dihedral restraint	dihedral_restraints	4	1	ϕ_0 (deg); $\Delta\phi$ (deg); k_{dih} (kJ mol ⁻¹ rad ⁻²)	all	4.3.4
orientation restraint	orientation_restraints	2	1	exp.; label; α ; c (U nm ^{α}); obs. (U); weight (U ⁻¹)		4.3.6
angle restraint	angle_restraints	4	1	θ_0 (deg); k_c (kJ mol ⁻¹); multiplicity	θ, k	4.3.3
angle restraint (z)	angle_restraints_z	2	1	θ_0 (deg); k_c (kJ mol ⁻¹); multiplicity	θ, k	4.3.3

Description of the file layout:

- Semicolon (;) and newline characters surround comments
- On a line ending with \ the newline character is ignored.
- Directives are surrounded by [and]
- The topology hierarchy (which must be followed) consists of three levels:
 - the parameter level, which defines certain force-field specifications (see Table 5.4)
 - the molecule level, which should contain one or more molecule definitions (see Table 5.5)
 - the system level, containing only system-specific information ([system] and [molecules])
- Items should be separated by spaces or tabs, not commas
- Atoms in molecules should be numbered consecutively starting at 1
- Atoms in the same charge group must be listed consecutively
- The file is parsed only once, which implies that no forward references can be treated: items must be defined before they can be used
- Exclusions can be generated from the bonds or overridden manually
- The bonded force types can be generated from the atom types or overridden per bond
- It is possible to apply multiple bonded interactions of the same type on the same atoms
- Descriptive comment lines and empty lines are highly recommended
- Starting with GROMACS version 3.1.3, all directives at the parameter level can be used multiple times and there are no restrictions on the order, except that an atom type needs to be defined before it can be used in other parameter definitions
- If parameters for a certain interaction are defined multiple times for the same combination of atom types the last definition is used; starting with GROMACS version 3.1.3 grompp generates a warning for parameter redefinitions with different values
- Using one of the [atoms], [bonds], [pairs], [angles], etc. without having used [moleculetype] before is meaningless and generates a warning
- Using [molecules] without having used [system] before is meaningless and generates a warning.
- After [system] the only allowed directive is [molecules]
- Using an unknown string in [] causes all the data until the next directive to be ignored and generates a warning

Here is an example of a topology file, urea.top:

```

;
;       Example topology file
;
; The force-field files to be included
#include "amber99.ff/forcefield.itp"

[ moleculetype ]
; name nrexcl
Urea      3

[ atoms ]
  1  C  1  URE      C      1      0.880229  12.01000  ; amber C  type
  2  O  1  URE      O      2     -0.613359  16.00000  ; amber O  type
  3  N  1  URE     N1      3     -0.923545  14.01000  ; amber N  type
  4  H  1  URE     H11     4      0.395055   1.00800  ; amber H  type
  5  H  1  URE     H12     5      0.395055   1.00800  ; amber H  type
  6  N  1  URE     N2      6     -0.923545  14.01000  ; amber N  type
  7  H  1  URE     H21     7      0.395055   1.00800  ; amber H  type
  8  H  1  URE     H22     8      0.395055   1.00800  ; amber H  type

[ bonds ]
  1  2
  1  3
  1  6
  3  4
  3  5
  6  7
  6  8

[ dihedrals ]
;   ai    aj    ak    al  funct  definition
    2     1     3     4    9
    2     1     3     5    9
    2     1     6     7    9
    2     1     6     8    9
    3     1     6     7    9
    3     1     6     8    9
    6     1     3     4    9
    6     1     3     5    9

[ dihedrals ]
    3     6     1     2    4
    1     4     3     5    4
    1     7     6     8    4

[ position_restraints ]
; you wouldn't normally use this for a molecule like Urea,
; but we include it here for didactic purposes
; ai    funct    fc
    1     1     1000    1000    1000 ; Restrain to a point

```

```

    2      1      1000      0      1000 ; Restrain to a line (Y-axis)
    3      1      1000      0          0 ; Restrain to a plane (Y-Z-plane)

[ dihedral_restraints ]
; ai   aj   ak   al   type   phi   dphi   fc
    3   6   1   2     1   180    0   10
    1   4   3   5     1   180    0   10

; Include TIP3P water topology
#include "amber99/tip3p.itp"

[ system ]
Urea in Water

[ molecules ]
;molecule name   nr.
Urea              1
SOL               1000

```

Here follows the explanatory text.

`#include "amber99.ff/forcefield.itp"` : this includes the information for the force field you are using, including bonded and non-bonded parameters. This example uses the AMBER99 force field, but your simulation may use a different force field. `grompp` will automatically go and find this file and copy-and-paste its content. That content can be seen in `share/top/amber99.ff/forcefield.itp`, and it is

```

#define _FF_AMBER
#define _FF_AMBER99

[ defaults ]
; nbfunc      comb-rule      gen-pairs      fudgeLJ  fudgeQQ
1             2             yes            0.5      0.8333

#include "ffnonbonded.itp"
#include "ffbonded.itp"
#include "gbsa.itp"

```

The two `#define` statements set up the conditions so that future parts of the topology can know that the AMBER 99 force field is in use.

[defaults] :

- `nbfunc` is the non-bonded function type. Use 1 (Lennard-Jones) or 2 (Buckingham)
- `comb-rule` is the number of the combination rule (see 5.3.2).
- `gen-pairs` is for pair generation. The default is 'no', *i.e.* get 1-4 parameters from the `pairtypes` list. When parameters are not present in the list, stop with a fatal error. Setting 'yes' generates 1-4 parameters that are not present in the pair list from normal Lennard-Jones parameters using `fudgeLJ`

- `fudgeLJ` is the factor by which to multiply Lennard-Jones 1-4 interactions, default 1
- `fudgeQQ` is the factor by which to multiply electrostatic 1-4 interactions, default 1
- N is the power for the repulsion term in a $6-N$ potential (with nonbonded-type Lennard-Jones only), starting with GROMACS version 4.5, `mdrun` also reads and applies N , for values not equal to 12 tabulated interaction functions are used (in older version you would have to use user tabulated interactions).

Note that `gen-pairs`, `fudgeLJ`, `fudgeQQ`, and N are optional. `fudgeLJ` is only used when `generate pairs` is set to ‘yes’, and `fudgeQQ` is always used. However, if you want to specify N you need to give a value for the other parameters as well.

Then some other `#include` statements add in the large amount of data needed to describe the rest of the force field. We will skip these and return to `urea.top`. There we will see

[`moleculetype`] : defines the name of your molecule in this `*.top` and `nrexcl = 3` stands for excluding non-bonded interactions between atoms that are no further than 3 bonds away.

[`atoms`] : defines the molecule, where `nr` and `type` are fixed, the rest is user defined. So `atom` can be named as you like, `cgnr` made larger or smaller (if possible, the total charge of a charge group should be zero), and charges can be changed here too.

[`bonds`] : no comment.

[`pairs`] : LJ and Coulomb 1-4 interactions

[`angles`] : no comment

[`dihedrals`] : in this case there are 9 proper dihedrals (`funct = 1`), 3 improper (`funct = 4`) and no Ryckaert-Bellemans type dihedrals. If you want to include Ryckaert-Bellemans type dihedrals in a topology, do the following (in case of *e.g.* decane):

```
[ dihedrals ]
; ai    aj    ak    al  funct      c0      c1      c2
   1     2     3     4     3
   2     3     4     5     3
```

In the original implementation of the potential for alkanes [136] no 1-4 interactions were used, which means that in order to implement that particular force field you need to remove the 1-4 interactions from the [`pairs`] section of your topology. In most modern force fields, like OPLS/AA or Amber the rules are different, and the Ryckaert-Bellemans potential is used as a cosine series in combination with 1-4 interactions.

[`position_restraints`] : harmonically restrain the selected particles to reference positions (4.3.1). The reference positions are read from a separate coordinate file by `grompp`.

[`dihedral_restraints`] : restrain selected dihedrals to a reference value. The implementation of dihedral restraints is described in section 4.3.4 of the manual. The parameters specified in the [`dihedral_restraints`] directive are as follows:

- `type` has only one possible value which is 1
- `phi` is the value of ϕ_0 in eqn. 4.83 and eqn. 4.84 of the manual.

- `dphi` is the value of $\Delta\phi$ in eqn. 4.84 of the manual.
- `fc` is the force constant k_{dihr} in eqn. 4.84 of the manual.

`#include "tip3p.itp"` : includes a topology file that was already constructed (see section 5.8.2).

`[system]` : title of your system, user-defined

`[molecules]` : this defines the total number of (sub)molecules in your system that are defined in this `*.top`. In this example file, it stands for 1 urea molecule dissolved in 1000 water molecules. The molecule type SOL is defined in the `tip3p.itp` file. Each name here must correspond to a name given with `[moleculetype]` earlier in the topology. The order of the blocks of molecule types and the numbers of such molecules must match the coordinate file that accompanies the topology when supplied to `grompp`. The blocks of molecules do not need to be contiguous, but some tools (e.g. `genion`) may act only on the first or last such block of a particular molecule type. Also, these blocks have nothing to do with the definition of groups (see sec. 3.3 and sec. 8.1).

5.8.2 Molecule.itp file

If you construct a topology file you will use frequently (like the water molecule, `tip3p.itp`, which is already constructed for you) it is good to make a `molecule.itp` file. This only lists the information of one particular molecule and allows you to re-use the `[moleculetype]` in multiple systems without re-invoking `pdb2gmx` or manually copying and pasting. An example `urea.itp` follows:

```
[ moleculetype ]
; molname nrexcl
URE 3

[ atoms ]
  1  C  1  URE      C      1      0.880229  12.01000  ; amber C  type
...
  8  H  1  URE     H22      8      0.395055   1.00800  ; amber H  type

[ bonds ]
  1  2
...
  6  8
[ dihedrals ]
;  ai    aj    ak    al  funct  definition
   2     1     3     4     9
...
   6     1     3     5     9
[ dihedrals ]
   3     6     1     2     4
   1     4     3     5     4
   1     7     6     8     4
```

Using *.itp files results in a very short *.top file:

```

;
;      Example topology file
;
; The force field files to be included
#include "amber99.ff/forcefield.itp"

#include "urea.itp"

; Include TIP3P water topology
#include "amber99/tip3p.itp"

[ system ]
Urea in Water

[ molecules ]
;molecule name      nr.
Urea                  1
SOL                   1000

```

5.8.3 Ifdef statements

A very powerful feature in GROMACS is the use of `#ifdef` statements in your *.top file. By making use of this statement, and associated `#define` statements like were seen in `amber99.ff/forcefield.itp` earlier, different parameters for one molecule can be used in the same *.top file. An example is given for TFE, where there is an option to use different charges on the atoms: charges derived by De Loof *et al.* [137] or by Van Buuren and Berendsen [138]. In fact, you can use much of the functionality of the C preprocessor, `cpp`, because `grompp` contains similar pre-processing functions to scan the file. The way to make use of the `#ifdef` option is as follows:

- either use the option `define = -DDeLoof` in the *.mdp file (containing `grompp` input parameters), or use the line `#define DeLoof` early in your *.top or *.itp file; and
- put the `#ifdef` statements in your *.top, as shown below:

...

```

[ atoms ]
; nr      type      resnr      residu      atom      cgnr      charge      mass
#ifdef DeLoof
; Use Charges from DeLoof
  1        C         1          TFE         C         1         0.74
  2        F         1          TFE         F         1        -0.25
  3        F         1          TFE         F         1        -0.25
  4        F         1          TFE         F         1        -0.25

```

```

    5      CH2      1      TFE      CH2      1      0.25
    6      OA      1      TFE      OA      1      -0.65
    7      HO      1      TFE      HO      1      0.41
#else
; Use Charges from VanBuuren
    1      C      1      TFE      C      1      0.59
    2      F      1      TFE      F      1      -0.2
    3      F      1      TFE      F      1      -0.2
    4      F      1      TFE      F      1      -0.2
    5      CH2     1      TFE      CH2     1      0.26
    6      OA      1      TFE      OA      1      -0.55
    7      HO      1      TFE      HO      1      0.3
#endif

[ bonds ]
; ai  aj  funct      c0      c1
    6   7   1  1.000000e-01  3.138000e+05
    1   2   1  1.360000e-01  4.184000e+05
    1   3   1  1.360000e-01  4.184000e+05
    1   4   1  1.360000e-01  4.184000e+05
    1   5   1  1.530000e-01  3.347000e+05
    5   6   1  1.430000e-01  3.347000e+05
...

```

This mechanism is used by `pdb2gmx` to implement optional position restraints (4.3.1) by `#including` an `.itp` file whose contents will be meaningful only if a particular `#define` is set (and spelled correctly!)

5.8.4 Topologies for free energy calculations

Free energy differences between two systems, A and B, can be calculated as described in sec. 3.12. Systems A and B are described by topologies consisting of the same number of molecules with the same number of atoms. Masses and non-bonded interactions can be perturbed by adding B parameters under the `[atoms]` directive. Bonded interactions can be perturbed by adding B parameters to the bonded types or the bonded interactions. The parameters that can be perturbed are listed in Tables 5.4 and 5.5. The λ -dependence of the interactions is described in section sec. 4.5. The bonded parameters that are used (on the line of the bonded interaction definition, or the ones looked up on atom types in the bonded type lists) is explained in Table 5.6. In most cases, things should work intuitively. When the A and B atom types in a bonded interaction are not all identical and parameters are not present for the B-state, either on the line or in the bonded types, `grompp` uses the A-state parameters and issues a warning. For free energy calculations, all or no parameters for topology B ($\lambda = 1$) should be added on the same line, after the normal parameters, in the same order as the normal parameters. From GROMACS 4.6 onward, if λ is treated as a vector, then the `bonded-lambdas` component controls all bonded terms that are not explicitly labeled as restraints. Restrain terms are controlled by the `restraint-lambdas` component.

Below is an example of a topology which changes from 200 propanols to 200 pentanes using the GROMOS-96 force field.

B-state atom types all identical to A-state atom types	parameters on line		parameters in bonded types				message
	A	B	A atom types		B atom types		
			A	B	A	B	
yes	+AB	–	x	x			error
	+A	+B	x	x			
	–	–	–	–			
	–	–	+AB	–			
	–	–	+A	+B			
no	+AB	–	x	x	x	x	warning
	+A	+B	x	x	x	x	error
	–	–	–	–	x	x	warning
	–	–	+AB	–	–	–	warning
	–	–	+A	+B	–	–	warning
	–	–	+A	x	+B	–	
	–	–	+A	x	+	+B	

Table 5.6: The bonded parameters that are used for free energy topologies, on the line of the bonded interaction definition or looked up in the bond types section based on atom types. A and B indicate the parameters used for state A and B respectively, + and – indicate the (non-)presence of parameters in the topology, x indicates that the presence has no influence.

```

; Include force field parameters
#include "gromos43a1.ff/forcefield.itp"

[ moleculetype ]
; Name                nrexcl
PropPent              3

[ atoms ]
; nr type resnr residue atom cgnr  charge    mass  typeB  chargeB  massB
  1  H   1     PROP   PH   1    0.398    1.008  CH3    0.0     15.035
  2  OA  1     PROP   PO   1   -0.548   15.9994 CH2    0.0     14.027
  3  CH2 1     PROP   PC1  1    0.150    14.027  CH2    0.0     14.027
  4  CH2 1     PROP   PC2  2    0.000    14.027
  5  CH3 1     PROP   PC3  2    0.000    15.035

[ bonds ]
; ai  aj funct  par_A  par_B
  1   2   2     gb_1  gb_26
  2   3   2     gb_17 gb_26
  3   4   2     gb_26 gb_26
  4   5   2     gb_26

[ pairs ]
; ai  aj funct
  1   4   1
  2   5   1

```

```
[ angles ]
; ai    aj    ak funct    par_A  par_B
    1    2    3     2    ga_11  ga_14
    2    3    4     2    ga_14  ga_14
    3    4    5     2    ga_14  ga_14

[ dihedrals ]
; ai    aj    ak    al funct    par_A  par_B
    1    2    3     4     1    gd_12  gd_17
    2    3    4     5     1    gd_17  gd_17

[ system ]
; Name
Propanol to Pentane

[ molecules ]
; Compound      #mols
PropPent        200
```

Atoms that are not perturbed, PC2 and PC3, do not need B-state parameter specifications, since the B parameters will be copied from the A parameters. Bonded interactions between atoms that are not perturbed do not need B parameter specifications, as is the case for the last bond in the example topology. Topologies using the OPLS/AA force field need no bonded parameters at all, since both the A and B parameters are determined by the atom types. Non-bonded interactions involving one or two perturbed atoms use the free-energy perturbation functional forms. Non-bonded interactions between two non-perturbed atoms use the normal functional forms. This means that when, for instance, only the charge of a particle is perturbed, its Lennard-Jones interactions will also be affected when lambda is not equal to zero or one.

Note that this topology uses the GROMOS-96 force field, in which the bonded interactions are not determined by the atom types. The bonded interaction strings are converted by the C-preprocessor. The force-field parameter files contain lines like:

```
#define gb_26      0.1530  7.1500e+06

#define gd_17      0.000    5.86      3
```

5.8.5 Constraint forces

The constraint force between two atoms in one molecule can be calculated with the free energy perturbation code by adding a constraint between the two atoms, with a different length in the A and B topology. When the B length is 1 nm longer than the A length and lambda is kept constant at zero, the derivative of the Hamiltonian with respect to lambda is the constraint force. For constraints between molecules, the pull code can be used, see sec. 6.4. Below is an example for calculating the constraint force at 0.7 nm between two methanes in water, by combining the two methanes into one “molecule.” **Note** that the definition of a “molecule” in GROMACS does not necessarily correspond to the chemical definition of a molecule. In GROMACS, a “molecule” can

be defined as any group of atoms that one wishes to consider simultaneously. The added constraint is of function type 2, which means that it is not used for generating exclusions (see sec. 5.4.4). Note that the constraint free energy term is included in the derivative term, and is specifically included in the `bonded-lambdas` component. However, the free energy for changing constraints is *not* included in the potential energy differences used for BAR and MBAR, as this requires reevaluating the energy at each of the constraint components. This functionality is planned for later versions.

```
; Include force-field parameters
#include "gromos43a1.ff/forcefield.itp"

[ moleculetype ]
; Name          nrexcl
Methanes        1

[ atoms ]
; nr   type   resnr  residu  atom   cgnr   charge   mass
  1   CH4     1     CH4     C1     1       0    16.043
  2   CH4     1     CH4     C2     2       0    16.043
[ constraints ]
; ai   aj  funct   length_A  length_B
  1    2    2       0.7       1.7

#include "gromos43a1.ff/spc.itp"

[ system ]
; Name
Methanes in Water

[ molecules ]
; Compound      #mols
Methanes        1
SOL              2002
```

5.8.6 Coordinate file

Files with the `.gro` file extension contain a molecular structure in GROMOS-87 format. A sample piece is included below:

```
MD of 2 waters, reformat step, PA aug-91
6
1WATER OW1 1 0.126 1.624 1.679 0.1227 -0.0580 0.0434
1WATER HW2 2 0.190 1.661 1.747 0.8085 0.3191 -0.7791
1WATER HW3 3 0.177 1.568 1.613 -0.9045 -2.6469 1.3180
2WATER OW1 4 1.275 0.053 0.622 0.2519 0.3140 -0.1734
2WATER HW2 5 1.337 0.002 0.680 -1.0641 -1.1349 0.0257
2WATER HW3 6 1.326 0.120 0.568 1.9427 -0.8216 -0.0244
1.82060 1.82060 1.82060
```

This format is fixed, *i.e.* all columns are in a fixed position. If you want to read such a file in your own program without using the GROMACS libraries you can use the following formats:

C-format: "%5i%5s%5s%5i%8.3f%8.3f%8.3f%8.4f%8.4f%8.4f"

Or to be more precise, with title *etc.* it looks like this:

```
"%s\n", Title
"%5d\n", natoms
for (i=0; (i<natoms); i++) {
    "%5d%-5s%5s%5d%8.3f%8.3f%8.3f%8.4f%8.4f%8.4f\n",
        residuenr, residuename, atomname, atomnr, x, y, z, vx, vy, vz
}
"%10.5f%10.5f%10.5f%10.5f%10.5f%10.5f%10.5f%10.5f%10.5f\n",
    box[X][X], box[Y][Y], box[Z][Z],
    box[X][Y], box[X][Z], box[Y][X], box[Y][Z], box[Z][X], box[Z][Y]
```

Fortran format: (i5, 2a5, i5, 3f8.3, 3f8.4)

So `conf.in.gro` is the GROMACS coordinate file and is almost the same as the GROMOS-87 file (for GROMOS users: when used with `ntx=7`). The only difference is the box for which GROMACS uses a tensor, not a vector.

5.9 Force field organization

5.9.1 Force-field files

Many force fields are available by default. Force fields are detected by the presence of `<name>.ff` directories in the `$GMXLIB/share/gromacs/top` sub-directory and/or the working directory. The information regarding the location of the force field files is printed by `pdb2gmx` so you can easily keep track of which version of a force field is being called, in case you have made modifications in one location or another. The force fields included with GROMACS are:

- AMBER03 protein, nucleic AMBER94 (Duan et al., J. Comp. Chem. 24, 1999-2012, 2003)
- AMBER94 force field (Cornell et al., JACS 117, 5179-5197, 1995)
- AMBER96 protein, nucleic AMBER94 (Kollman et al., Acc. Chem. Res. 29, 461-469, 1996)
- AMBER99 protein, nucleic AMBER94 (Wang et al., J. Comp. Chem. 21, 1049-1074, 2000)
- AMBER99SB protein, nucleic AMBER94 (Hornak et al., Proteins 65, 712-725, 2006)
- AMBER99SB-ILDN protein, nucleic AMBER94 (Lindorff-Larsen et al., Proteins 78, 1950-58, 2010)
- AMBERGS force field (Garcia & Sanbonmatsu, PNAS 99, 2782-2787, 2002)
- CHARMM27 all-atom force field (CHARM22 plus CMAP for proteins)
- GROMOS96 43a1 force field
- GROMOS96 43a2 force field (improved alkane dihedrals)
- GROMOS96 45a3 force field (Schuler JCC 2001 22 1205)

- GROMOS96 53a5 force field (JCC 2004 vol 25 pag 1656)
- GROMOS96 53a6 force field (JCC 2004 vol 25 pag 1656)
- GROMOS96 54a7 force field (Eur. Biophys. J. (2011), 40., 843-856, DOI: 10.1007/s00249-011-0700-9)
- OPLS-AA/L all-atom force field (2001 aminoacid dihedrals)

A force field is included at the beginning of a topology file with an `#include` statement followed by `<name>.ff/forcefield.itp`. This statement includes the force-field file, which, in turn, may include other force-field files. All the force fields are organized in the same way. An example of the `amber99.ff/forcefield.itp` was shown in 5.8.1.

For each force field, there several files which are only used by `pdb2gmx`. These are: residue databases (`.rtp`, see 5.7.1) the hydrogen database (`.hdb`, see 5.7.4), two termini databases (`.n.tdb` and `.c.tdb`, see 5.7.5) and the atom type database (`.atp`, see 5.2.1), which contains only the masses. Other optional files are described in sec. 5.7.

5.9.2 Changing force-field parameters

If one wants to change the parameters of few bonded interactions in a molecule, this is most easily accomplished by typing the parameters behind the definition of the bonded interaction directly in the `*.top` file under the `[moleculetype]` section (see 5.8.1 for the format and units). If one wants to change the parameters for all instances of a certain interaction one can change them in the force-field file or add a new `[???types]` section after including the force field. When parameters for a certain interaction are defined multiple times, the last definition is used. As of GROMACS version 3.1.3, a warning is generated when parameters are redefined with a different value. Changing the Lennard-Jones parameters of an atom type is not recommended, because in the GROMOS force fields the Lennard-Jones parameters for several combinations of atom types are not generated according to the standard combination rules. Such combinations (and possibly others that do follow the combination rules) are defined in the `[nonbond_params]` section, and changing the Lennard-Jones parameters of an atom type has no effect on these combinations.

5.9.3 Adding atom types

As of GROMACS version 3.1.3, atom types can be added in an extra `[atomtypes]` section after the the inclusion of the normal force field. After the definition of the new atom type(s), additional non-bonded and pair parameters can be defined. In pre-3.1.3 versions of GROMACS, the new atom types needed to be added in the `[atomtypes]` section of the force-field files, because all non-bonded parameters above the last `[atomtypes]` section would be overwritten using the standard combination rules.

Chapter 6

Special Topics

6.1 Free energy implementation

For free energy calculations, there are two things that must be specified; the end states, and the pathway connecting the end states. The end states can be specified in two ways. The most straightforward is through the specification of end states in the topology file. Most potential forms support both an *A* state and a *B* state. Whenever both states are specified, then the *A* state corresponds to the initial free energy state, and the *B* state corresponds to the final state.

In some cases, the end state can also be defined in some cases without altering the topology, solely through the `.mdp` file, through the use of the `couple-moltype`, `couple-lambda0`, `couple-lambda1`, and `couple-intramol` `mdp` keywords. Any molecule type selected in `couple-moltype` will automatically have a *B* state implicitly constructed (and the *A* state redefined) according to the `couple-lambda` keywords. `couple-lambda0` and `couple-lambda1` define the non-bonded parameters that are present in the *A* state (`couple-lambda0`) and the *B* state (`couple-lambda1`). The choices are 'q', 'vdw', and 'vdw-q'; these indicate the Coulombic, van der Waals, or both parameters that are turned on in the respective state.

Once the end states are defined, then the path between the end states has to be defined. This path is defined solely in the `.mdp` file. Starting in 4.6, λ is a vector of components, with Coulombic, van der Waals, bonded, restraint, and mass components all able to be adjusted independently. This makes it possible to turn off the Coulombic term linearly, and then the van der Waals using soft core, all in the same simulation. This is especially useful for replica exchange or expanded ensemble simulations, where it is important to sample all the way from interacting to non-interacting states in the same simulation to improve sampling.

`fep-lambdas` is the default array of λ values ranging from 0 to 1. All of the other `lambda` arrays use the values in this array if they are not specified. The previous behavior, where the pathway is controlled by a single λ variable, can be preserved by using only `fep-lambdas` to define the pathway.

For example, if you wanted to first to change the Coulombic terms, then the van der Waals terms, changing bonded at the same time rate as the van der Waals, but changing the restraints throughout

the first two-thirds of the simulation, then you could use this λ vector:

```
coul-lambdas      = 0.0 0.2 0.5 1.0 1.0 1.0 1.0 1.0 1.0 1.0
vdw-lambdas       = 0.0 0.0 0.0 0.0 0.4 0.5 0.6 0.7 0.8 1.0
bonded-lambdas    = 0.0 0.0 0.0 0.0 0.4 0.5 0.6 0.7 0.8 1.0
restraint-lambdas = 0.0 0.0 0.1 0.2 0.3 0.5 0.7 1.0 1.0 1.0
```

This is also equivalent to:

```
fep-lambdas       = 0.0 0.0 0.0 0.0 0.4 0.5 0.6 0.7 0.8 1.0
coul-lambdas      = 0.0 0.2 0.5 1.0 1.0 1.0 1.0 1.0 1.0 1.0
restraint-lambdas = 0.0 0.0 0.1 0.2 0.3 0.5 0.7 1.0 1.0 1.0
```

The `fep-lambda` array, in this case, is being used as the default to fill in the bonded and van der Waals λ arrays. Usually, it's best to fill in all arrays explicitly, just to make sure things are properly assigned.

If you want to turn on only restraints going from A to B , then it would be:

```
restraint-lambdas = 0.0 0.1 0.2 0.4 0.6 1.0
```

and all of the other components of the λ vector would be left in the A state.

To compute free energies with a vector λ using thermodynamic integration, then the TI equation becomes vector equation:

$$\Delta F = \int \langle \nabla H \rangle \cdot d\vec{\lambda} \quad (6.1)$$

or for finite differences:

$$\Delta F \approx \int \sum \langle \nabla H \rangle \cdot \Delta \lambda \quad (6.2)$$

The external `pymbar` script downloaded from <https://SimTK.org/home/pymbar> can compute this integral automatically from the GROMACS `dhdl.xvg` output.

6.2 Potential of mean force

A potential of mean force (PMF) is a potential that is obtained by integrating the mean force from an ensemble of configurations. In GROMACS, there are several different methods to calculate the mean force. Each method has its limitations, which are listed below.

- **pull code:** between the centers of mass of molecules or groups of molecules.
- **AWH code:** currently acts on coordinates provided by the pull code.
- **free-energy code with harmonic bonds or constraints:** between single atoms.
- **free-energy code with position restraints:** changing the conformation of a relatively immobile group of atoms.

- **pull code in limited cases:** between groups of atoms that are part of a larger molecule for which the bonds are constrained with SHAKE or LINCS. If the pull group is relatively large, the pull code can be used.

The pull and free-energy code are described in more detail in the following two sections.

Entropic effects

When a distance between two atoms or the centers of mass of two groups is constrained or restrained, there will be a purely entropic contribution to the PMF due to the rotation of the two groups [139]. For a system of two non-interacting masses the potential of mean force is:

$$V_{pmf}(r) = -(n_c - 1)k_B T \log(r) \quad (6.3)$$

where n_c is the number of dimensions in which the constraint works (i.e. $n_c = 3$ for a normal constraint and $n_c = 1$ when only the z -direction is constrained). Whether one needs to correct for this contribution depends on what the PMF should represent. When one wants to pull a substrate into a protein, this entropic term indeed contributes to the work to get the substrate into the protein. But when calculating a PMF between two solutes in a solvent, for the purpose of simulating without solvent, the entropic contribution should be removed. **Note** that this term can be significant; when at 300K the distance is halved, the contribution is 3.5 kJ mol⁻¹.

6.3 Non-equilibrium pulling

When the distance between two groups is changed continuously, work is applied to the system, which means that the system is no longer in equilibrium. Although in the limit of very slow pulling the system is again in equilibrium, for many systems this limit is not reachable within reasonable computational time. However, one can use the Jarzynski relation [140] to obtain the equilibrium free-energy difference ΔG between two distances from many non-equilibrium simulations:

$$\Delta G_{AB} = -k_B T \log \left\langle e^{-\beta W_{AB}} \right\rangle_A \quad (6.4)$$

where W_{AB} is the work performed to force the system along one path from state A to B, the angular bracket denotes averaging over a canonical ensemble of the initial state A and $\beta = 1/k_B T$.

6.4 The pull code

The pull code applies forces or constraints between the centers of mass of one or more pairs of groups of atoms. Each pull reaction coordinate is called a “coordinate” and it operates on usually two, but sometimes more, pull groups. A pull group can be part of one or more pull coordinates. Furthermore, a coordinate can also operate on a single group and an absolute reference position in space. The distance between a pair of groups can be determined in 1, 2 or 3 dimensions, or can be along a user-defined vector. The reference distance can be constant or can change linearly with time. Normally all atoms are weighted by their mass, but an additional weighting factor can also be used.

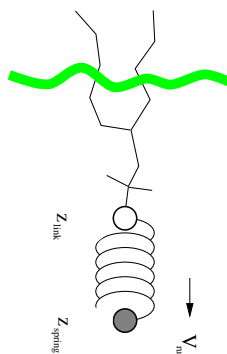


Figure 6.1: Schematic picture of pulling a lipid out of a lipid bilayer with umbrella pulling. V_{rup} is the velocity at which the spring is retracted, Z_{link} is the atom to which the spring is attached and Z_{spring} is the location of the spring.

Several different pull types, i.e. ways to apply the pull force, are supported, and in all cases the reference distance can be constant or linearly changing with time.

1. **Umbrella pulling** A harmonic potential is applied between the centers of mass of two groups. Thus, the force is proportional to the displacement.
2. **Constraint pulling** The distance between the centers of mass of two groups is constrained. The constraint force can be written to a file. This method uses the SHAKE algorithm but only needs 1 iteration to be exact if only two groups are constrained.
3. **Constant force pulling** A constant force is applied between the centers of mass of two groups. Thus, the potential is linear. In this case there is no reference distance or pull rate.
4. **Flat bottom pulling** Like umbrella pulling, but the potential and force are zero for coordinate values below (`pull-coord?-type = flat-bottom`) or above (`pull-coord?-type = flat-bottom-high`) a reference value. This is useful for restraining e.g. the distance between two molecules to a certain region.

In addition, there are different types of reaction coordinates, so-called pull geometries. These are set with the `.mdp` option `pull-coord?-geometry`.

Definition of the center of mass

In GROMACS, there are three ways to define the center of mass of a group. The standard way is a “plain” center of mass, possibly with additional weighting factors. With periodic boundary conditions it is no longer possible to uniquely define the center of mass of a group of atoms. Therefore, a reference atom is used. For determining the center of mass, for all other atoms in the group, the closest periodic image to the reference atom is used. This uniquely defines the center of mass. By default, the middle (determined by the order in the topology) atom is used as a reference atom, but the user can also select any other atom if it would be closer to center of the group.

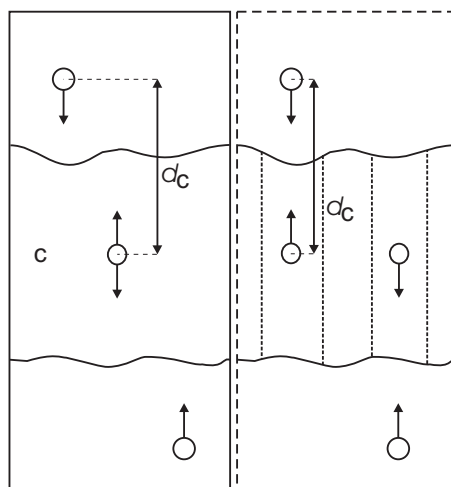


Figure 6.2: Comparison of a plain center of mass reference group versus a cylinder reference group applied to interface systems. C is the reference group. The circles represent the center of mass of two groups plus the reference group, d_c is the reference distance.

For a layered system, for instance a lipid bilayer, it may be of interest to calculate the PMF of a lipid as function of its distance from the whole bilayer. The whole bilayer can be taken as reference group in that case, but it might also be of interest to define the reaction coordinate for the PMF more locally. The `.mdp` option `pull-coord?-geometry = cylinder` does not use all the atoms of the reference group, but instead dynamically only those within a cylinder with radius `pull-cylinder-r` around the pull vector going through the pull group. This only works for distances defined in one dimension, and the cylinder is oriented with its long axis along this one dimension. To avoid jumps in the pull force, contributions of atoms are weighted as a function of distance (in addition to the mass weighting):

$$w(r < r_{\text{cyl}}) = 1 - 2 \left(\frac{r}{r_{\text{cyl}}} \right)^2 + \left(\frac{r}{r_{\text{cyl}}} \right)^4 \quad (6.5)$$

$$w(r \geq r_{\text{cyl}}) = 0 \quad (6.6)$$

Note that the radial dependence on the weight causes a radial force on both cylinder group and the other pull group. This is an undesirable, but unavoidable effect. To minimize this effect, the cylinder radius should be chosen sufficiently large. The effective mass is 0.47 times that of a cylinder with uniform weights and equal to the mass of uniform cylinder of 0.79 times the radius.

For a group of molecules in a periodic system, a plain reference group might not be well-defined. An example is a water slab that is connected periodically in x and y , but has two liquid-vapor interfaces along z . In such a setup, water molecules can evaporate from the liquid and they will move through the vapor, through the periodic boundary, to the other interface. Such a system is inherently periodic and there is no proper way of defining a “plain” center of mass along z . A proper solution is to using a cosine shaped weighting profile for all atoms in the reference group. The profile is a cosine with a single period in the unit cell. Its phase is optimized to give the maximum sum of weights, including mass weighting. This provides a unique and continuous reference position that is nearly identical to the plain center of mass position in case all atoms are

all within a half of the unit-cell length. See ref [141] for details.

When relative weights w_i are used during the calculations, either by supplying weights in the input or due to cylinder geometry or due to cosine weighting, the weights need to be scaled to conserve momentum:

$$w'_i = w_i \sum_{j=1}^N w_j m_j / \sum_{j=1}^N w_j^2 m_j \quad (6.7)$$

where m_j is the mass of atom j of the group. The mass of the group, required for calculating the constraint force, is:

$$M = \sum_{i=1}^N w'_i m_i \quad (6.8)$$

The definition of the weighted center of mass is:

$$\mathbf{r}_{com} = \sum_{i=1}^N w'_i m_i \mathbf{r}_i / M \quad (6.9)$$

From the centers of mass the AFM, constraint, or umbrella force \mathbf{F}_{com} on each group can be calculated. The force on the center of mass of a group is redistributed to the atoms as follows:

$$\mathbf{F}_i = \frac{w'_i m_i}{M} \mathbf{F}_{com} \quad (6.10)$$

Definition of the pull direction

The most common setup is to pull along the direction of the vector containing the two pull groups, this is selected with `pull-coord?-geometry = distance`. You might want to pull along a certain vector instead, which is selected with `pull-coord?-geometry = direction`. But this can cause unwanted torque forces in the system, unless you pull against a reference group with (nearly) fixed orientation, e.g. a membrane protein embedded in a membrane along x/y while pulling along z. If your reference group does not have a fixed orientation, you should probably use `pull-coord?-geometry = direction-relative`, see Fig. 6.3. Since the potential now depends on the coordinates of two additional groups defining the orientation, the torque forces will work on these two groups.

Definition of the angle and dihedral pull geometries

Four pull groups are required for `pull-coord?-geometry = angle`. In the same way as for geometries with two groups, each consecutive pair of groups i and $i + 1$ define a vector connecting the COMs of groups i and $i + 1$. The angle is defined as the angle between the two resulting vectors. E.g., the `.mdp` option `pull-coord?-groups = 1 2 2 4` defines the angle between the vector from the COM of group 1 to the COM of group 2 and the vector from the COM of group 2 to the COM of group 4. The angle takes values in the closed interval $[0, 180]$ deg. For `pull-coord?-geometry = angle-axis` the angle is defined with respect to a reference axis given by `pull-coord?-vec` and only two groups need to be given. The dihedral geometry requires six pull groups. These pair up in the same way as described above and

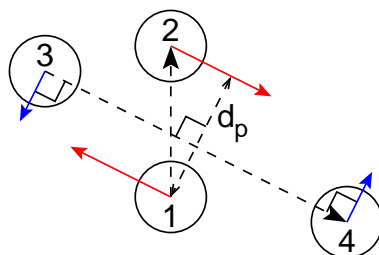


Figure 6.3: The pull setup for geometry `direction-relative`. The “normal” pull groups are 1 and 2. Groups 3 and 4 define the pull direction and thus the direction of the normal pull forces (red). This leads to reaction forces (blue) on groups 3 and 4, which are perpendicular to the pull direction. Their magnitude is given by the “normal” pull force times the ratio of d_p and the distance between groups 3 and 4.

so define three vectors. The dihedral angle is defined as the angle between the two planes spanned by the two first and the two last vectors. Equivalently, the dihedral angle can be seen as the angle between the first and the third vector when these vectors are projected onto a plane normal to the second vector (the axis vector). As an example, consider a dihedral angle involving four groups: 1, 5, 8 and 9. Here, the `.mdp` option `pull-coord?-groups = 8 1 1 5 5 9` specifies the three vectors that define the dihedral angle: the first vector is the COM distance vector from group 8 to 1, the second vector is the COM distance vector from group 1 to 5, and the third vector is the COM distance vector from group 5 to 9. The dihedral angle takes values in the interval $(-180, 180]$ deg and has periodic boundaries.

Limitations

There is one theoretical limitation: strictly speaking, constraint forces can only be calculated between groups that are not connected by constraints to the rest of the system. If a group contains part of a molecule of which the bond lengths are constrained, the pull constraint and LINCS or SHAKE bond constraint algorithms should be iterated simultaneously. This is not done in GROMACS. This means that for simulations with `constraints = all-bonds` in the `.mdp` file pulling is, strictly speaking, limited to whole molecules or groups of molecules. In some cases this limitation can be avoided by using the free energy code, see sec. 6.9. In practice, the errors caused by not iterating the two constraint algorithms can be negligible when the pull group consists of a large amount of atoms and/or the pull force is small. In such cases, the constraint correction displacement of the pull group is small compared to the bond lengths.

6.5 Adaptive biasing with AWH

The accelerated weight histogram method (AWH) [142] calculates the PMF along a reaction coordinate by adding an adaptively determined biasing potential. AWH flattens free energy barriers along the reaction coordinate by applying a history-dependent potential to the system that “fills up” free energy minima. This is similar in spirit to other adaptive biasing potential methods, e.g. the Wang-Landau [143], local elevation [144] and metadynamics [145] methods. The initial sam-

pling stage of AWH makes the method robust against the choice of input parameters. Furthermore, the target distribution along the reaction coordinate may be chosen freely.

6.5.1 Basics of the method

Rather than biasing the reaction coordinate $\xi(x)$ directly, AWH acts on a *reference coordinate* λ . The reaction coordinate $\xi(x)$ is coupled to λ with a harmonic potential

$$Q(\xi, \lambda) = \frac{1}{2}\beta k(\xi - \lambda)^2, \quad (6.11)$$

so that for large force constants k , $\xi \approx \lambda$. Note the use of dimensionless energies for compatibility with previously published work. Units of energy are obtained by multiplication with $k_B T = 1/\beta$. In the simulation, λ samples the user-defined sampling interval I . For a multidimensional reaction coordinate ξ , the sampling interval is the Cartesian product $I = \prod_{\mu} I_{\mu}$ (a rectangular domain). The connection between atom coordinates and λ is established through the extended ensemble [68],

$$P(x, \lambda) = \frac{1}{\mathcal{Z}} e^{g(\lambda) - Q(\xi(x), \lambda) - V(x)}, \quad (6.12)$$

where $g(\lambda)$ is a bias function (a free variable) and $V(x)$ is the unbiased potential energy of the system. The distribution along λ can be tuned to be any predefined *target distribution* $\rho(\lambda)$ (often chosen to be flat) by choosing $g(\lambda)$ wisely. This is evident from

$$P(\lambda) = \int P(x, \lambda) dx = \frac{1}{\mathcal{Z}} e^{g(\lambda)} \int e^{-Q(\xi(x), \lambda) - V(x)} dx \equiv \frac{1}{\mathcal{Z}} e^{g(\lambda) - F(\lambda)}, \quad (6.13)$$

where $F(\lambda)$ is the free energy

$$F(\lambda) = -\ln \int e^{-Q(\xi(x), \lambda) - V(x)} dx. \quad (6.14)$$

Being the convolution of the PMF with the Gaussian defined by the harmonic potential, $F(\lambda)$ is a smoothed version of the PMF. Eq. 6.13 shows that in order to obtain $P(\lambda) = \rho(\lambda)$, $F(\lambda)$ needs to be determined accurately. Thus, AWH adaptively calculates $F(\lambda)$ and simultaneously converges $P(\lambda)$ toward $\rho(\lambda)$.

The free energy update

AWH is initialized with an estimate of the free energy $F_0(\lambda)$. At regular time intervals this estimate is updated using data collected in between the updates. At update n , the applied bias $g_n(\lambda)$ is a function of the current free energy estimate $F_n(\lambda)$ and target distribution $\rho_n(\lambda)$,

$$g_n(\lambda) = \ln \rho_n(\lambda) + F_n(\lambda), \quad (6.15)$$

which is consistent with Eq. 6.13. Note that also the target distribution may be updated during the simulation (see examples in section 6.5.3). Substituting this choice of $g = g_n$ back into Eq. 6.13 yields the simple free energy update

$$\Delta F_n(\lambda) = F(\lambda) - F_n(\lambda) = -\ln \frac{P_n(\lambda)}{\rho_n(\lambda)}, \quad (6.16)$$

which would yield a better estimate $F_{n+1} = F_n + \Delta F_n$, assuming $P_n(\lambda)$ can be measured accurately. AWH estimates $P_n(\lambda)$ by regularly calculating the conditional distribution

$$\omega_n(\lambda|x) \equiv P_n(\lambda|x) = \frac{e^{g_n(\lambda) - Q(\xi(x), \lambda)}}{\sum_{\lambda'} e^{g_n(\lambda') - Q(\xi(x), \lambda')}}. \quad (6.17)$$

Accumulating these probability weights yields $\sum_t \omega(\lambda|x(t)) \sim P_n(\lambda)$, where $\int P_n(\lambda|x)P_n(x)dx = P_n(\lambda)$ has been used. The $\omega_n(\lambda|x)$ weights are thus the samples of the AWH method. With the limited amount of sampling one has in practice, update scheme 6.16 yields very noisy results. AWH instead applies a free energy update that has the same form but which can be applied repeatedly with limited and localized sampling,

$$\Delta F_n = -\ln \frac{W_n(\lambda) + \sum_t \omega_n(\lambda|x(t))}{W_n(\lambda) + \sum_t \rho_n(\lambda)}. \quad (6.18)$$

Here $W_n(\lambda)$ is the *reference weight histogram* representing prior sampling. The update for $W(\lambda)$, disregarding the initial stage (see section 6.5.2), is

$$W_{n+1}(\lambda) = W_n(\lambda) + \sum_t \rho_n(\lambda). \quad (6.19)$$

Thus, the weight histogram equals the targeted, “ideal” history of samples. There are two important things to note about the free energy update. First, sampling is driven away from oversampled, currently local regions. For such λ values, $\omega_n(\lambda) > \rho_n(\lambda)$ and $\Delta F_n(\lambda) < 0$, which by Eq. 6.15 implies $\Delta g_n(\lambda) < 0$ (assuming $\Delta \rho_n \equiv 0$). Thus, the probability to sample λ decreases after the update (see Eq. 6.13). Secondly, the normalization of the histogram $N_n = \sum_{\lambda} W_n(\lambda)$, determines the update size $|\Delta F(\lambda)|$. For instance, for a single sample $\omega(\lambda|x)$, the shape of the update is approximately a Gaussian function of width $\sigma = 1/\sqrt{\beta k}$ and height $\propto 1/N_n$ [142],

$$|\Delta F_n(\lambda)| \propto \frac{1}{N_n} e^{-\frac{1}{2}\beta k(\xi(x) - \lambda)^2}. \quad (6.20)$$

Therefore, as samples accumulate in $W(\lambda)$ and N_n grows, the updates get smaller, allowing for the free energy to converge.

Note that quantity of interest to the user is not $F(\lambda)$ but the PMF $\Phi(\xi)$. $\Phi(\xi)$ is extracted by reweighting samples $\xi(t)$ on the fly [142] (see also section 6.5.5) and will converge at the same rate as $F(\lambda)$, see Fig. 6.4. The PMF will be written to output (see section 6.5.7).

Applying the bias to the system

The bias potential can be applied to the system in two ways. Either by applying a harmonic potential centered at $\lambda(t)$, which is sampled using (rejection-free) Monte-Carlo sampling from the conditional distribution $\omega_n(\lambda|x(t)) = P_n(\lambda|x(t))$, see Eq. 6.17. This is also called Gibbs sampling or independence sampling. Alternatively, and by default in the code, the following *convolved bias potential* can be applied,

$$U_n(\xi) = -\ln \int e^{g_n(\lambda) - Q(\xi, \lambda)} d\lambda. \quad (6.21)$$

These two approaches are equivalent in the sense that they give rise to the same biased probabilities $P_n(x)$ (cf. 6.12) while the dynamics are clearly different in the two cases. This choice does not affect the internals of the AWH algorithm, only what force and potential AWH returns to the MD engine.

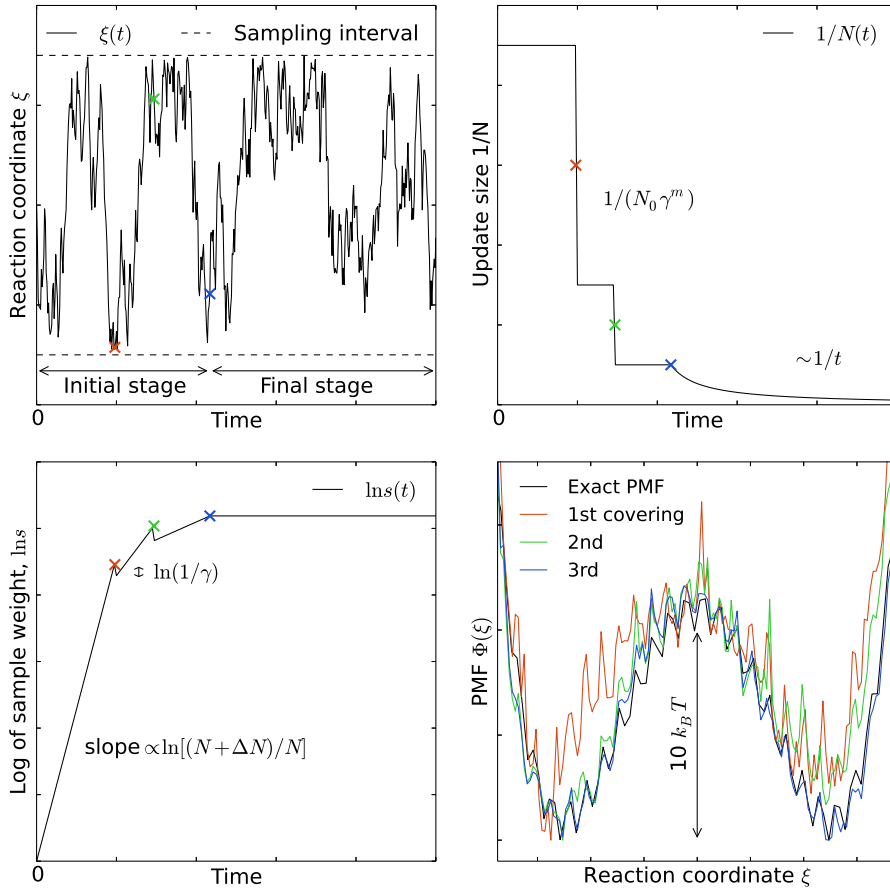


Figure 6.4: AWH evolution in time for a Brownian particle in a double-well potential. The reaction coordinate $\xi(t)$ traverses the sampling interval multiple times in the initial stage before exiting and entering the final stage (top left). In the final stage, the dynamics of ξ becomes increasingly diffusive. The times of covering are shown as \times -markers of different colors. At these times the free energy update size $\sim 1/N$, where N is the size of the weight histogram, is decreased by scaling N by a factor of $\gamma = 3$ (top right). In the final stage, N grows at the sampling rate and thus $1/N \sim 1/t$. The exit from the final stage is determined on the fly by ensuring that the effective sample weight s of data collected in the final stage exceeds that of initial stage data (bottom left; note that $\ln s(t)$ is plotted). An estimate of the PMF is also extracted from the simulation (bottom right), which after exiting the initial stage should estimate global free energy differences fairly accurately.

6.5.2 The initial stage

Initially, when the bias potential is far from optimal, samples will be highly correlated. In such cases, letting $W(\lambda)$ accumulate samples as prescribed by Eq. 6.19, entails a too rapid decay of the free energy update size. This motivates splitting the simulation into an *initial stage* where the weight histogram grows according to a more restrictive and robust protocol, and a *final stage* where the the weight histogram grows linearly at the sampling rate (Eq. 6.19). The AWH initial stage takes inspiration from the well-known Wang-Landau algorithm [143], although there are differences in the details.

In the initial stage the update size is kept constant (by keeping N_n constant) until a transition across the sampling interval has been detected, a “covering”. For the definition of a covering, see Eq. 6.22 below. After a covering has occurred, N_n is scaled up by a constant “growth factor” γ , chosen heuristically as $\gamma = 3$. Thus, in the initial stage N_n is set dynamically as $N_n = \gamma^m N_0$, where m is the number of coverings. Since the update size scales as $1/N$ (Eq. 6.20) this leads to a close to exponential decay of the update size in the initial stage, see Fig. 6.4.

The update size directly determines the rate of change of $F_n(\lambda)$ and hence, from Eq. 6.15, also the rate of change of the bias function $g_n(\lambda)$. Thus initially, when N_n is kept small and updates large, the system will be driven along the reaction coordinate by the constantly fluctuating bias. If N_0 is set small enough, the first transition will typically be fast because of the large update size and will quickly give a first rough estimate of the free energy. The second transition, using $N_1 = \gamma N_0$ refines this estimate further. Thus, rather than very carefully filling free energy minima using a small initial update size, the sampling interval is swepted back-and-forth multiple times, using a wide range of update sizes, see Fig. 6.4. This way, the initial stage also makes AWH robust against the choice of N_0 .

The covering criterion

In the general case of a multidimensional reaction coordinate $\lambda = (\lambda_\mu)$, the sampling interval I is considered covered when all dimensions have been covered. A dimension d is covered if all points λ_μ in the one-dimensional sampling interval I_μ have been “visited”. Finally, a point $\lambda_\mu \in I_\mu$ has been visited if there is at least one point $\lambda^* \in I$ with $\lambda_\mu^* = \lambda_\mu$ that since the last covering has accumulated probability weight corresponding to the peak of a multidimensional Gaussian distribution

$$\Delta W(\lambda^*) \geq w_{\text{peak}} \equiv \prod_{\mu} \frac{\Delta \lambda_{\mu}}{\sqrt{2\pi}\sigma_k}. \quad (6.22)$$

Here, $\Delta \lambda_{\mu}$ is the point spacing of the discretized I_{μ} and $\sigma_k = 1/\sqrt{\beta k_{\mu}}$ (where k_{μ} is the force constant) is the Gaussian width.

Exit from the initial stage

For longer times, when major free energy barriers have largely been flattened by the converging bias potential, the histogram $W(\lambda)$ should grow at the actual sampling rate and the initial stage needs to be exited [146]. There are multiple reasonable (heuristic) ways of determining when this transition should take place. One option is to postulate that the number of samples in the

weight histogram N_n should never exceed the actual number of collected samples, and exit the initial stage when this condition breaks [142]. In the initial stage, N grows close to exponentially while the collected number of samples grows linearly, so an exit will surely occur eventually. Here we instead apply an exit criterion based on the observation that “artificially” keeping N constant while continuing to collect samples corresponds to scaling down the relative weight of old samples relative to new ones. Similarly, the subsequent scaling up of N by a factor γ corresponds to scaling up the weight of old data. Briefly, the exit criterion is devised such that the weight of a sample collected *after* the initial stage is always larger or equal to the weight of a sample collected *during* the initial stage, see Fig. 6.4. This is consistent with scaling down early, noisy data.

The initial stage exit criterion will now be described in detail. We start out at the beginning of a covering stage, so that N has just been scaled by γ and is now kept constant. Thus, the first sample of this stage has the weight $s = 1/\gamma$ relative to the last sample of the previous covering stage. We assume that ΔN samples are collected and added to W for each update. To keep N constant, W needs to be scaled down by a factor $N/(N + \Delta N)$ after every update. Equivalently, this means that new data is scaled up relative to old data by the inverse factor. Thus, after Δn updates a new sample has the relative weight $s = (1/\gamma)[(N_n + \Delta N)/N_n]^{\Delta n}$. Now assume covering occurs at this time. To continue to the next covering stage, N should be scaled by γ , which corresponds to again multiplying s by $1/\gamma$. If at this point $s \geq \gamma$, then after rescaling $s \geq 1$; i.e. overall the relative weight of a new sample relative to an old sample is still growing fast. If on the contrary $s < \gamma$, and this defines the exit from the initial stage, then the initial stage is over and from now N simply grows at the sampling rate (see Eq. 6.19). To really ensure that $s \geq 1$ holds before exiting, so that samples after the exit have at least the sample weight of older samples, the last covering stage is extended by a sufficient number of updates.

6.5.3 Choice of target distribution

The target distribution $\rho(\lambda)$ is traditionally chosen to be uniform

$$\rho_{\text{const}}(\lambda) = \text{const.} \quad (6.23)$$

This choice exactly flattens $F(\lambda)$ in user-defined sampling interval I . Generally, $\rho(\lambda) = 0, \lambda \notin I$. In certain cases other choices may be preferable. For instance, in the multidimensional case the rectangular sampling interval is likely to contain regions of very high free energy, e.g. where atoms are clashing. To exclude such regions, $\rho(\lambda)$ can be specified by the following function of the free energy

$$\rho_{\text{cut}}(\lambda) \propto \frac{1}{1 + e^{F(\lambda) - F_{\text{cut}}}}, \quad (6.24)$$

where F_{cut} is a free energy cutoff (relative to $\min_{\lambda} F(\lambda)$). Thus, regions of the sampling interval where $F(\lambda) > F_{\text{cut}}$ will be exponentially suppressed (in a smooth fashion). Alternatively, very high free energy regions could be avoided while still flattening more moderate free energy barriers by targeting a Boltzmann distribution corresponding to scaling $\beta = 1/k_B T$ by a factor $0 < s_{\beta} < 1$,

$$\rho_{\text{Boltz}}(\lambda) \propto e^{-s_{\beta} F(\lambda)}, \quad (6.25)$$

The parameter s_{β} determines to what degree the free energy landscape is flattened; the lower s_{β} , the flatter. Note that both $\rho_{\text{cut}}(\lambda)$ and $\rho_{\text{Boltz}}(\lambda)$ depend on $F(\lambda)$, which needs to be substituted

by the current best estimate $F_n(\lambda)$. Thus, the target distribution is also updated (consistently with Eq. 6.15).

There is in fact an alternative approach to obtaining $\rho_{\text{Boltz}}(\lambda)$ as the limiting target distribution in AWH, which is particular in the way the weight histogram $W(\lambda)$ and the target distribution ρ are updated and coupled to each other. This yields an evolution of the bias potential which is very similar to that of well-tempered metadynamics [147], see [142] for details. Because of the popularity and success of well-tempered metadynamics, this is a special case worth considering. In this case ρ is a function of the reference weight histogram

$$\rho_{\text{Boltz,loc}}(\lambda) \propto W(\lambda), \quad (6.26)$$

and the update of the weight histogram is modified (cf. Eq. 6.19)

$$W_{n+1}(\lambda) = W_n(\lambda) + s_\beta \sum_t \omega(\lambda|x(t)). \quad (6.27)$$

Thus, here the weight histogram equals the real history of samples, but scaled by s_β . This target distribution is called *local* Boltzmann since W is only modified locally, where sampling has taken place. We see that when $s_\beta \approx 0$ the histogram essentially does not grow and the size of the free energy update will stay at a constant value (as in the original formulation of metadynamics). Thus, the free energy estimate will not converge, but continue to fluctuate around the correct value. This illustrates the inherent coupling between the convergence and choice of target distribution for this special choice of target. Furthermore note that when using $\rho = \rho_{\text{Boltz,loc}}$ there is no initial stage (section 6.5.2). The rescaling of the weight histogram applied in the initial stage is a global operation, which is incompatible $\rho_{\text{Boltz,loc}}$ only depending locally on the sampling history.

Lastly, the target distribution can be modulated by arbitrary probability weights

$$\rho(\lambda) = \rho_0(\lambda)w_{\text{user}}(\lambda). \quad (6.28)$$

where $w_{\text{user}}(\lambda)$ is provided by user data and in principle $\rho_0(\lambda)$ can be any of the target distributions mentioned above.

6.5.4 Multiple independent or sharing biases

Multiple independent bias potentials may be applied within one simulation. This only makes sense if the biased coordinates $\xi^{(1)}$, $\xi^{(2)}$, \dots evolve essentially independently from one another. A typical example of this would be when applying an independent bias to each monomer of a protein. Furthermore, multiple AWH simulations can be launched in parallel, each with a (set of) independent biases.

If the defined sampling interval is large relative to the diffusion time of the reaction coordinate, traversing the sampling interval multiple times as is required by the initial stage (section 6.5.2) may take an infeasible amount of simulation time. In these cases it could be advantageous to parallelize the work and have a group of multiple “walkers” $\xi^{(i)}(t)$ share a single bias potential. This can be achieved by collecting samples from all $\xi^{(i)}$ of the same sharing group into a single histogram and update a common free energy estimate. Samples can be shared between walkers within the simulation and/or between multiple simulations. However, currently only sharing between simulations is supported in the code while all biases within a simulation are independent.

Note that when attempting to shorten the simulation time by using bias-sharing walkers, care must be taken to ensure the simulations are still long enough to properly explore and equilibrate all regions of the sampling interval. To begin, the walkers in a group should be decorrelated and distributed approximately according to the target distribution before starting to refine the free energy. This can be achieved e.g. by “equilibrating” the shared weight histogram before letting it grow; for instance, $W(\lambda)/N \approx \rho(\lambda)$ with some tolerance.

Furthermore, the “covering” or transition criterion of the initial stage should to be generalized to detect when the sampling interval has been collectively traversed. One alternative is to just use the same criterion as for a single walker (but now with more samples), see Eq. 6.22. However, in contrast to the single walker case this does not ensure that any real transitions across the sampling interval has taken place; in principle all walkers could be sampling only very locally and still cover the whole interval. Just as with a standard umbrella sampling procedure, the free energy may appear to be converged while in reality simulations sampling closeby λ values are sampling disconnected regions of phase space. A stricter criterion, which helps avoid such issues, is to require that before a simulation marks a point λ_μ along dimension μ as visited, and shares this with the other walkers, also all points within a certain diameter D_{cover} should have been visited (i.e. fulfill Eq. 6.22). Increasing D_{cover} increases robustness, but may slow down convergence. For the maximum value of D_{cover} , equal to the length of the sampling interval, the sampling interval is considered covered when at least one walker has independently traversed the sampling interval.

6.5.5 Reweighting and combining biased data

Often one may want to, post-simulation, calculate the unbiased PMF $\Phi(u)$ of another variable $u(x)$. $\Phi(u)$ can be estimated using ξ -biased data by reweighting (“unbiasing”) the trajectory using the bias potential $U_{n(t)}$, see Eq. 6.21. Essentially, one bins the biased data along u and removes the effect of $U_{n(t)}$ by dividing the weight of samples $u(t)$ by $e^{-U_{n(t)}(\xi(t))}$,

$$\hat{\Phi}(u) = -\ln \sum_t 1_u(u(t)) e^{U_{n(t)}(\xi(t))} \mathcal{Z}_{n(t)}. \quad (6.29)$$

Here the indicator function 1_u denotes the binning procedure: $1_u(u') = 1$ if u' falls into the bin labeled by u and 0 otherwise. The normalization factor $\mathcal{Z}_n = \int e^{-\Phi(\xi) - U_n(\xi)} d\xi$ is the partition function of the extended ensemble. As can be seen \mathcal{Z}_n depends on $\Phi(\xi)$, the PMF of the (biased) reaction coordinate ξ (which is calculated and written to file by the AWH simulation). It is advisable to use only final stage data in the reweighting procedure due to the rapid change of the bias potential during the initial stage. If one would include initial stage data, one should use the sample weights that are inferred by the repeated rescaling of the histogram in the initial stage, for the sake of consistency. Initial stage samples would then in any case be heavily scaled down relative to final stage samples. Note that Eq. 6.29 can also be used to combine data from multiple simulations (by adding another sum also over the trajectory set). Furthermore, when multiple independent AWH biases have generated a set of PMF estimates $\{\hat{\Phi}^{(i)}(\xi)\}$, a combined best estimate $\hat{\Phi}(\xi)$ can be obtained by applying self-consistent exponential averaging. More details on this procedure and a derivation of Eq. 6.29 (using slightly different notation) can be found in [148].

6.5.6 The friction metric

During the AWH simulation, the following time-integrated force correlation function is calculated,

$$\eta_{\mu\nu}(\lambda) = \beta \int_0^\infty \frac{\langle \delta\mathcal{F}_\mu(x(t), \lambda) \delta\mathcal{F}_\nu(x(0), \lambda) \omega(\lambda|x(t)) \omega(\lambda|x(0)) \rangle}{\langle \omega^2(\lambda|x) \rangle} dt. \quad (6.30)$$

Here $\mathcal{F}_\mu(x, \lambda) = k_\mu(\xi_\mu(x) - \lambda_\mu)$ is the force along dimension μ from an harmonic potential centered at λ and $\delta\mathcal{F}_\mu(x, \lambda) = \mathcal{F}_\mu(x, \lambda) - \langle \mathcal{F}_\mu(x, \lambda) \rangle$ is the deviation of the force. The factors $\omega(\lambda|x(t))$, see Eq. 6.17, reweight the samples. $\eta_{\mu\nu}(\lambda)$ is a friction tensor [149]. Its matrix elements are inversely proportional to local diffusion coefficients. A measure of sampling (in)efficiency at each λ is given by

$$\eta^{\frac{1}{2}}(\lambda) = \sqrt{\det \eta_{\mu\nu}(\lambda)}. \quad (6.31)$$

A large value of $\eta^{\frac{1}{2}}(\lambda)$ indicates slow dynamics and long correlation times, which may require more sampling.

6.5.7 Usage

AWH stores data in the energy file (`.edr`) with a frequency set by the user. The data – the PMF, the convolved bias, distributions of the λ and ξ coordinates, etc. – can be extracted after the simulation using the `gmx_awh` tool. Furthermore, the trajectory of the reaction coordinate $\xi(t)$ is printed to the pull output file `pullx.xvg`. The log file (`.log`) also contains information; check for messages starting with “awh”, they will tell you about covering and potential sampling issues.

Setting the initial update size

The initial value of the weight histogram size N sets the initial update size (and the rate of change of the bias). When N is kept constant, like in the initial stage, the average variance of the free energy scales as $\varepsilon^2 \sim 1/(ND)$ [142], for a simple model system with constant diffusion D along the reaction coordinate. This provides a ballpark estimate used by AWH to initialize N in terms of more meaningful quantities

$$\frac{1}{N_0} = \frac{1}{N_0(\varepsilon_0, D)} \sim D\varepsilon_0^2. \quad (6.32)$$

Essentially, this tells us that a slower system (small D) requires more samples (larger N^0) to attain the same level of accuracy (ε_0) at a given sampling rate. Conversely, for a system of given diffusion, how to choose the initial biasing rate depends on how good the initial accuracy is. Both the initial error ε_0 and the diffusion D only need to be roughly estimated or guessed. In the typical case, one would only tweak the D parameter, and use a default value for ε_0 . For good convergence, D should be chosen as large as possible (while maintaining a stable system) giving large initial bias updates and fast initial transitions. Choosing D too small can lead to slow initial convergence. It may be a good idea to run a short trial simulation and after the first covering check the maximum free energy difference of the PMF estimate. If this is much larger than the expected magnitude of the free energy barriers that should be crossed, then the system is probably being pulled too hard and D should be decreased. ε_0 on the other hand, would only be tweaked when starting an AWH simulation using a fairly accurate guess of the PMF as input.

Tips for efficient sampling

The force constant k should be larger than the curvature of the PMF landscape. If this is not the case, the distributions of the reaction coordinate ξ and the reference coordinate λ , will differ significantly and warnings will be printed in the log file. One can choose k as large as the time step supports. This will necessarily increase the number of points of the discretized sampling interval I . In general however, it should not affect the performance of the simulation noticeably because the AWH update is implemented such that only sampled points are accessed at free energy update time.

As with any method, the choice of reaction coordinate(s) is critical. If a single reaction coordinate does not suffice, identifying a second reaction coordinate and sampling the two-dimensional landscape may help. In this case, using a target distribution with a free energy cutoff (see Eq. 6.24) might be required to avoid sampling uninteresting regions of very high free energy. Obtaining accurate free energies for reaction coordinates of much higher dimensionality than 3 or possibly 4 is generally not feasible.

Monitoring the transition rate of $\xi(t)$, across the sampling interval is also advisable. For reliable statistics (e.g. when reweighting the trajectory as described in section 6.5.5), one would generally want to observe at least a few transitions after having exited the initial stage. Furthermore, if the dynamics of the reaction coordinate suddenly changes, this may be a sign of e.g. a reaction coordinate problem.

Difficult regions of sampling may also be detected by calculating the friction tensor $\eta_{\mu\nu}(\lambda)$ in the sampling interval, see section 6.5.6. $\eta_{\mu\nu}(\lambda)$ as well as the sampling efficiency measure $\eta^{\frac{1}{2}}(\lambda)$ (Eq. 6.31) are written to the energy file and can be extracted with `gmx awh`. A high peak in $\eta^{\frac{1}{2}}(\lambda)$ indicates that this region requires longer time to sample properly.

6.6 Enforced Rotation

This module can be used to enforce the rotation of a group of atoms, as e.g. a protein subunit. There are a variety of rotation potentials, among them complex ones that allow flexible adaptations of both the rotated subunit as well as the local rotation axis during the simulation. An example application can be found in ref. [150].

6.6.1 Fixed Axis Rotation

Stationary Axis with an Isotropic Potential

In the fixed axis approach (see Fig. 6.5B), torque on a group of N atoms with positions \mathbf{x}_i (denoted “rotation group”) is applied by rotating a reference set of atomic positions – usually their initial positions \mathbf{y}_i^0 – at a constant angular velocity ω around an axis defined by a direction vector $\hat{\mathbf{v}}$ and a pivot point \mathbf{u} . To that aim, each atom with position \mathbf{x}_i is attracted by a “virtual spring” potential to its moving reference position $\mathbf{y}_i = \mathbf{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u})$, where $\mathbf{\Omega}(t)$ is a matrix that describes the

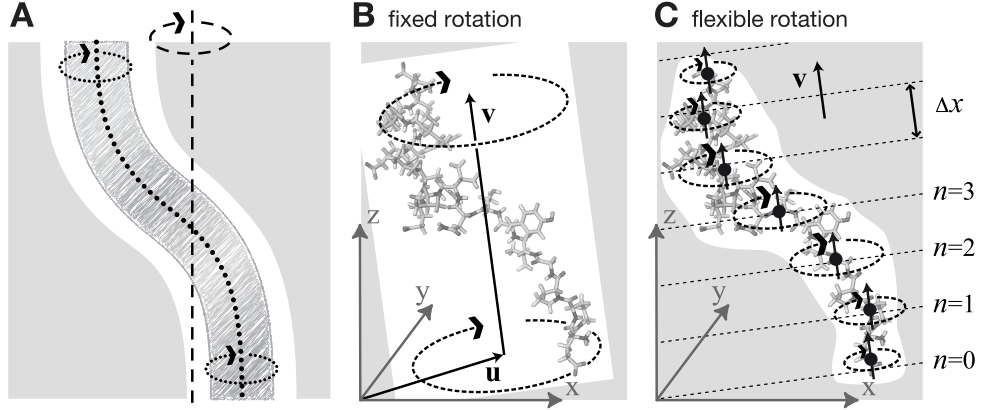


Figure 6.5: Comparison of fixed and flexible axis rotation. **A**: Rotating the sketched shape inside the white tubular cavity can create artifacts when a fixed rotation axis (dashed) is used. More realistically, the shape would revolve like a flexible pipe-cleaner (dotted) inside the bearing (gray). **B**: Fixed rotation around an axis \mathbf{v} with a pivot point specified by the vector \mathbf{u} . **C**: Subdividing the rotating fragment into slabs with separate rotation axes (\uparrow) and pivot points (\bullet) for each slab allows for flexibility. The distance between two slabs with indices n and $n + 1$ is Δx .

rotation around the axis. In the simplest case, the “springs” are described by a harmonic potential,

$$V^{\text{iso}} = \frac{k}{2} \sum_{i=1}^N w_i \left[\mathbf{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u}) - (\mathbf{x}_i - \mathbf{u}) \right]^2, \quad (6.33)$$

with optional mass-weighted prefactors $w_i = N m_i / M$ with total mass $M = \sum_{i=1}^N m_i$. The rotation matrix $\mathbf{\Omega}(t)$ is

$$\mathbf{\Omega}(t) = \begin{pmatrix} \cos \omega t + v_x^2 \xi & v_x v_y \xi - v_z \sin \omega t & v_x v_z \xi + v_y \sin \omega t \\ v_x v_y \xi + v_z \sin \omega t & \cos \omega t + v_y^2 \xi & v_y v_z \xi - v_x \sin \omega t \\ v_x v_z \xi - v_y \sin \omega t & v_y v_z \xi + v_x \sin \omega t & \cos \omega t + v_z^2 \xi \end{pmatrix},$$

where v_x, v_y , and v_z are the components of the normalized rotation vector $\hat{\mathbf{v}}$, and $\xi := 1 - \cos(\omega t)$. As illustrated in Fig. 6.6A for a single atom j , the rotation matrix $\mathbf{\Omega}(t)$ operates on the initial reference positions $\mathbf{y}_j^0 = \mathbf{x}_j(t_0)$ of atom j at $t = t_0$. At a later time t , the reference position has rotated away from its initial place (along the blue dashed line), resulting in the force

$$\mathbf{F}_j^{\text{iso}} = -\nabla_j V^{\text{iso}} = k w_j \left[\mathbf{\Omega}(t)(\mathbf{y}_j^0 - \mathbf{u}) - (\mathbf{x}_j - \mathbf{u}) \right], \quad (6.34)$$

which is directed towards the reference position.

Pivot-Free Isotropic Potential

Instead of a fixed pivot vector \mathbf{u} this potential uses the center of mass \mathbf{x}_c of the rotation group as pivot for the rotation axis,

$$\mathbf{x}_c = \frac{1}{M} \sum_{i=1}^N m_i \mathbf{x}_i \quad \text{and} \quad \mathbf{y}_c^0 = \frac{1}{M} \sum_{i=1}^N m_i \mathbf{y}_i^0, \quad (6.35)$$

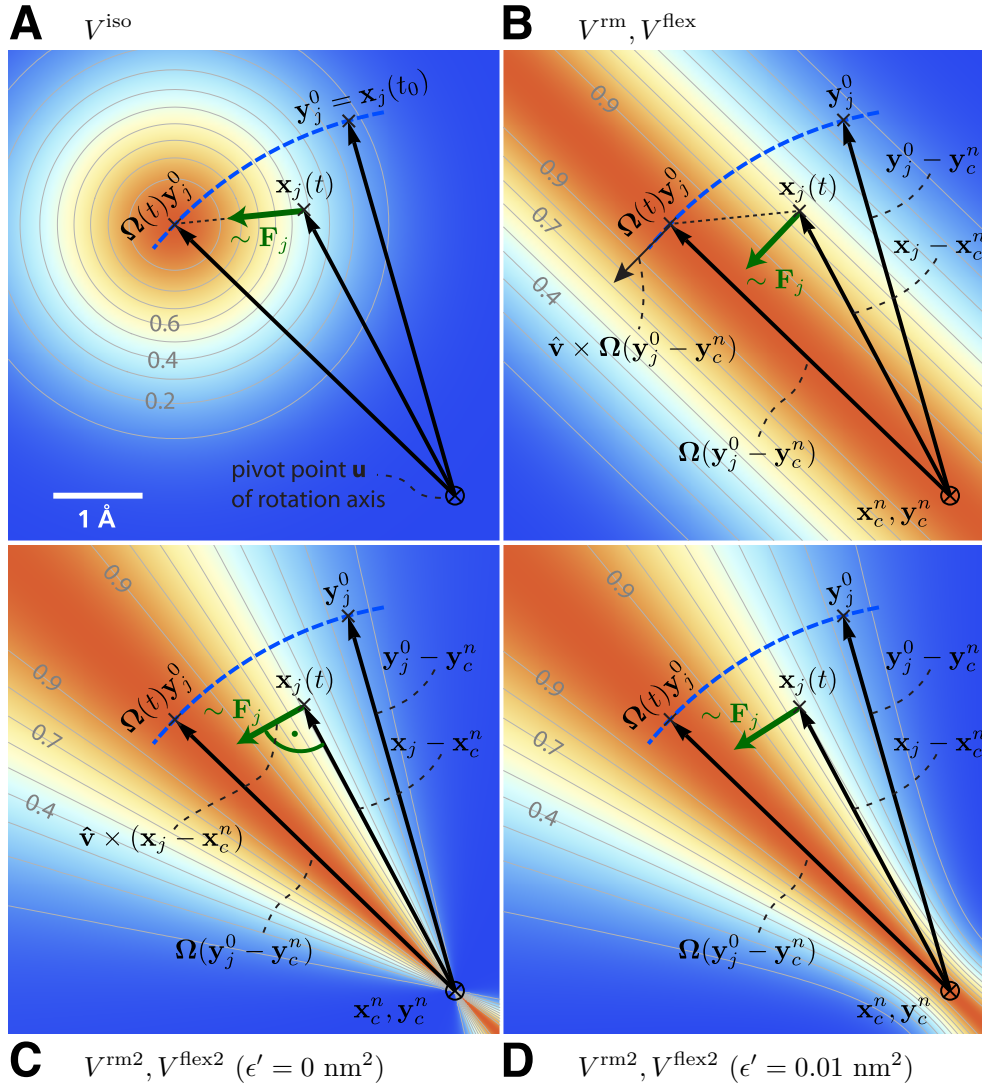


Figure 6.6: Selection of different rotation potentials and definition of notation. All four potentials V (color coded) are shown for a single atom at position $\mathbf{x}_j(t)$. **A**: Isotropic potential V^{iso} , **B**: radial motion potential V^{rm} and flexible potential V^{flex} , **C–D**: radial motion 2 potential $V^{\text{rm}2}$ and flexible 2 potential $V^{\text{flex}2}$ for $\epsilon' = 0 \text{ nm}^2$ (**C**) and $\epsilon' = 0.01 \text{ nm}^2$ (**D**). The rotation axis is perpendicular to the plane and marked by \otimes . The light gray contours indicate Boltzmann factors $e^{-V/(k_B T)}$ in the \mathbf{x}_j -plane for $T = 300 \text{ K}$ and $k = 200 \text{ kJ}/(\text{mol}\cdot\text{nm}^2)$. The green arrow shows the direction of the force \mathbf{F}_j acting on atom j ; the blue dashed line indicates the motion of the reference position.

which yields the “pivot-free” isotropic potential

$$V^{\text{iso-pf}} = \frac{k}{2} \sum_{i=1}^N w_i \left[\boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^0) - (\mathbf{x}_i - \mathbf{x}_c) \right]^2, \quad (6.36)$$

with forces

$$\mathbf{F}_j^{\text{iso-pf}} = k w_j \left[\boldsymbol{\Omega}(t)(\mathbf{y}_j^0 - \mathbf{y}_c^0) - (\mathbf{x}_j - \mathbf{x}_c) \right]. \quad (6.37)$$

Without mass-weighting, the pivot \mathbf{x}_c is the geometrical center of the group.

Parallel Motion Potential Variant

The forces generated by the isotropic potentials (eqns. 6.33 and 6.36) also contain components parallel to the rotation axis and thereby restrain motions along the axis of either the whole rotation group (in case of V^{iso}) or within the rotation group (in case of $V^{\text{iso-pf}}$). For cases where unrestrained motion along the axis is preferred, we have implemented a “parallel motion” variant by eliminating all components parallel to the rotation axis for the potential. This is achieved by projecting the distance vectors between reference and actual positions

$$\mathbf{r}_i = \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u}) - (\mathbf{x}_i - \mathbf{u}) \quad (6.38)$$

onto the plane perpendicular to the rotation vector,

$$\mathbf{r}_i^\perp := \mathbf{r}_i - (\mathbf{r}_i \cdot \hat{\mathbf{v}}) \hat{\mathbf{v}}, \quad (6.39)$$

yielding

$$\begin{aligned} V^{\text{pm}} &= \frac{k}{2} \sum_{i=1}^N w_i (\mathbf{r}_i^\perp)^2 \\ &= \frac{k}{2} \sum_{i=1}^N w_i \left\{ \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u}) - (\mathbf{x}_i - \mathbf{u}) \right. \\ &\quad \left. - \left\{ \left[\boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u}) - (\mathbf{x}_i - \mathbf{u}) \right] \cdot \hat{\mathbf{v}} \right\} \hat{\mathbf{v}} \right\}^2, \end{aligned} \quad (6.40)$$

and similarly

$$\mathbf{F}_j^{\text{pm}} = k w_j \mathbf{r}_j^\perp. \quad (6.41)$$

Pivot-Free Parallel Motion Potential

Replacing in eqn. 6.40 the fixed pivot \mathbf{u} by the center of mass \mathbf{x}_c yields the pivot-free variant of the parallel motion potential. With

$$\mathbf{s}_i = \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^0) - (\mathbf{x}_i - \mathbf{x}_c) \quad (6.42)$$

the respective potential and forces are

$$V^{\text{pm-pf}} = \frac{k}{2} \sum_{i=1}^N w_i (\mathbf{s}_i^\perp)^2, \quad (6.43)$$

$$\mathbf{F}_j^{\text{pm-pf}} = k w_j \mathbf{s}_j^\perp. \quad (6.44)$$

Radial Motion Potential

In the above variants, the minimum of the rotation potential is either a single point at the reference position \mathbf{y}_i (for the isotropic potentials) or a single line through \mathbf{y}_i parallel to the rotation axis (for the parallel motion potentials). As a result, radial forces restrict radial motions of the atoms. The two subsequent types of rotation potentials, V^{rm} and $V^{\text{rm}2}$, drastically reduce or even eliminate this effect. The first variant, V^{rm} (Fig. 6.6B), eliminates all force components parallel to the vector connecting the reference atom and the rotation axis,

$$V^{\text{rm}} = \frac{k}{2} \sum_{i=1}^N w_i [\mathbf{p}_i \cdot (\mathbf{x}_i - \mathbf{u})]^2, \quad (6.45)$$

with

$$\mathbf{p}_i := \frac{\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u})}{\|\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u})\|}. \quad (6.46)$$

This variant depends only on the distance $\mathbf{p}_i \cdot (\mathbf{x}_i - \mathbf{u})$ of atom i from the plane spanned by $\hat{\mathbf{v}}$ and $\boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u})$. The resulting force is

$$\mathbf{F}_j^{\text{rm}} = -k w_j [\mathbf{p}_j \cdot (\mathbf{x}_j - \mathbf{u})] \mathbf{p}_j. \quad (6.47)$$

Pivot-Free Radial Motion Potential

Proceeding similar to the pivot-free isotropic potential yields a pivot-free version of the above potential. With

$$\mathbf{q}_i := \frac{\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^0)}{\|\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^0)\|}, \quad (6.48)$$

the potential and force for the pivot-free variant of the radial motion potential read

$$V^{\text{rm-pf}} = \frac{k}{2} \sum_{i=1}^N w_i [\mathbf{q}_i \cdot (\mathbf{x}_i - \mathbf{x}_c)]^2, \quad (6.49)$$

$$\mathbf{F}_j^{\text{rm-pf}} = -k w_j [\mathbf{q}_j \cdot (\mathbf{x}_j - \mathbf{x}_c)] \mathbf{q}_j + k \frac{m_j}{M} \sum_{i=1}^N w_i [\mathbf{q}_i \cdot (\mathbf{x}_i - \mathbf{x}_c)] \mathbf{q}_i. \quad (6.50)$$

Radial Motion 2 Alternative Potential

As seen in Fig. 6.6B, the force resulting from V^{rm} still contains a small, second-order radial component. In most cases, this perturbation is tolerable; if not, the following alternative, $V^{\text{rm}2}$, fully eliminates the radial contribution to the force, as depicted in Fig. 6.6C,

$$V^{\text{rm}2} = \frac{k}{2} \sum_{i=1}^N w_i \frac{[(\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{u})) \cdot \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u})]^2}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{u})\|^2 + \epsilon'}, \quad (6.51)$$

where a small parameter ϵ' has been introduced to avoid singularities. For $\epsilon' = 0 \text{ nm}^2$, the equipotential planes are spanned by $\mathbf{x}_i - \mathbf{u}$ and $\hat{\mathbf{v}}$, yielding a force perpendicular to $\mathbf{x}_i - \mathbf{u}$, thus not contracting or expanding structural parts that moved away from or toward the rotation axis.

Choosing a small positive ϵ' (e.g., $\epsilon' = 0.01 \text{ nm}^2$, Fig. 6.6D) in the denominator of eqn. 6.51 yields a well-defined potential and continuous forces also close to the rotation axis, which is not the case for $\epsilon' = 0 \text{ nm}^2$ (Fig. 6.6C). With

$$\mathbf{r}_i := \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{u}) \quad (6.52)$$

$$\mathbf{s}_i := \frac{\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{u})}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{u})\|} \equiv \Psi_i \hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{u}) \quad (6.53)$$

$$\Psi_i^* := \frac{1}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{u})\|^2 + \epsilon'} \quad (6.54)$$

the force on atom j reads

$$\mathbf{F}_j^{\text{rm}2} = -k \left\{ w_j (\mathbf{s}_j \cdot \mathbf{r}_j) \left[\frac{\Psi_j^*}{\Psi_j} \mathbf{r}_j - \frac{\Psi_j^{*2}}{\Psi_j^3} (\mathbf{s}_j \cdot \mathbf{r}_j) \mathbf{s}_j \right] \right\} \times \hat{\mathbf{v}}. \quad (6.55)$$

Pivot-Free Radial Motion 2 Potential

The pivot-free variant of the above potential is

$$V^{\text{rm}2\text{-pf}} = \frac{k}{2} \sum_{i=1}^N w_i \frac{[(\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c)) \cdot \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c)]^2}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c)\|^2 + \epsilon'}. \quad (6.56)$$

With

$$\mathbf{r}_i := \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c) \quad (6.57)$$

$$\mathbf{s}_i := \frac{\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c)}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c)\|} \equiv \Psi_i \hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c) \quad (6.58)$$

$$\Psi_i^* := \frac{1}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c)\|^2 + \epsilon'} \quad (6.59)$$

the force on atom j reads

$$\begin{aligned} \mathbf{F}_j^{\text{rm}2\text{-pf}} = & -k \left\{ w_j (\mathbf{s}_j \cdot \mathbf{r}_j) \left[\frac{\Psi_j^*}{\Psi_j} \mathbf{r}_j - \frac{\Psi_j^{*2}}{\Psi_j^3} (\mathbf{s}_j \cdot \mathbf{r}_j) \mathbf{s}_j \right] \right\} \times \hat{\mathbf{v}} \\ & + k \frac{m_j}{M} \left\{ \sum_{i=1}^N w_i (\mathbf{s}_i \cdot \mathbf{r}_i) \left[\frac{\Psi_i^*}{\Psi_i} \mathbf{r}_i - \frac{\Psi_i^{*2}}{\Psi_i^3} (\mathbf{s}_i \cdot \mathbf{r}_i) \mathbf{s}_i \right] \right\} \times \hat{\mathbf{v}}. \end{aligned} \quad (6.60)$$

6.6.2 Flexible Axis Rotation

As sketched in Fig. 6.5A–B, the rigid body behavior of the fixed axis rotation scheme is a drawback for many applications. In particular, deformations of the rotation group are suppressed when the equilibrium atom positions directly depend on the reference positions. To avoid this limitation, eqns. 6.50 and 6.56 will now be generalized towards a “flexible axis” as sketched in Fig. 6.5C. This will be achieved by subdividing the rotation group into a set of equidistant slabs perpendicular to the rotation vector, and by applying a separate rotation potential to each of these slabs. Fig. 6.5C shows the midplanes of the slabs as dotted straight lines and the centers as thick black dots.

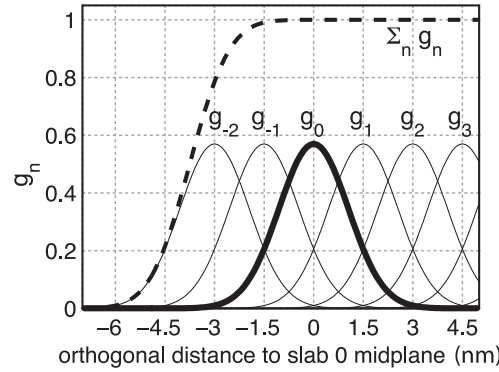


Figure 6.7: Gaussian functions g_n centered at $n \Delta x$ for a slab distance $\Delta x = 1.5$ nm and $n \geq -2$. Gaussian function g_0 is highlighted in bold; the dashed line depicts the sum of the shown Gaussian functions.

To avoid discontinuities in the potential and in the forces, we define “soft slabs” by weighing the contributions of each slab n to the total potential function V^{flex} by a Gaussian function

$$g_n(\mathbf{x}_i) = \Gamma \exp\left(-\frac{\beta_n^2(\mathbf{x}_i)}{2\sigma^2}\right), \quad (6.61)$$

centered at the midplane of the n th slab. Here σ is the width of the Gaussian function, Δx the distance between adjacent slabs, and

$$\beta_n(\mathbf{x}_i) := \mathbf{x}_i \cdot \hat{\mathbf{v}} - n \Delta x. \quad (6.62)$$

A most convenient choice is $\sigma = 0.7\Delta x$ and

$$1/\Gamma = \sum_{n \in \mathbb{Z}} \exp\left(-\frac{(n - \frac{1}{4})^2}{2 \cdot 0.7^2}\right) \approx 1.75464,$$

which yields a nearly constant sum, essentially independent of \mathbf{x}_i (dashed line in Fig. 6.7), *i.e.*,

$$\sum_{n \in \mathbb{Z}} g_n(\mathbf{x}_i) = 1 + \epsilon(\mathbf{x}_i), \quad (6.63)$$

with $|\epsilon(\mathbf{x}_i)| < 1.3 \cdot 10^{-4}$. This choice also implies that the individual contributions to the force from the slabs add up to unity such that no further normalization is required.

To each slab center \mathbf{x}_c^n , all atoms contribute by their Gaussian-weighted (optionally also mass-weighted) position vectors $g_n(\mathbf{x}_i) \mathbf{x}_i$. The instantaneous slab centers \mathbf{x}_c^n are calculated from the current positions \mathbf{x}_i ,

$$\mathbf{x}_c^n = \frac{\sum_{i=1}^N g_n(\mathbf{x}_i) m_i \mathbf{x}_i}{\sum_{i=1}^N g_n(\mathbf{x}_i) m_i}, \quad (6.64)$$

while the reference centers \mathbf{y}_c^n are calculated from the reference positions \mathbf{y}_i^0 ,

$$\mathbf{y}_c^n = \frac{\sum_{i=1}^N g_n(\mathbf{y}_i^0) m_i \mathbf{y}_i^0}{\sum_{i=1}^N g_n(\mathbf{y}_i^0) m_i}. \quad (6.65)$$

Due to the rapid decay of g_n , each slab will essentially involve contributions from atoms located within $\approx 3\Delta x$ from the slab center only.

Flexible Axis Potential

We consider two flexible axis variants. For the first variant, the slab segmentation procedure with Gaussian weighting is applied to the radial motion potential (eqn. 6.50 / Fig. 6.6B), yielding as the contribution of slab n

$$V^n = \frac{k}{2} \sum_{i=1}^N w_i g_n(\mathbf{x}_i) [\mathbf{q}_i^n \cdot (\mathbf{x}_i - \mathbf{x}_c^n)]^2,$$

and a total potential function

$$V^{\text{flex}} = \sum_n V^n. \quad (6.66)$$

Note that the global center of mass \mathbf{x}_c used in eqn. 6.50 is now replaced by \mathbf{x}_c^n , the center of mass of the slab. With

$$\mathbf{q}_i^n := \frac{\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^n)}{\|\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^n)\|} \quad (6.67)$$

$$b_i^n := \mathbf{q}_i^n \cdot (\mathbf{x}_i - \mathbf{x}_c^n), \quad (6.68)$$

the resulting force on atom j reads

$$\begin{aligned} \mathbf{F}_j^{\text{flex}} = & -k w_j \sum_n g_n(\mathbf{x}_j) b_j^n \left\{ \mathbf{q}_j^n - b_j^n \frac{\beta_n(\mathbf{x}_j)}{2\sigma^2} \hat{\mathbf{v}} \right\} \\ & + k m_j \sum_n \frac{g_n(\mathbf{x}_j)}{\sum_h g_n(\mathbf{x}_h)} \sum_{i=1}^N w_i g_n(\mathbf{x}_i) b_i^n \left\{ \mathbf{q}_i^n - \frac{\beta_n(\mathbf{x}_j)}{\sigma^2} [\mathbf{q}_i^n \cdot (\mathbf{x}_j - \mathbf{x}_c^n)] \hat{\mathbf{v}} \right\}. \end{aligned} \quad (6.69)$$

Note that for V^{flex} , as defined, the slabs are fixed in space and so are the reference centers \mathbf{y}_c^n . If during the simulation the rotation group moves too far in \mathbf{v} direction, it may enter a region where – due to the lack of nearby reference positions – no reference slab centers are defined, rendering the potential evaluation impossible. We therefore have included a slightly modified version of this potential that avoids this problem by attaching the midplane of slab $n = 0$ to the center of mass of the rotation group, yielding slabs that move with the rotation group. This is achieved by subtracting the center of mass \mathbf{x}_c of the group from the positions,

$$\tilde{\mathbf{x}}_i = \mathbf{x}_i - \mathbf{x}_c, \quad \text{and} \quad \tilde{\mathbf{y}}_i^0 = \mathbf{y}_i^0 - \mathbf{y}_c^0, \quad (6.70)$$

such that

$$V^{\text{flex-t}} = \frac{k}{2} \sum_n \sum_{i=1}^N w_i g_n(\tilde{\mathbf{x}}_i) \left[\frac{\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\tilde{\mathbf{y}}_i^0 - \tilde{\mathbf{y}}_c^n)}{\|\hat{\mathbf{v}} \times \boldsymbol{\Omega}(t)(\tilde{\mathbf{y}}_i^0 - \tilde{\mathbf{y}}_c^n)\|} \cdot (\tilde{\mathbf{x}}_i - \tilde{\mathbf{x}}_c^n) \right]^2. \quad (6.71)$$

To simplify the force derivation, and for efficiency reasons, we here assume \mathbf{x}_c to be constant, and thus $\partial \mathbf{x}_c / \partial x = \partial \mathbf{x}_c / \partial y = \partial \mathbf{x}_c / \partial z = 0$. The resulting force error is small (of order $O(1/N)$ or $O(m_j/M)$ if mass-weighting is applied) and can therefore be tolerated. With this assumption, the forces $\mathbf{F}^{\text{flex-t}}$ have the same form as eqn. 6.69.

Flexible Axis 2 Alternative Potential

In this second variant, slab segmentation is applied to $V^{\text{rm}2}$ (eqn. 6.56), resulting in a flexible axis potential without radial force contributions (Fig. 6.6C),

$$V^{\text{flex}2} = \frac{k}{2} \sum_{i=1}^N \sum_n w_i g_n(\mathbf{x}_i) \frac{[(\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c^n)) \cdot \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^n)]^2}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c^n)\|^2 + \epsilon'}. \quad (6.72)$$

With

$$\mathbf{r}_i^n := \boldsymbol{\Omega}(t)(\mathbf{y}_i^0 - \mathbf{y}_c^n) \quad (6.73)$$

$$\mathbf{s}_i^n := \frac{\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c^n)}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c^n)\|} \equiv \psi_i \hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c^n) \quad (6.74)$$

$$\psi_i^* := \frac{1}{\|\hat{\mathbf{v}} \times (\mathbf{x}_i - \mathbf{x}_c^n)\|^2 + \epsilon'} \quad (6.75)$$

$$W_j^n := \frac{g_n(\mathbf{x}_j) m_j}{\sum_h g_n(\mathbf{x}_h) m_h} \quad (6.76)$$

$$\mathbf{S}^n := \sum_{i=1}^N w_i g_n(\mathbf{x}_i) (\mathbf{s}_i^n \cdot \mathbf{r}_i^n) \left[\frac{\psi_i^*}{\psi_i} \mathbf{r}_i^n - \frac{\psi_i^{*2}}{\psi_i^3} (\mathbf{s}_i^n \cdot \mathbf{r}_i^n) \mathbf{s}_i^n \right] \quad (6.77)$$

the force on atom j reads

$$\begin{aligned} \mathbf{F}_j^{\text{flex}2} &= -k \left\{ \sum_n w_j g_n(\mathbf{x}_j) (\mathbf{s}_j^n \cdot \mathbf{r}_j^n) \left[\frac{\psi_j^*}{\psi_j} \mathbf{r}_j^n - \frac{\psi_j^{*2}}{\psi_j^3} (\mathbf{s}_j^n \cdot \mathbf{r}_j^n) \mathbf{s}_j^n \right] \right\} \times \hat{\mathbf{v}} \\ &+ k \left\{ \sum_n W_j^n \mathbf{S}^n \right\} \times \hat{\mathbf{v}} - k \left\{ \sum_n W_j^n \frac{\beta_n(\mathbf{x}_j)}{\sigma^2} \frac{1}{\psi_j} \mathbf{s}_j^n \cdot \mathbf{S}^n \right\} \hat{\mathbf{v}} \\ &+ \frac{k}{2} \left\{ \sum_n w_j g_n(\mathbf{x}_j) \frac{\beta_n(\mathbf{x}_j)}{\sigma^2} \frac{\psi_j^*}{\psi_j^2} (\mathbf{s}_j^n \cdot \mathbf{r}_j^n)^2 \right\} \hat{\mathbf{v}}. \end{aligned} \quad (6.78)$$

Applying transformation (6.70) yields a “translation-tolerant” version of the flexible 2 potential, $V^{\text{flex}2\text{-t}}$. Again, assuming that $\partial \mathbf{x}_c / \partial x$, $\partial \mathbf{x}_c / \partial y$, $\partial \mathbf{x}_c / \partial z$ are small, the resulting equations for $V^{\text{flex}2\text{-t}}$ and $\mathbf{F}^{\text{flex}2\text{-t}}$ are similar to those of $V^{\text{flex}2}$ and $\mathbf{F}^{\text{flex}2}$.

6.6.3 Usage

To apply enforced rotation, the particles i that are to be subjected to one of the rotation potentials are defined via index groups `rot-group0`, `rot-group1`, etc., in the `.mdp` input file. The reference positions \mathbf{y}_i^0 are read from a special `.trr` file provided to `grompp`. If no such file is found, $\mathbf{x}_i(t=0)$ are used as reference positions and written to `.trr` such that they can be used for subsequent setups. All parameters of the potentials such as k , ϵ' , etc. (Table 6.1) are provided as `.mdp` parameters; `rot-type` selects the type of the potential. The option `rot-massw` allows to choose whether or not to use mass-weighted averaging. For the flexible potentials, a cutoff value g_n^{min} (typically $g_n^{\text{min}} = 0.001$) makes shure that only significant contributions to V and \mathbf{F} are evaluated, *i.e.* terms with $g_n(\mathbf{x}) < g_n^{\text{min}}$ are omitted. Table 6.2 summarizes observables that are written to additional output files and which are described below.

Table 6.1: Parameters used by the various rotation potentials. x's indicate which parameter is actually used for a given potential.

parameter			k	\hat{v}	\mathbf{u}	ω	ϵ'	Δx	g_n^{\min}
.mdp input variable name			k	vec	pivot	rate	eps	slab-dist	min-gauss
unit			$\frac{\text{kJ}}{\text{mol}\cdot\text{nm}^2}$	-	nm	$^\circ/\text{ps}$	nm^2	nm	-
fixed axis potentials:		eqn.							
isotropic	V^{iso}	(6.33)	X	X	X	X	-	-	-
— pivot-free	$V^{\text{iso-pf}}$	(6.36)	X	X	-	X	-	-	-
parallel motion	V^{pm}	(6.40)	X	X	X	X	-	-	-
— pivot-free	$V^{\text{pm-pf}}$	(6.44)	X	X	-	X	-	-	-
radial motion	V^{rm}	(6.45)	X	X	X	X	-	-	-
— pivot-free	$V^{\text{rm-pf}}$	(6.50)	X	X	-	X	-	-	-
radial motion 2	V^{rm2}	(6.51)	X	X	X	X	X	-	-
— pivot-free	$V^{\text{rm2-pf}}$	(6.56)	X	X	-	X	X	-	-
flexible axis potentials:		eqn.							
flexible	V^{flex}	(6.66)	X	X	-	X	-	X	X
— transl. tol.	$V^{\text{flex-t}}$	(6.71)	X	X	-	X	-	X	X
flexible 2	V^{flex2}	(6.72)	X	X	-	X	X	X	X
— transl. tol.	$V^{\text{flex2-t}}$	-	X	X	-	X	X	X	X

Table 6.2: Quantities recorded in output files during enforced rotation simulations. All slab-wise data is written every `nstfout` steps, other rotation data every `nstrout` steps.

quantity	unit	equation	output file	fixed	flexible
$V(t)$	kJ/mol	see 6.1	rotation	X	X
$\theta_{\text{ref}}(t)$	degrees	$\theta_{\text{ref}}(t) = \omega t$	rotation	X	X
$\theta_{\text{av}}(t)$	degrees	(6.79)	rotation	X	-
$\theta_{\text{fit}}(t), \theta_{\text{fit}}(t, n)$	degrees	(6.81)	rotangles	-	X
$\mathbf{y}_0(n), \mathbf{x}_0(t, n)$	nm	(6.64, 6.65)	rotslabs	-	X
$\tau(t)$	kJ/mol	(6.82)	rotation	X	-
$\tau(t, n)$	kJ/mol	(6.82)	rottorque	-	X

Angle of Rotation Groups: Fixed Axis

For fixed axis rotation, the average angle $\theta_{\text{av}}(t)$ of the group relative to the reference group is determined via the distance-weighted angular deviation of all rotation group atoms from their reference positions,

$$\theta_{\text{av}} = \frac{\sum_{i=1}^N r_i \theta_i}{\sum_{i=1}^N r_i}. \quad (6.79)$$

Here, r_i is the distance of the reference position to the rotation axis, and the difference angles θ_i are determined from the atomic positions, projected onto a plane perpendicular to the rotation axis through pivot point \mathbf{u} (see eqn. 6.39 for the definition of \perp),

$$\cos \theta_i = \frac{(\mathbf{y}_i - \mathbf{u})^\perp \cdot (\mathbf{x}_i - \mathbf{u})^\perp}{\|(\mathbf{y}_i - \mathbf{u})^\perp \cdot (\mathbf{x}_i - \mathbf{u})^\perp\|}. \quad (6.80)$$

The sign of θ_{av} is chosen such that $\theta_{\text{av}} > 0$ if the actual structure rotates ahead of the reference.

Angle of Rotation Groups: Flexible Axis

For flexible axis rotation, two outputs are provided, the angle of the entire rotation group, and separate angles for the segments in the slabs. The angle of the entire rotation group is determined by an RMSD fit of \mathbf{x}_i to the reference positions \mathbf{y}_i^0 at $t = 0$, yielding θ_{fit} as the angle by which the reference has to be rotated around $\hat{\mathbf{v}}$ for the optimal fit,

$$\text{RMSD}(\mathbf{x}_i, \mathbf{\Omega}(\theta_{\text{fit}})\mathbf{y}_i^0) \stackrel{!}{=} \min. \quad (6.81)$$

To determine the local angle for each slab n , both reference and actual positions are weighted with the Gaussian function of slab n , and $\theta_{\text{fit}}(t, n)$ is calculated as in eqn. 6.81) from the Gaussian-weighted positions.

For all angles, the `.mdp` input option `rot-fit-method` controls whether a normal RMSD fit is performed or whether for the fit each position \mathbf{x}_i is put at the same distance to the rotation axis as its reference counterpart \mathbf{y}_i^0 . In the latter case, the RMSD measures only angular differences, not radial ones.

Angle Determination by Searching the Energy Minimum

Alternatively, for `rot-fit-method = potential`, the angle of the rotation group is determined as the angle for which the rotation potential energy is minimal. Therefore, the used rotation potential is additionally evaluated for a set of angles around the current reference angle. In this case, the `rotangles.log` output file contains the values of the rotation potential at the chosen set of angles, while `rotation.svg` lists the angle with minimal potential energy.

Torque

The torque $\boldsymbol{\tau}(t)$ exerted by the rotation potential is calculated for fixed axis rotation via

$$\boldsymbol{\tau}(t) = \sum_{i=1}^N \mathbf{r}_i(t) \times \mathbf{f}_i^\perp(t), \quad (6.82)$$

where $\mathbf{r}_i(t)$ is the distance vector from the rotation axis to $\mathbf{x}_i(t)$ and $\mathbf{f}_i^\perp(t)$ is the force component perpendicular to $\mathbf{r}_i(t)$ and $\hat{\mathbf{v}}$. For flexible axis rotation, torques $\boldsymbol{\tau}_n$ are calculated for each slab using the local rotation axis of the slab and the Gaussian-weighted positions.

6.7 Electric fields

A pulsed and oscillating electric field can be applied according to:

$$E(t) = E_0 \exp\left[-\frac{(t-t_0)^2}{2\sigma^2}\right] \cos[\omega(t-t_0)] \quad (6.83)$$

where E_0 is the field strength, the angular frequency $\omega = 2\pi c/\lambda$, t_0 is the time at of the peak in the field strength and σ is the with of the pulse. Special cases occur when $\sigma = 0$ (non-pulsed field) and for ω is 0 (static field).

This simulated laser-pulse was applied to simulations of melting ice [151]. A pulsed electric field may look like Fig. 6.8. In the supporting information of that paper the impact of an applied electric field on a system under periodic boundary conditions is analyzed. It is described that the effective electric field under PBC is larger than the applied field, by a factor depending on the size of the box and the dielectric properties of molecules in the box. For a system with static dielectric properties this factor can be corrected for. But for a system where the dielectric varies over time, for example a membrane protein with a pore that opens and closes during the simulatippn, this way of applying an electric field is not useful. In such cases one can use the computational electrophysiology protocol described in the next section (sec. 6.8).

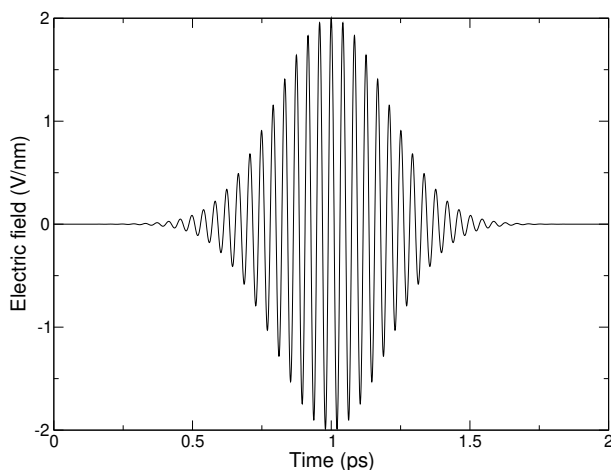


Figure 6.8: A simulated laser pulse in GROMACS.

Electric fields are applied when the following options are specified in the `grompp.mdp` file. You specify, in order, E_0 , ω , t_0 and σ :

```
electric-field-x = 0.04 0 0 0
```

yields a static field with $E_0 = 0.04$ V/nm in the X-direction. In contrast,

```
electric-field-x = 2.0 150 5 0
```

yields an oscillating electric field with $E_0 = 2$ V/nm, $\omega = 150$ /ps and $t_0 = 5$ ps. Finally

```
electric-field-x = 2.0 150 5 1
```

yields an pulsed-oscillating electric field with $E_0 = 2$ V/nm, $\omega = 150$ /ps and $t_0 = 5$ ps and $\sigma = 1$ ps. Read more in ref. [151]. Note that the input file format is changed from the undocumented older version. A figure like Fig. 6.8 may be produced by passing the `-field` option to `gmx mdrun`.

6.8 Computational Electrophysiology

The Computational Electrophysiology (CompEL) protocol [152] allows the simulation of ion flux through membrane channels, driven by transmembrane potentials or ion concentration gradients. Just as in real cells, CompEL establishes transmembrane potentials by sustaining a small imbalance of charges Δq across the membrane, which gives rise to a potential difference ΔU according to the membrane capacitance:

$$\Delta U = \Delta q / C_{\text{membrane}} \quad (6.84)$$

The transmembrane electric field and concentration gradients are controlled by `.mdp` options, which allow the user to set reference counts for the ions on either side of the membrane. If a difference between the actual and the reference numbers persists over a certain time span, specified by the user, a number of ion/water pairs are exchanged between the compartments until the reference numbers are restored. Alongside the calculation of channel conductance and ion selectivity, CompEL simulations also enable determination of the channel reversal potential, an important characteristic obtained in electrophysiology experiments.

In a CompEL setup, the simulation system is divided into two compartments **A** and **B** with independent ion concentrations. This is best achieved by using double bilayer systems with a copy (or copies) of the channel/pore of interest in each bilayer (Fig. 6.9 A, B). If the channel axes point in the same direction, channel flux is observed simultaneously at positive and negative potentials in this way, which is for instance important for studying channel rectification.

The potential difference ΔU across the membrane is easily calculated with the `gmx potential` utility. By this, the potential drop along z or the pore axis is exactly known in each time interval of the simulation (Fig. 6.9 C). Type and number of ions n_i of charge q_i , traversing the channel in the simulation, are written to the `swapions.xvg` output file, from which the average channel conductance G in each interval Δt is determined by:

$$G = \frac{\sum_i n_i q_i}{\Delta t \Delta U}. \quad (6.85)$$

The ion selectivity is calculated as the number flux ratio of different species. Best results are obtained by averaging these values over several overlapping time intervals.

The calculation of reversal potentials is best achieved using a small set of simulations in which a given transmembrane concentration gradient is complemented with small ion imbalances of varying magnitude. For example, if one compartment contains 1 M salt and the other 0.1 M, and

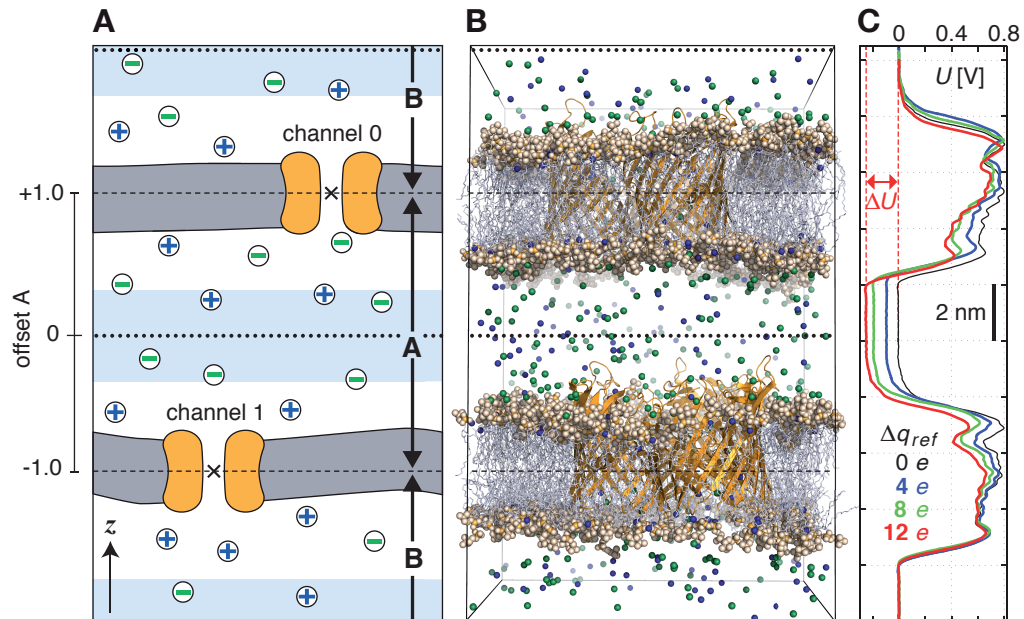


Figure 6.9: Typical double-membrane setup for CompEL simulations (A, B). Ion / water molecule exchanges will be performed as needed between the two light blue volumes around the dotted black lines (A). Plot (C) shows the potential difference ΔU resulting from the selected charge imbalance Δq_{ref} between the compartments.

given charge neutrality otherwise, a set of simulations with $\Delta q = 0 e$, $\Delta q = 2 e$, $\Delta q = 4 e$ could be used. Fitting a straight line through the current-voltage relationship of all obtained I - U pairs near zero current will then yield U_{rev} .

6.8.1 Usage

The following `.mdp` options control the CompEL protocol:

```
swapcoords      = Z           ; Swap positions: no, X, Y, Z
swap-frequency  = 100        ; Swap attempt frequency
```

Choose `Z` if your membrane is in the xy -plane (Fig. 6.9). Ions will be exchanged between compartments depending on their z -positions alone. `swap-frequency` determines how often a swap attempt will be made. This step requires that the positions of the split groups, the ions, and possibly the solvent molecules are communicated between the parallel processes, so if chosen too small it can decrease the simulation performance. The `Position swapping` entry in the cycle and time accounting table at the end of the `md.log` file summarizes the amount of runtime spent in the swap module.

```
split-group0    = channel0   ; Defines compartment boundary
split-group1    = channel1   ; Defines other compartment boundary
massw-split0    = no         ; use mass-weighted center?
massw-split1    = no
```

`split-group0` and `split-group1` are two index groups that define the boundaries between the two compartments, which are usually the centers of the channels. If `massw-split0` or `massw-split1` are set to `yes`, the center of mass of each index group is used as boundary, here in z -direction. Otherwise, the geometrical centers will be used (\times in Fig. 6.9 A). If, such as here, a membrane channel is selected as split group, the center of the channel will define the dividing plane between the compartments (dashed horizontal lines). All index groups must be defined in the index file.

If, to restore the requested ion counts, an ion from one compartment has to be exchanged with a water molecule from the other compartment, then those molecules are swapped which have the largest distance to the compartment-defining boundaries (dashed horizontal lines). Depending on the ion concentration, this effectively results in exchanges of molecules between the light blue volumes. If a channel is very asymmetric in z -direction and would extend into one of the swap volumes, one can offset the swap exchange plane with the `bulk-offset` parameter. A value of 0.0 means no offset b , values $-1.0 < b < 0$ move the swap exchange plane closer to the lower, values $0 < b < 1.0$ closer to the upper membrane. Fig. 6.9 A (left) depicts that for the A compartment.

```
solvent-group = SOL      ; Group containing the solvent molecules
iontypes      = 3       ; Number of different ion types to control
iontype0-name = NA      ; Group name of the ion type
iontype0-in-A = 51      ; Reference count of ions of type 0 in A
iontype0-in-B = 35      ; Reference count of ions of type 0 in B
iontype1-name = K       ;
iontype1-in-A = 10      ;
iontype1-in-B = 38      ;
iontype2-name = CL      ;
iontype2-in-A = -1      ;
iontype2-in-B = -1      ;
```

The group name of solvent molecules acting as exchange partners for the ions has to be set with `solvent-group`. The number of different ionic species under control of the CompEL protocol is given by the `iontypes` parameter, while `iontype0-name` gives the name of the index group containing the atoms of this ionic species. The reference number of ions of this type can be set with the `iontype0-in-A` and `iontype0-in-B` options for compartments **A** and **B**, respectively. Obviously, the sum of `iontype0-in-A` and `iontype0-in-B` needs to equal the number of ions in the group defined by `iontype0-name`. A reference number of -1 means: use the number of ions as found at the beginning of the simulation as the reference value.

```
coupl-steps   = 10      ; Average over these many swap steps
threshold     = 1       ; Do not swap if < threshold
```

If `coupl-steps` is set to 1, then the momentary ion distribution determines whether ions are exchanged. `coupl-steps > 1` will use the time-average of ion distributions over the selected number of attempt steps instead. This can be useful, for example, when ions diffuse near compartment boundaries, which would lead to numerous unproductive ion exchanges. A `threshold` of 1 means that a swap is performed if the average ion count in a compartment differs by at least 1

from the requested values. Higher thresholds will lead to toleration of larger differences. Ions are exchanged until the requested number \pm the threshold is reached.

```

cyl0-r           = 5.0           ; Split cylinder 0 radius (nm)
cyl0-up          = 0.75          ; Split cylinder 0 upper extension (nm)
cyl0-down        = 0.75          ; Split cylinder 0 lower extension (nm)
cyl1-r           = 5.0           ; same for other channel
cyl1-up          = 0.75
cyl1-down        = 0.75

```

The cylinder options are used to define virtual geometric cylinders around the channel's pore to track how many ions of which type have passed each channel. Ions will be counted as having traveled through a channel according to the definition of the channel's cylinder radius, upper and lower extension, relative to the location of the respective split group. This will not affect the actual flux or exchange, but will provide you with the ion permeation numbers across each of the channels. Note that an ion can only be counted as passing through a particular channel if it is detected *within* the defined split cylinder in a swap step. If `swap-frequency` is chosen too high, a particular ion may be detected in compartment **A** in one swap step, and in compartment **B** in the following swap step, so it will be unclear through which of the channels it has passed.

A double-layered system for CompEL simulations can be easily prepared by duplicating an existing membrane/channel MD system in the direction of the membrane normal (typically z) with `gmx editconf -translate 0 0 <l_z>`, where `l_z` is the box length in that direction. If you have already defined index groups for the channel for the single-layered system, `gmx make_ndx -n index.ndx -twin` will provide you with the groups for the double-layered system.

To suppress large fluctuations of the membranes along the swap direction, it may be useful to apply a harmonic potential (acting only in the swap dimension) between each of the two channel and/or bilayer centers using umbrella pulling (see section 6.4).

Multimeric channels

If a split group consists of more than one molecule, the correct PBC image of all molecules with respect to each other has to be chosen such that the channel center can be correctly determined. GROMACS assumes that the starting structure in the `.tpr` file has the correct PBC representation. Set the following environment variable to check whether that is the case:

- `GMX_COMPELDUMP`: output the starting structure after it has been made whole to `.pdb` file.

6.9 Calculating a PMF using the free-energy code

The free-energy coupling-parameter approach (see sec. 3.12) provides several ways to calculate potentials of mean force. A potential of mean force between two atoms can be calculated by connecting them with a harmonic potential or a constraint. For this purpose there are special potentials that avoid the generation of extra exclusions, see sec. 5.4.4. When the position of the

minimum or the constraint length is 1 nm more in state B than in state A, the restraint or constraint force is given by $\partial H/\partial\lambda$. The distance between the atoms can be changed as a function of λ and time by setting `delta-lambda` in the `.mdp` file. The results should be identical (although not numerically due to the different implementations) to the results of the pull code with umbrella sampling and constraint pulling. Unlike the pull code, the free energy code can also handle atoms that are connected by constraints.

Potentials of mean force can also be calculated using position restraints. With position restraints, atoms can be linked to a position in space with a harmonic potential (see 4.3.1). These positions can be made a function of the coupling parameter λ . The positions for the A and the B states are supplied to `grompp` with the `-r` and `-rb` options, respectively. One could use this approach to do targeted MD; note that we do not encourage the use of targeted MD for proteins. A protein can be forced from one conformation to another by using these conformations as position restraint coordinates for state A and B. One can then slowly change λ from 0 to 1. The main drawback of this approach is that the conformational freedom of the protein is severely limited by the position restraints, independent of the change from state A to B. Also, the protein is forced from state A to B in an almost straight line, whereas the real pathway might be very different. An example of a more fruitful application is a solid system or a liquid confined between walls where one wants to measure the force required to change the separation between the boundaries or walls. Because the boundaries (or walls) already need to be fixed, the position restraints do not limit the system in its sampling.

6.10 Removing fastest degrees of freedom

The maximum time step in MD simulations is limited by the smallest oscillation period that can be found in the simulated system. Bond-stretching vibrations are in their quantum-mechanical ground state and are therefore better represented by a constraint instead of a harmonic potential.

For the remaining degrees of freedom, the shortest oscillation period (as measured from a simulation) is 13 fs for bond-angle vibrations involving hydrogen atoms. Taking as a guideline that with a Verlet (leap-frog) integration scheme a minimum of 5 numerical integration steps should be performed per period of a harmonic oscillation in order to integrate it with reasonable accuracy, the maximum time step will be about 3 fs. Disregarding these very fast oscillations of period 13 fs, the next shortest periods are around 20 fs, which will allow a maximum time step of about 4 fs.

Removing the bond-angle degrees of freedom from hydrogen atoms can best be done by defining them as virtual interaction sites instead of normal atoms. Whereas a normal atom is connected to the molecule with bonds, angles and dihedrals, a virtual site's position is calculated from the position of three nearby heavy atoms in a predefined manner (see also sec. 4.7). For the hydrogens in water and in hydroxyl, sulfhydryl, or amine groups, no degrees of freedom can be removed, because rotational freedom should be preserved. The only other option available to slow down these motions is to increase the mass of the hydrogen atoms at the expense of the mass of the connected heavy atom. This will increase the moment of inertia of the water molecules and the hydroxyl, sulfhydryl, or amine groups, without affecting the equilibrium properties of the system and without affecting the dynamical properties too much. These constructions will shortly be described in sec. 6.10.1 and have previously been described in full detail [153].

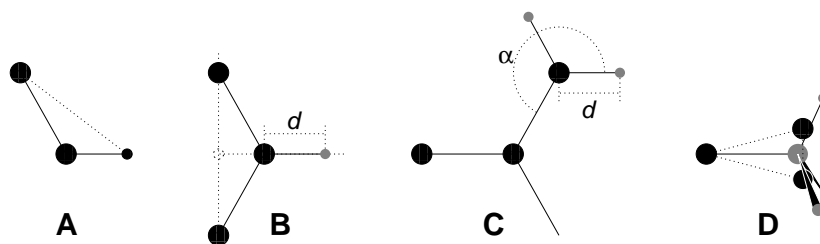


Figure 6.10: The different types of virtual site constructions used for hydrogen atoms. The atoms used in the construction of the virtual site(s) are depicted as black circles, virtual sites as gray ones. Hydrogens are smaller than heavy atoms. **A**: fixed bond angle, note that here the hydrogen is not a virtual site; **B**: in the plane of three atoms, with fixed distance; **C**: in the plane of three atoms, with fixed angle and distance; **D**: construction for amine groups ($-\text{NH}_2$ or $-\text{NH}_3^+$), see text for details.

Using both virtual sites and modified masses, the next bottleneck is likely to be formed by the improper dihedrals (which are used to preserve planarity or chirality of molecular groups) and the peptide dihedrals. The peptide dihedral cannot be changed without affecting the physical behavior of the protein. The improper dihedrals that preserve planarity mostly deal with aromatic residues. Bonds, angles, and dihedrals in these residues can also be replaced with somewhat elaborate virtual site constructions.

All modifications described in this section can be performed using the GROMACS topology building tool `pdb2gmx`. Separate options exist to increase hydrogen masses, virtualize all hydrogen atoms, or also virtualize all aromatic residues. **Note** that when all hydrogen atoms are virtualized, those inside the aromatic residues will be virtualized as well, *i.e.* hydrogens in the aromatic residues are treated differently depending on the treatment of the aromatic residues.

Parameters for the virtual site constructions for the hydrogen atoms are inferred from the force-field parameters (*vis.* bond lengths and angles) directly by `grompp` while processing the topology file. The constructions for the aromatic residues are based on the bond lengths and angles for the geometry as described in the force fields, but these parameters are hard-coded into `pdb2gmx` due to the complex nature of the construction needed for a whole aromatic group.

6.10.1 Hydrogen bond-angle vibrations

Construction of virtual sites

The goal of defining hydrogen atoms as virtual sites is to remove all high-frequency degrees of freedom from them. In some cases, not all degrees of freedom of a hydrogen atom should be removed, *e.g.* in the case of hydroxyl or amine groups the rotational freedom of the hydrogen atom(s) should be preserved. Care should be taken that no unwanted correlations are introduced by the construction of virtual sites, *e.g.* bond-angle vibration between the constructing atoms could translate into hydrogen bond-length vibration. Additionally, since virtual sites are by definition massless, in order to preserve total system mass, the mass of each hydrogen atom that is treated as virtual site should be added to the bonded heavy atom.

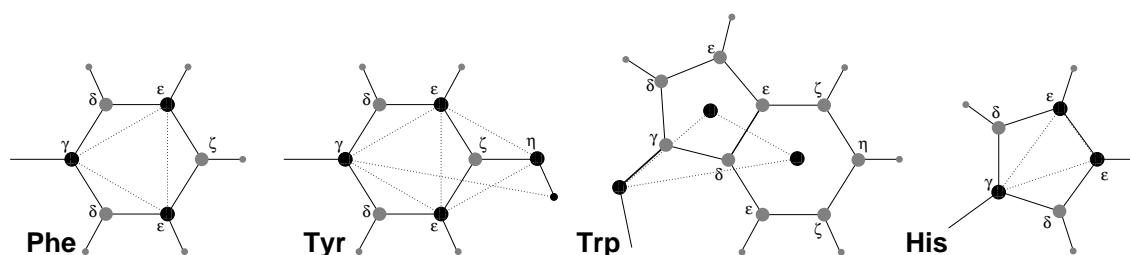


Figure 6.11: The different types of virtual site constructions used for aromatic residues. The atoms used in the construction of the virtual site(s) are depicted as black circles, virtual sites as gray ones. Hydrogens are smaller than heavy atoms. **A:** phenylalanine; **B:** tyrosine (note that the hydroxyl hydrogen is *not* a virtual site); **C:** tryptophan; **D:** histidine.

Taking into account these considerations, the hydrogen atoms in a protein naturally fall into several categories, each requiring a different approach (see also Fig. 6.10).

- *hydroxyl (-OH) or sulfhydryl (-SH) hydrogen:* The only internal degree of freedom in a hydroxyl group that can be constrained is the bending of the C-O-H angle. This angle is fixed by defining an additional bond of appropriate length, see Fig. 6.10A. Doing so removes the high-frequency angle bending, but leaves the dihedral rotational freedom. The same goes for a sulfhydryl group. **Note** that in these cases the hydrogen is not treated as a virtual site.
- *single amine or amide (-NH-) and aromatic hydrogens (-CH-):* The position of these hydrogens cannot be constructed from a linear combination of bond vectors, because of the flexibility of the angle between the heavy atoms. Instead, the hydrogen atom is positioned at a fixed distance from the bonded heavy atom on a line going through the bonded heavy atom and a point on the line through both second bonded atoms, see Fig. 6.10B.
- *planar amine (-NH₂) hydrogens:* The method used for the single amide hydrogen is not well suited for planar amine groups, because no suitable two heavy atoms can be found to define the direction of the hydrogen atoms. Instead, the hydrogen is constructed at a fixed distance from the nitrogen atom, with a fixed angle to the carbon atom, in the plane defined by one of the other heavy atoms, see Fig. 6.10C.
- *amine group (umbrella -NH₂ or -NH₃⁺) hydrogens:* Amine hydrogens with rotational freedom cannot be constructed as virtual sites from the heavy atoms they are connected to, since this would result in loss of the rotational freedom of the amine group. To preserve the rotational freedom while removing the hydrogen bond-angle degrees of freedom, two “dummy masses” are constructed with the same total mass, moment of inertia (for rotation around the C-N bond) and center of mass as the amine group. These dummy masses have no interaction with any other atom, except for the fact that they are connected to the carbon and to each other, resulting in a rigid triangle. From these three particles, the positions of the nitrogen and hydrogen atoms are constructed as linear combinations of the two carbon-mass vectors and their outer product, resulting in an amine group with rotational freedom intact, but without other internal degrees of freedom. See Fig. 6.10D.

6.10.2 Out-of-plane vibrations in aromatic groups

The planar arrangements in the side chains of the aromatic residues lends itself perfectly to a virtual-site construction, giving a perfectly planar group without the inherently unstable constraints that are necessary to keep normal atoms in a plane. The basic approach is to define three atoms or dummy masses with constraints between them to fix the geometry and create the rest of the atoms as simple virtual sites type (see sec. 4.7) from these three. Each of the aromatic residues require a different approach:

- *Phenylalanine*: C_γ , $C_{\epsilon 1}$, and $C_{\epsilon 2}$ are kept as normal atoms, but with each a mass of one third the total mass of the phenyl group. See Fig. 6.10A.
- *Tyrosine*: The ring is treated identically to the phenylalanine ring. Additionally, constraints are defined between $C_{\epsilon 1}$, $C_{\epsilon 2}$, and O_η . The original improper dihedral angles will keep both triangles (one for the ring and one with O_η) in a plane, but due to the larger moments of inertia this construction will be much more stable. The bond-angle in the hydroxyl group will be constrained by a constraint between C_γ and H_η . **Note** that the hydrogen is not treated as a virtual site. See Fig. 6.10B.
- *Tryptophan*: C_β is kept as a normal atom and two dummy masses are created at the center of mass of each of the rings, each with a mass equal to the total mass of the respective ring ($C_{\delta 2}$ and $C_{\epsilon 2}$ are each counted half for each ring). This keeps the overall center of mass and the moment of inertia almost (but not quite) equal to what it was. See Fig. 6.10C.
- *Histidine*: C_γ , $C_{\epsilon 1}$ and $N_{\epsilon 2}$ are kept as normal atoms, but with masses redistributed such that the center of mass of the ring is preserved. See Fig. 6.10D.

6.11 Viscosity calculation

The shear viscosity is a property of liquids that can be determined easily by experiment. It is useful for parameterizing a force field because it is a kinetic property, while most other properties which are used for parameterization are thermodynamic. The viscosity is also an important property, since it influences the rates of conformational changes of molecules solvated in the liquid.

The viscosity can be calculated from an equilibrium simulation using an Einstein relation:

$$\eta = \frac{1}{2} \frac{V}{k_B T} \lim_{t \rightarrow \infty} \frac{d}{dt} \left\langle \left(\int_{t_0}^{t_0+t} P_{xz}(t') dt' \right)^2 \right\rangle_{t_0} \quad (6.86)$$

This can be done with `gmx energy`. This method converges very slowly [154], and as such a nanosecond simulation might not be long enough for an accurate determination of the viscosity. The result is very dependent on the treatment of the electrostatics. Using a (short) cut-off results in large noise on the off-diagonal pressure elements, which can increase the calculated viscosity by an order of magnitude.

GROMACS also has a non-equilibrium method for determining the viscosity [154]. This makes use of the fact that energy, which is fed into system by external forces, is dissipated through viscous

friction. The generated heat is removed by coupling to a heat bath. For a Newtonian liquid adding a small force will result in a velocity gradient according to the following equation:

$$a_x(z) + \frac{\eta}{\rho} \frac{\partial^2 v_x(z)}{\partial z^2} = 0 \quad (6.87)$$

Here we have applied an acceleration $a_x(z)$ in the x -direction, which is a function of the z -coordinate. In GROMACS the acceleration profile is:

$$a_x(z) = A \cos\left(\frac{2\pi z}{l_z}\right) \quad (6.88)$$

where l_z is the height of the box. The generated velocity profile is:

$$v_x(z) = V \cos\left(\frac{2\pi z}{l_z}\right) \quad (6.89)$$

$$V = A \frac{\rho}{\eta} \left(\frac{l_z}{2\pi}\right)^2 \quad (6.90)$$

The viscosity can be calculated from A and V :

$$\eta = \frac{A}{V} \rho \left(\frac{l_z}{2\pi}\right)^2 \quad (6.91)$$

In the simulation V is defined as:

$$V = \frac{\sum_{i=1}^N m_i v_{i,x} \cos\left(\frac{2\pi z}{l_z}\right)}{\sum_{i=1}^N m_i} \quad (6.92)$$

The generated velocity profile is not coupled to the heat bath. Moreover, the velocity profile is excluded from the kinetic energy. One would like V to be as large as possible to get good statistics. However, the shear rate should not be so high that the system gets too far from equilibrium. The maximum shear rate occurs where the cosine is zero, the rate being:

$$\text{sh}_{\max} = \max_z \left| \frac{\partial v_x(z)}{\partial z} \right| = A \frac{\rho}{\eta} \frac{l_z}{2\pi} \quad (6.93)$$

For a simulation with: $\eta = 10^{-3}$ [kg m⁻¹ s⁻¹], $\rho = 10^3$ [kg m⁻³] and $l_z = 2\pi$ [nm], $\text{sh}_{\max} = 1$ [ps nm⁻¹] A . This shear rate should be smaller than one over the longest correlation time in the system. For most liquids, this will be the rotation correlation time, which is around 10 ps. In this case, A should be smaller than 0.1 [nm ps⁻²]. When the shear rate is too high, the observed viscosity will be too low. Because V is proportional to the square of the box height, the optimal box is elongated in the z -direction. In general, a simulation length of 100 ps is enough to obtain an accurate value for the viscosity.

The heat generated by the viscous friction is removed by coupling to a heat bath. Because this coupling is not instantaneous the real temperature of the liquid will be slightly lower than the

observed temperature. Berendsen derived this temperature shift [31], which can be written in terms of the shear rate as:

$$T_s = \frac{\eta \tau}{2\rho C_v} \dot{\gamma}_{\max}^2 \quad (6.94)$$

where τ is the coupling time for the Berendsen thermostat and C_v is the heat capacity. Using the values of the example above, $\tau = 10^{-13}$ [s] and $C_v = 2 \cdot 10^3$ [J kg⁻¹ K⁻¹], we get: $T_s = 25$ [K ps⁻²] $\dot{\gamma}_{\max}^2$. When we want the shear rate to be smaller than 1/10 [ps⁻¹], T_s is smaller than 0.25 [K], which is negligible.

Note that the system has to build up the velocity profile when starting from an equilibrium state. This build-up time is of the order of the correlation time of the liquid.

Two quantities are written to the energy file, along with their averages and fluctuations: V and $1/\eta$, as obtained from (6.91).

6.12 Tabulated interaction functions

6.12.1 Cubic splines for potentials

In some of the inner loops of GROMACS, look-up tables are used for computation of potential and forces. The tables are interpolated using a cubic spline algorithm. There are separate tables for electrostatic, dispersion, and repulsion interactions, but for the sake of caching performance these have been combined into a single array. The cubic spline interpolation for $x_i \leq x < x_{i+1}$ looks like this:

$$V_s(x) = A_0 + A_1 \epsilon + A_2 \epsilon^2 + A_3 \epsilon^3 \quad (6.95)$$

where the table spacing h and fraction ϵ are given by:

$$h = x_{i+1} - x_i \quad (6.96)$$

$$\epsilon = (x - x_i)/h \quad (6.97)$$

so that $0 \leq \epsilon < 1$. From this, we can calculate the derivative in order to determine the forces:

$$-V'_s(x) = -\frac{dV_s(x)}{d\epsilon} \frac{d\epsilon}{dx} = -(A_1 + 2A_2 \epsilon + 3A_3 \epsilon^2)/h \quad (6.98)$$

The four coefficients are determined from the four conditions that V_s and $-V'_s$ at both ends of each interval should match the exact potential V and force $-V'$. This results in the following errors for each interval:

$$|V_s - V|_{\max} = V'''' \frac{h^4}{384} + O(h^5) \quad (6.99)$$

$$|V'_s - V'|_{\max} = V'''' \frac{h^3}{72\sqrt{3}} + O(h^4) \quad (6.100)$$

$$|V''_s - V''|_{\max} = V'''' \frac{h^2}{12} + O(h^3) \quad (6.101)$$

V and V' are continuous, while V'' is the first discontinuous derivative. The number of points per nanometer is 500 and 2000 for mixed- and double-precision versions of GROMACS, respectively. This means that the errors in the potential and force will usually be smaller than the mixed precision accuracy.

GROMACS stores A_0 , A_1 , A_2 and A_3 . The force routines get a table with these four parameters and a scaling factor s that is equal to the number of points per nm. (**Note** that h is s^{-1}). The algorithm goes a little something like this:

1. Calculate distance vector (\mathbf{r}_{ij}) and distance r_{ij}
2. Multiply r_{ij} by s and truncate to an integer value n_0 to get a table index
3. Calculate fractional component ($\epsilon = sr_{ij} - n_0$) and ϵ^2
4. Do the interpolation to calculate the potential V and the scalar force f
5. Calculate the vector force \mathbf{F} by multiplying f with \mathbf{r}_{ij}

Note that table look-up is significantly *slower* than computation of the most simple Lennard-Jones and Coulomb interaction. However, it is much faster than the shifted Coulomb function used in conjunction with the PPPM method. Finally, it is much easier to modify a table for the potential (and get a graphical representation of it) than to modify the inner loops of the MD program.

6.12.2 User-specified potential functions

You can also use your own potential functions without editing the GROMACS code. The potential function should be according to the following equation

$$V(r_{ij}) = \frac{q_i q_j}{4\pi\epsilon_0} f(r_{ij}) + C_6 g(r_{ij}) + C_{12} h(r_{ij}) \quad (6.102)$$

where f , g , and h are user defined functions. **Note** that if $g(r)$ represents a normal dispersion interaction, $g(r)$ should be < 0 . C_6 , C_{12} and the charges are read from the topology. Also note that combination rules are only supported for Lennard-Jones and Buckingham, and that your tables should match the parameters in the binary topology.

When you add the following lines in your `.mdp` file:

```
rlist           = 1.0
coulombtype    = User
rcoulomb       = 1.0
vdwtype        = User
rvdw           = 1.0
```

`mdrun` will read a single non-bonded table file, or multiple when `energygrp-table` is set (see below). The name of the file(s) can be set with the `mdrun` option `-table`. The table file should contain seven columns of table look-up data in the order: x , $f(x)$, $-f'(x)$, $g(x)$, $-g'(x)$, $h(x)$, $-h'(x)$. The x should run from 0 to $r_c + 1$ (the value of `table_extension` can be changed in the `.mdp` file). You can choose the spacing you like; for the standard tables GROMACS uses a spacing of 0.002 and 0.0005 nm when you run in mixed and double precision, respectively. In this context, r_c denotes the maximum of the two cut-offs `rvdw` and `rcoulomb` (see above). These variables need not be the same (and need not be 1.0 either). Some functions used for potentials contain a singularity at $x = 0$, but since atoms are normally not closer to each other than 0.1 nm,

the function value at $x = 0$ is not important. Finally, it is also possible to combine a standard Coulomb with a modified LJ potential (or vice versa). One then specifies *e.g.* `coulombtype = Cut-off` or `coulombtype = PME`, combined with `vdwtype = User`. The table file must always contain the 7 columns however, and meaningful data (i.e. not zeroes) must be entered in all columns. A number of pre-built table files can be found in the `GMXLIB` directory for 6-8, 6-9, 6-10, 6-11, and 6-12 Lennard-Jones potentials combined with a normal Coulomb.

If you want to have different functional forms between different groups of atoms, this can be set through energy groups. Different tables can be used for non-bonded interactions between different energy groups pairs through the `.mdp` option `energygrp-table` (see details in the User Guide). Atoms that should interact with a different potential should be put into different energy groups. Between group pairs which are not listed in `energygrp-table`, the normal user tables will be used. This makes it easy to use a different functional form between a few types of atoms.

6.13 Mixed Quantum-Classical simulation techniques

In a molecular mechanics (MM) force field, the influence of electrons is expressed by empirical parameters that are assigned on the basis of experimental data, or on the basis of results from high-level quantum chemistry calculations. These are valid for the ground state of a given covalent structure, and the MM approximation is usually sufficiently accurate for ground-state processes in which the overall connectivity between the atoms in the system remains unchanged. However, for processes in which the connectivity does change, such as chemical reactions, or processes that involve multiple electronic states, such as photochemical conversions, electrons can no longer be ignored, and a quantum mechanical description is required for at least those parts of the system in which the reaction takes place.

One approach to the simulation of chemical reactions in solution, or in enzymes, is to use a combination of quantum mechanics (QM) and molecular mechanics (MM). The reacting parts of the system are treated quantum mechanically, with the remainder being modeled using the force field. The current version of GROMACS provides interfaces to several popular Quantum Chemistry packages (MOPAC [155], GAMESS-UK [156], Gaussian [157] and CPMD [158]).

GROMACS interactions between the two subsystems are either handled as described by Field *et al.* [159] or within the ONIOM approach by Morokuma and coworkers [160, 161].

6.13.1 Overview

Two approaches for describing the interactions between the QM and MM subsystems are supported in this version:

1. **Electronic Embedding** The electrostatic interactions between the electrons of the QM region and the MM atoms and between the QM nuclei and the MM atoms are included in the Hamiltonian for the QM subsystem:

$$H^{QM/MM} = H_e^{QM} - \sum_i^n \sum_J^M \frac{e^2 Q_J}{4\pi\epsilon_0 r_{iJ}} + \sum_A^N \sum_J^M \frac{e^2 Z_A Q_J}{e\pi\epsilon_0 R_{AJ}}, \quad (6.103)$$

where n and N are the number of electrons and nuclei in the QM region, respectively, and M is the number of charged MM atoms. The first term on the right hand side is the original electronic Hamiltonian of an isolated QM system. The first of the double sums is the total electrostatic interaction between the QM electrons and the MM atoms. The total electrostatic interaction of the QM nuclei with the MM atoms is given by the second double sum. Bonded interactions between QM and MM atoms are described at the MM level by the appropriate force-field terms. Chemical bonds that connect the two subsystems are capped by a hydrogen atom to complete the valence of the QM region. The force on this atom, which is present in the QM region only, is distributed over the two atoms of the bond. The cap atom is usually referred to as a link atom.

2. **ONIOM** In the ONIOM approach, the energy and gradients are first evaluated for the isolated QM subsystem at the desired level of *ab initio* theory. Subsequently, the energy and gradients of the total system, including the QM region, are computed using the molecular mechanics force field and added to the energy and gradients calculated for the isolated QM subsystem. Finally, in order to correct for counting the interactions inside the QM region twice, a molecular mechanics calculation is performed on the isolated QM subsystem and the energy and gradients are subtracted. This leads to the following expression for the total QM/MM energy (and gradients likewise):

$$E_{tot} = E_I^{QM} + E_{I+II}^{MM} - E_I^{MM}, \quad (6.104)$$

where the subscripts I and II refer to the QM and MM subsystems, respectively. The superscripts indicate at what level of theory the energies are computed. The ONIOM scheme has the advantage that it is not restricted to a two-layer QM/MM description, but can easily handle more than two layers, with each layer described at a different level of theory.

6.13.2 Usage

To make use of the QM/MM functionality in GROMACS, one needs to:

1. introduce link atoms at the QM/MM boundary, if needed;
2. specify which atoms are to be treated at a QM level;
3. specify the QM level, basis set, type of QM/MM interface and so on.

Adding link atoms

At the bond that connects the QM and MM subsystems, a link atom is introduced. In GROMACS the link atom has special atomtype, called LA. This atomtype is treated as a hydrogen atom in the QM calculation, and as a virtual site in the force-field calculation. The link atoms, if any, are part of the system, but have no interaction with any other atom, except that the QM force working on it is distributed over the two atoms of the bond. In the topology, the link atom (LA), therefore, is defined as a virtual site atom:

```
[ virtual_sites2 ]  
LA QMatom MMatom 1 0.65
```

See sec. 5.2.2 for more details on how virtual sites are treated. The link atom is replaced at every step of the simulation.

In addition, the bond itself is replaced by a constraint:

```
[ constraints ]  
QMatom MMatom 2 0.153
```

Note that, because in our system the QM/MM bond is a carbon-carbon bond (0.153 nm), we use a constraint length of 0.153 nm, and dummy position of 0.65. The latter is the ratio between the ideal C-H bond length and the ideal C-C bond length. With this ratio, the link atom is always 0.1 nm away from the QMatom, consistent with the carbon-hydrogen bond length. If the QM and MM subsystems are connected by a different kind of bond, a different constraint and a different dummy position, appropriate for that bond type, are required.

Specifying the QM atoms

Atoms that should be treated at a QM level of theory, including the link atoms, are added to the index file. In addition, the chemical bonds between the atoms in the QM region are to be defined as connect bonds (bond type 5) in the topology file:

```
[ bonds ]  
QMatom1 QMatom2 5  
QMatom2 QMatom3 5
```

Specifying the QM/MM simulation parameters

In the .mdp file, the following parameters control a QM/MM simulation.

```
QMMM = no
```

If this is set to *yes*, a QM/MM simulation is requested. Several groups of atoms can be described at different QM levels separately. These are specified in the QMMM-grps field separated by spaces. The level of *ab initio* theory at which the groups are described is specified by QMmethod and QMbasis Fields. Describing the groups at different levels of theory is only possible with the ONIOM QM/MM scheme, specified by QMMMscheme.

```
QMMM-grps =  
groups to be described at the QM level
```

```
QMMMscheme = normal
```

Options are *normal* and *ONIOM*. This selects the QM/MM interface. *normal* implies that the QM subsystem is electronically embedded in the MM subsystem. There can only be one QMMM-grps that is modeled at the QMmethod and QMbasis level of *ab initio*

theory. The rest of the system is described at the MM level. The QM and MM subsystems interact as follows: MM point charges are included in the QM one-electron Hamiltonian and all Lennard-Jones interactions are described at the MM level. If ONIOM is selected, the interaction between the subsystem is described using the ONIOM method by Morokuma and co-workers. There can be more than one QMMM-grps each modeled at a different level of QM theory (QMmethod and QMbasis).

QMmethod =

Method used to compute the energy and gradients on the QM atoms. Available methods are AM1, PM3, RHF, UHF, DFT, B3LYP, MP2, CASSCF, MMVB and CPMD. For CASSCF, the number of electrons and orbitals included in the active space is specified by CASelectrons and CASorbitals. For CPMD, the plane-wave cut-off is specified by the planewavecutoff keyword.

QMbasis =

Gaussian basis set used to expand the electronic wave-function. Only Gaussian basis sets are currently available, i.e. STO-3G, 3-21G, 3-21G*, 3-21+G*, 6-21G, 6-31G, 6-31G*, 6-31+G*, and 6-311G. For CPMD, which uses plane wave expansion rather than atom-centered basis functions, the planewavecutoff keyword controls the plane wave expansion.

QMcharge =

The total charge in e of the QMMM-grps. In case there are more than one QMMM-grps, the total charge of each ONIOM layer needs to be specified separately.

QMmult =

The multiplicity of the QMMM-grps. In case there are more than one QMMM-grps, the multiplicity of each ONIOM layer needs to be specified separately.

CASorbitals =

The number of orbitals to be included in the active space when doing a CASSCF computation.

CASelectrons =

The number of electrons to be included in the active space when doing a CASSCF computation.

SH = no

If this is set to yes, a QM/MM MD simulation on the excited state-potential energy surface and enforce a diabatic hop to the ground-state when the system hits the conical intersection hyperline in the course the simulation. This option only works in combination with the CASSCF method.

6.13.3 Output

The energies and gradients computed in the QM calculation are added to those computed by GRO-MACS. In the `.edr` file there is a section for the total QM energy.

6.13.4 Future developments

Several features are currently under development to increase the accuracy of the QM/MM interface. One useful feature is the use of delocalized MM charges in the QM computations. The most important benefit of using such smeared-out charges is that the Coulombic potential has a finite value at interatomic distances. In the point charge representation, the partially-charged MM atoms close to the QM region tend to “over-polarize” the QM system, which leads to artifacts in the calculation.

What is needed as well is a transition state optimizer.

6.14 Using VMD plug-ins for trajectory file I/O

GROMACS tools are able to use the plug-ins found in an existing installation of [VMD](#) in order to read and write trajectory files in formats that are not native to GROMACS. You will be able to supply an AMBER DCD-format trajectory filename directly to GROMACS tools, for example.

This requires a VMD installation not older than version 1.8, that your system provides the `dlopen` function so that programs can determine at run time what plug-ins exist, and that you build shared libraries when building GROMACS. CMake will find the `vmd` executable in your path, and from it, or the environment variable `VMDDIR` at configuration or run time, locate the plug-ins. Alternatively, the `VMD_PLUGIN_PATH` can be used at run time to specify a path where these plug-ins can be found. Note that these plug-ins are in a binary format, and that format must match the architecture of the machine attempting to use them.

6.15 Interactive Molecular Dynamics

GROMACS supports the interactive molecular dynamics (IMD) protocol as implemented by [VMD](#) to control a running simulation in NAMD. IMD allows to monitor a running GROMACS simulation from a VMD client. In addition, the user can interact with the simulation by pulling on atoms, residues or fragments with a mouse or a force-feedback device. Additional information about the GROMACS implementation and an exemplary GROMACS IMD system can be found [on this homepage](#).

6.15.1 Simulation input preparation

The GROMACS implementation allows transmission and interaction with a part of the running simulation only, e.g. in cases where no water molecules should be transmitted or pulled. The group is specified via the `.mdp` option `IMD-group`. When `IMD-group` is empty, the IMD protocol is disabled and cannot be enabled via the switches in `mdrun`. To interact with the entire system, `IMD-group` can be set to `System`. When using `grompp`, a `.gro` file to be used as VMD input is written out (`-imd` switch of `grompp`).

6.15.2 Starting the simulation

Communication between VMD and GROMACS is achieved via TCP sockets and thus enables controlling an `mdrun` running locally or on a remote cluster. The port for the connection can be specified with the `-imdport` switch of `mdrun`, 8888 is the default. If a port number of 0 or smaller is provided, GROMACS automatically assigns a free port to use with IMD.

Every N steps, the `mdrun` client receives the applied forces from VMD and sends the new positions to the client. VMD permits increasing or decreasing the communication frequency interactively. By default, the simulation starts and runs even if no IMD client is connected. This behavior is changed by the `-imdwait` switch of `mdrun`. After startup and whenever the client has disconnected, the integration stops until reconnection of the client. When the `-imdterm` switch is used, the simulation can be terminated by pressing the stop button in VMD. This is disabled by default. Finally, to allow interacting with the simulation (i.e. pulling from VMD) the `-imdpull` switch has to be used. Therefore, a simulation can only be monitored but not influenced from the VMD client when none of `-imdwait`, `-imdterm` or `-imdpull` are set. However, since the IMD protocol requires no authentication, it is not advisable to run simulations on a host directly reachable from an insecure environment. Secure shell forwarding of TCP can be used to connect to running simulations not directly reachable from the interacting host. Note that the IMD command line switches of `mdrun` are hidden by default and show up in the help text only with `gmx mdrun -h -hidden`.

6.15.3 Connecting from VMD

In VMD, first the structure corresponding to the IMD group has to be loaded (*File* → *New Molecule*). Then the IMD connection window has to be used (*Extensions* → *Simulation* → *IMD Connect (NAMD)*). In the IMD connection window, hostname and port have to be specified and followed by pressing *Connect*. *Detach Sim* allows disconnecting without terminating the simulation, while *Stop Sim* ends the simulation on the next neighbor searching step (if allowed by `-imdterm`).

The timestep transfer rate allows adjusting the communication frequency between simulation and IMD client. Setting the keep rate loads every N^{th} frame into VMD instead of discarding them when a new one is received. The displayed energies are in SI units in contrast to energies displayed from NAMD simulations.

6.16 Embedding proteins into the membranes

GROMACS is capable of inserting the protein into pre-equilibrated lipid bilayers with minimal perturbation of the lipids using the method, which was initially described as a ProtSqueeze technique,[162] and later implemented as `g_membed` tool.[163] Currently the functionality of `g_membed` is available in `mdrun` as described in the user guide.

This method works by first artificially shrinking the protein in the xy -plane, then it removes lipids that overlap with that much smaller core. Then the protein atoms are gradually resized back to their initial configuration, using normal dynamics for the rest of the system, so the lipids adapt to

the protein. Further lipids are removed as required.

Chapter 7

Run parameters and Programs

7.1 Online documentation

More documentation is available online from the GROMACS web site, <http://manual.gromacs.org/documentation>.

In addition, we install standard UNIX man pages for all the programs. If you have sourced the GMXRC script in the GROMACS binary directory for your host they should already be present in your MANPATH environment variable, and you should be able to type *e.g.* `man gmx-grompp`. You can also use the `-h` flag on the command line (*e.g.* `gmx grompp -h`) to see the same information, as well as `gmx help grompp`. The list of all programs are available from `gmx help`.

7.2 File types

Table 7.1 lists the file types used by GROMACS along with a short description, and you can find a more detail description for each file in your HTML reference, or in our online version.

GROMACS files written in XDR format can be read on any architecture with GROMACS version 1.6 or later if the configuration script found the XDR libraries on your system. They should always be present on UNIX since they are necessary for NFS support.

7.3 Run Parameters

The descriptions of `.mdp` parameters can be found at <http://manual.gromacs.org/current/mdp-options.html> or in your installation at `share/gromacs/html/mdp-options.html`

Default Name	Ext.	Type	Default Option	Description
atomtp.atp		Asc		Atomtype file used by pdb2gmx
eiwit.brk		Asc	-f	Brookhaven data bank file
state.cpt		xdr		Checkpoint file
nndice.dat		Asc		Generic data file
user.dlg		Asc		Dialog Box data for ngmx
sam.edi		Asc		ED sampling input
sam.edo		Asc		ED sampling output
ener.edr				Generic energy: edr ene
ener.edr		xdr		Energy file in portable xdr format
ener.ene		Bin		Energy file
eiwit.ent		Asc	-f	Entry in the protein data bank
plot.eps		Asc		Encapsulated PostScript (tm) file
conf.esp		Asc	-c	Coordinate file in ESPResSo format
conf.g96		Asc	-c	Coordinate file in Gromos-96 format
conf.gro		Asc	-c	Coordinate file in Gromos-87 format
conf.gro			-c	Structure: gro g96 pdb esp tpr
out.gro			-o	Structure: gro g96 pdb esp
polar.hdb		Asc		Hydrogen data base
topinc.itp		Asc		Include file for topology
run.log		Asc	-l	Log file
ps.m2p		Asc		Input file for mat2ps
ss.map		Asc		File that maps matrix data to colors
ss.mat		Asc		Matrix Data file
grompp.mdp		Asc	-f	grompp input file with MD parameters
hessian.mtx		Bin	-m	Hessian matrix
index.ndx		Asc	-n	Index file
hello.out		Asc	-o	Generic output file
eiwit.pdb		Asc	-f	Protein data bank file
residue.rtp		Asc		Residue Type file used by pdb2gmx
doc.tex		Asc	-o	LaTeX file
topol.top		Asc	-p	Topology file
topol.tpr			-s	Generic run input: tpr
topol.tpr			-s	Structure+mass(db): tpr gro g96 pdb
topol.tpr		xdr	-s	Portable xdr run input file
traj.trr				Full precision trajectory: trr cpt
traj.trr		xdr		Trajectory in portable xdr format
root.xpm		Asc		X PixMap compatible matrix file
traj.xtc			-f	Trajec., input: xtc trr tng cpt gro g96 pdb
traj.xtc			-f	Trajectory, output: xtc trr tng gro g96 pdb
traj.xtc		xdr		Compressed trajectory (portable xdr format)
graph.xvg		Asc	-o	xvgr/xmgr file

Table 7.1: The GROMACS file types.

Chapter 8

Analysis

In this chapter different ways of analyzing your trajectory are described. The names of the corresponding analysis programs are given. Specific information on the in- and output of these programs can be found in the online manual at www.gromacs.org. The output files are often produced as finished Grace/Xmgr graphs.

First, in sec. 8.1, the group concept in analysis is explained. 8.1.2 explains a newer concept of dynamic selections, which is currently supported by a few tools. Then, the different analysis tools are presented.

8.1 Using Groups

`gmx make_ndx`, `gmx mk_angndx`, `gmx select`

In chapter 3, it was explained how *groups of atoms* can be used in `mdrun` (see sec. 3.3). In most analysis programs, groups of atoms must also be chosen. Most programs can generate several default index groups, but groups can always be read from an index file. Let's consider the example of a simulation of a binary mixture of components A and B. When we want to calculate the radial distribution function (RDF) $g_{AB}(r)$ of A with respect to B, we have to calculate:

$$4\pi r^2 g_{AB}(r) = V \sum_{i \in A} \sum_{j \in B} P(r) \quad (8.1)$$

where V is the volume and $P(r)$ is the probability of finding a B atom at distance r from an A atom.

By having the user define the *atom numbers* for groups A and B in a simple file, we can calculate this g_{AB} in the most general way, without having to make any assumptions in the RDF program about the type of particles.

Groups can therefore consist of a series of *atom numbers*, but in some cases also of *molecule numbers*. It is also possible to specify a series of angles by *triples* of *atom numbers*, dihedrals by *quadruples* of *atom numbers* and bonds or vectors (in a molecule) by *pairs* of *atom numbers*.

When appropriate the type of index file will be specified for the following analysis programs. To help creating such index files (`index.ndx`), there are a couple of programs to generate them, using either your input configuration or the topology. To generate an index file consisting of a series of *atom numbers* (as in the example of g_{AB}), use `gmx make_ndx` or `gmx select`. To generate an index file with angles or dihedrals, use `gmx mk_angndx`. Of course you can also make them by hand. The general format is presented here:

```
[ Oxygen ]
  1      4      7

[ Hydrogen ]
  2      3      5      6
  8      9
```

First, the group name is written between square brackets. The following atom numbers may be spread out over as many lines as you like. The atom numbering starts at 1.

Each tool that can use groups will offer the available alternatives for the user to choose. That choice can be made with the number of the group, or its name. In fact, the first few letters of the group name will suffice if that will distinguish the group from all others. There are ways to use Unix shell features to choose group names on the command line, rather than interactively. Consult www.gromacs.org for suggestions.

8.1.1 Default Groups

When no index file is supplied to analysis tools or `grompp`, a number of default groups are generated to choose from:

```
System
  all atoms in the system

Protein
  all protein atoms

Protein-H
  protein atoms excluding hydrogens

C-alpha
  C $_{\alpha}$  atoms

Backbone
  protein backbone atoms; N, C $_{\alpha}$  and C

MainChain
  protein main chain atoms: N, C $_{\alpha}$ , C and O, including oxygens in C-terminus

MainChain+Cb
  protein main chain atoms including C $_{\beta}$ 
```

MainChain+H	protein main chain atoms including backbone amide hydrogens and hydrogens on the N-terminus
SideChain	protein side chain atoms; that is all atoms except N, C _α , C, O, backbone amide hydrogens, oxygens in C-terminus and hydrogens on the N-terminus
SideChain-H	protein side chain atoms excluding all hydrogens
Prot-Masses	protein atoms excluding dummy masses (as used in virtual site constructions of NH ₃ groups and tryptophan side-chains), see also sec. 5.2.2; this group is only included when it differs from the “Protein” group
Non-Protein	all non-protein atoms
DNA	all DNA atoms
RNA	all RNA atoms
Water	water molecules (names like SOL, WAT, HOH, etc.) See <code>residuetypes.dat</code> for a full listing
non-Water	anything not covered by the Water group
Ion	any name matching an Ion entry in <code>residuetypes.dat</code>
Water_and_Ions	combination of the Water and Ions groups
molecule_name	for all residues/molecules which are not recognized as protein, DNA, or RNA; one group per residue/molecule name is generated
Other	all atoms which are neither protein, DNA, nor RNA.

Empty groups will not be generated. Most of the groups only contain protein atoms. An atom is considered a protein atom if its residue name is listed in the `residuetypes.dat` file and is listed as a “Protein” entry. The process for determining DNA, RNA, etc. is analogous. If you need to modify these classifications, then you can copy the file from the library directory into your working directory and edit the local copy.

8.1.2 Selections

`gmx select`

Currently, a few analysis tools support an extended concept of (*dynamic*) *selections*. There are three main differences to traditional index groups:

- The selections are specified as text instead of reading fixed atom indices from a file, using a syntax similar to VMD. The text can be entered interactively, provided on the command line, or from a file.
- The selections are not restricted to atoms, but can also specify that the analysis is to be performed on, e.g., center-of-mass positions of a group of atoms. Some tools may not support selections that do not evaluate to single atoms, e.g., if they require information that is available only for single atoms, like atom names or types.
- The selections can be dynamic, i.e., evaluate to different atoms for different trajectory frames. This allows analyzing only a subset of the system that satisfies some geometric criteria.

As an example of a simple selection, `resname ABC and within 2 of resname DEF` selects all atoms in residues named ABC that are within 2 nm of any atom in a residue named DEF.

Tools that accept selections can also use traditional index files similarly to older tools: it is possible to give an `.ndx` file to the tool, and directly select a group from the index file as a selection, either by group number or by group name. The index groups can also be used as a part of a more complicated selection.

To get started, you can run `gmx select` with a single structure, and use the interactive prompt to try out different selections. The tool provides, among others, output options `-on` and `-ofpdb` to write out the selected atoms to an index file and to a `.pdb` file, respectively. This does not allow testing selections that evaluate to center-of-mass positions, but other selections can be tested and the result examined.

The detailed syntax and the individual keywords that can be used in selections can be accessed by typing `help` in the interactive prompt of any selection-enabled tool, as well as with `gmx help selections`. The help is divided into subtopics that can be accessed with, e.g., `help syntax` / `gmx help selections syntax`. Some individual selection keywords have extended help as well, which can be accessed with, e.g., `help keywords within`.

The interactive prompt does not currently provide much editing capabilities. If you need them, you can run the program under `rlwrap`.

For tools that do not yet support the selection syntax, you can use `gmx select -on` to generate static index groups to pass to the tool. However, this only allows for a small subset (only the first bullet from the above list) of the flexibility that fully selection-aware tools offer.

It is also possible to write your own analysis tools to take advantage of the flexibility of these selections: see the `template.cpp` file in the `share/gromacs/template` directory of your installation for an example.

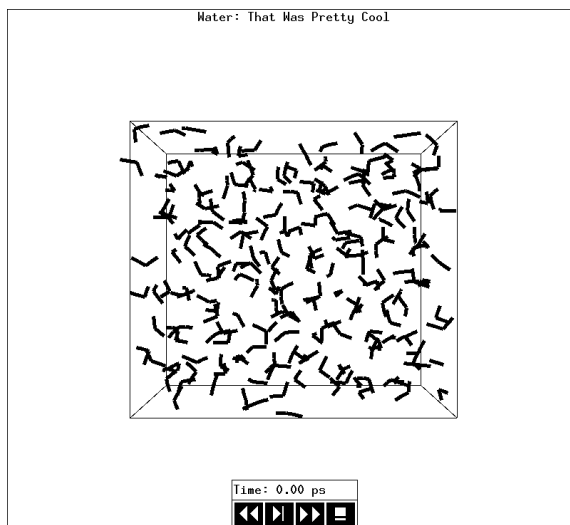


Figure 8.1: The window of `gmx view` showing a box of water.

8.2 Looking at your trajectory

`gmx view`

Before analyzing your trajectory it is often informative to look at your trajectory first. GROMACS comes with a simple trajectory viewer `gmx view`; the advantage with this one is that it does not require OpenGL, which usually isn't present on *e.g.* supercomputers. It is also possible to generate a hard-copy in Encapsulated Postscript format (see Fig. 8.1). If you want a faster and more fancy viewer there are several programs that can read the GROMACS trajectory formats – have a look at our homepage (www.gromacs.org) for updated links.

8.3 General properties

`gmx energy`, `gmx traj`

To analyze some or all *energies* and other properties, such as *total pressure*, *pressure tensor*, *density*, *box-volume* and *box-sizes*, use the program `gmx energy`. A choice can be made from a list a set of energies, like potential, kinetic or total energy, or individual contributions, like Lennard-Jones or dihedral energies.

The *center-of-mass velocity*, defined as

$$\mathbf{v}_{com} = \frac{1}{M} \sum_{i=1}^N m_i \mathbf{v}_i \quad (8.2)$$

with $M = \sum_{i=1}^N m_i$ the total mass of the system, can be monitored in time by the program `gmx traj -com -ov`. It is however recommended to remove the center-of-mass velocity every step (see chapter 3)!

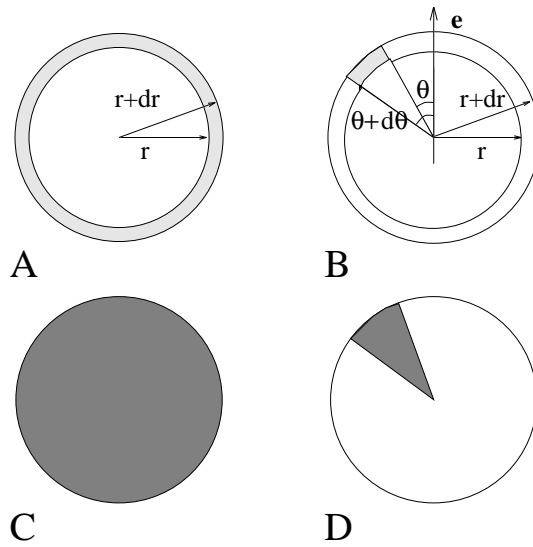


Figure 8.2: Definition of slices in `gmx rdf`: A. $g_{AB}(r)$. B. $g_{AB}(r, \theta)$. The slices are colored gray. C. Normalization $\langle \rho_B \rangle_{local}$. D. Normalization $\langle \rho_B \rangle_{local, \theta}$. Normalization volumes are colored gray.

8.4 Radial distribution functions

`gmx rdf`

The *radial distribution function* (RDF) or pair correlation function $g_{AB}(r)$ between particles of type *A* and *B* is defined in the following way:

$$\begin{aligned}
 g_{AB}(r) &= \frac{\langle \rho_B(r) \rangle}{\langle \rho_B \rangle_{local}} \\
 &= \frac{1}{\langle \rho_B \rangle_{local}} \frac{1}{N_A} \sum_{i \in A} \sum_{j \in B} \frac{\delta(r_{ij} - r)}{4\pi r^2}
 \end{aligned} \tag{8.3}$$

with $\langle \rho_B(r) \rangle$ the particle density of type *B* at a distance *r* around particles *A*, and $\langle \rho_B \rangle_{local}$ the particle density of type *B* averaged over all spheres around particles *A* with radius r_{max} (see Fig. 8.2C).

Usually the value of r_{max} is half of the box length. The averaging is also performed in time. In practice the analysis program `gmx rdf` divides the system into spherical slices (from *r* to $r + dr$, see Fig. 8.2A) and makes a histogram in stead of the δ -function. An example of the RDF of oxygen-oxygen in SPC water [85] is given in Fig. 8.3.

With `gmx rdf` it is also possible to calculate an angle dependent rdf $g_{AB}(r, \theta)$, where the angle θ is defined with respect to a certain laboratory axis **e**, see Fig. 8.2B.

$$g_{AB}(r, \theta) = \frac{1}{\langle \rho_B \rangle_{local, \theta}} \frac{1}{N_A} \sum_{i \in A} \sum_{j \in B} \frac{\delta(r_{ij} - r) \delta(\theta_{ij} - \theta)}{2\pi r^2 \sin(\theta)} \tag{8.4}$$

$$\cos(\theta_{ij}) = \frac{\mathbf{r}_{ij} \cdot \mathbf{e}}{\|\mathbf{r}_{ij}\| \|\mathbf{e}\|} \tag{8.5}$$

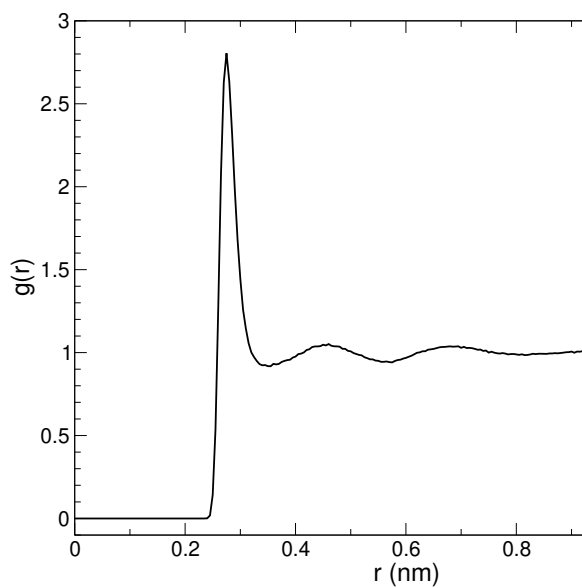


Figure 8.3: $g_{OO}(r)$ for Oxygen-Oxygen of SPC-water.

This $g_{AB}(r, \theta)$ is useful for analyzing anisotropic systems. **Note** that in this case the normalization $\langle \rho_B \rangle_{local, \theta}$ is the average density in all angle slices from θ to $\theta + d\theta$ up to r_{max} , so angle dependent, see Fig. 8.2D.

8.5 Correlation functions

8.5.1 Theory of correlation functions

The theory of correlation functions is well established [113]. We describe here the implementation of the various correlation function flavors in the GROMACS code. The definition of the autocorrelation function (ACF) $C_f(t)$ for a property $f(t)$ is:

$$C_f(t) = \langle f(\xi)f(\xi + t) \rangle_{\xi} \quad (8.6)$$

where the notation on the right hand side indicates averaging over ξ , *i.e.* over time origins. It is also possible to compute cross-correlation function from two properties $f(t)$ and $g(t)$:

$$C_{fg}(t) = \langle f(\xi)g(\xi + t) \rangle_{\xi} \quad (8.7)$$

however, in GROMACS there is no standard mechanism to do this (**note:** you can use the `xmgr` program to compute cross correlations). The integral of the correlation function over time is the correlation time τ_f :

$$\tau_f = \int_0^{\infty} C_f(t) dt \quad (8.8)$$

In practice, correlation functions are calculated based on data points with discrete time intervals Δt , so that the ACF from an MD simulation is:

$$C_f(j\Delta t) = \frac{1}{N-j} \sum_{i=0}^{N-1-j} f(i\Delta t)f((i+j)\Delta t) \quad (8.9)$$

where N is the number of available time frames for the calculation. The resulting ACF is obviously only available at time points with the same interval Δt . Since, for many applications, it is necessary to know the short time behavior of the ACF (*e.g.* the first 10 ps) this often means that we have to save the data with intervals much shorter than the time scale of interest. Another implication of eqn. 8.9 is that in principle we can not compute all points of the ACF with the same accuracy, since we have $N - 1$ data points for $C_f(\Delta t)$ but only 1 for $C_f((N - 1)\Delta t)$. However, if we decide to compute only an ACF of length $M\Delta t$, where $M \leq N/2$ we can compute all points with the same statistical accuracy:

$$C_f(j\Delta t) = \frac{1}{M} \sum_{i=0}^{N-1-M} f(i\Delta t)f((i+j)\Delta t) \quad (8.10)$$

Here of course $j < M$. M is sometimes referred to as the time lag of the correlation function. When we decide to do this, we intentionally do not use all the available points for very short time intervals ($j \ll M$), but it makes it easier to interpret the results. Another aspect that may not be neglected when computing ACFs from simulation is that usually the time origins ξ (eqn. 8.6) are not statistically independent, which may introduce a bias in the results. This can be tested using a

block-averaging procedure, where only time origins with a spacing at least the length of the time lag are included, *e.g.* using k time origins with spacing of $M\Delta t$ (where $kM \leq N$):

$$C_f(j\Delta t) = \frac{1}{k} \sum_{i=0}^{k-1} f(iM\Delta t)f((iM+j)\Delta t) \quad (8.11)$$

However, one needs very long simulations to get good accuracy this way, because there are many fewer points that contribute to the ACF.

8.5.2 Using FFT for computation of the ACF

The computational cost for calculating an ACF according to eqn. 8.9 is proportional to N^2 , which is considerable. However, this can be improved by using fast Fourier transforms to do the convolution [113].

8.5.3 Special forms of the ACF

There are some important varieties on the ACF, *e.g.* the ACF of a vector \mathbf{p} :

$$C_{\mathbf{p}}(t) = \int_0^\infty P_n(\cos \angle(\mathbf{p}(\xi), \mathbf{p}(\xi+t))) d\xi \quad (8.12)$$

where $P_n(x)$ is the n^{th} order Legendre polynomial¹. Such correlation times can actually be obtained experimentally using *e.g.* NMR or other relaxation experiments. GROMACS can compute correlations using the 1st and 2nd order Legendre polynomial (eqn. 8.12). This can also be used for rotational autocorrelation (`gmx rotacf`) and dipole autocorrelation (`gmx dipoles`).

In order to study torsion angle dynamics, we define a dihedral autocorrelation function as [164]:

$$C(t) = \langle \cos(\theta(\tau) - \theta(\tau+t)) \rangle_\tau \quad (8.13)$$

Note that this is not a product of two functions as is generally used for correlation functions, but it may be rewritten as the sum of two products:

$$C(t) = \langle \cos(\theta(\tau)) \cos(\theta(\tau+t)) + \sin(\theta(\tau)) \sin(\theta(\tau+t)) \rangle_\tau \quad (8.14)$$

8.5.4 Some Applications

The program `gmx velacc` calculates the *velocity autocorrelation function*.

$$C_{\mathbf{v}}(\tau) = \langle \mathbf{v}_i(\tau) \cdot \mathbf{v}_i(0) \rangle_{i \in A} \quad (8.15)$$

The self diffusion coefficient can be calculated using the Green-Kubo relation [113]:

$$D_A = \frac{1}{3} \int_0^\infty \langle \mathbf{v}_i(t) \cdot \mathbf{v}_i(0) \rangle_{i \in A} dt \quad (8.16)$$

¹ $P_0(x) = 1, P_1(x) = x, P_2(x) = (3x^2 - 1)/2$

which is just the integral of the velocity autocorrelation function. There is a widely-held belief that the velocity ACF converges faster than the mean square displacement (sec. 8.7), which can also be used for the computation of diffusion constants. However, Allen & Tildesley [113] warn us that the long-time contribution to the velocity ACF can not be ignored, so care must be taken.

Another important quantity is the dipole correlation time. The *dipole correlation function* for particles of type A is calculated as follows by `gmx dipoles`:

$$C_{\mu}(\tau) = \langle \mu_i(\tau) \cdot \mu_i(0) \rangle_{i \in A} \quad (8.17)$$

with $\mu_i = \sum_{j \in i} \mathbf{r}_j q_j$. The dipole correlation time can be computed using eqn. 8.8. For some applications see [165].

The viscosity of a liquid can be related to the correlation time of the Pressure tensor \mathbf{P} [166, 167]. `gmx energy` can compute the viscosity, but this is not very accurate [154], and actually the values do not converge.

8.6 Curve fitting in GROMACS

8.6.1 Sum of exponential functions

Sometimes it is useful to fit a curve to an analytical function, for example in the case of autocorrelation functions with noisy tails. GROMACS is not a general purpose curve-fitting tool however and therefore GROMACS only supports a limited number of functions. Table 8.1 lists the available options with the corresponding command-line options. The underlying routines for fitting use the Levenberg-Marquardt algorithm as implemented in the `lmfit` package [168] (a bare-bones version of which is included in GROMACS in which an option for error-weighted fitting was implemented).

Table 8.1: Overview of fitting functions supported in (most) analysis tools that compute autocorrelation functions. The “Note” column describes properties of the output parameters.

Command line option	Functional form $f(t)$	Note
<code>exp</code>	e^{-t/a_0}	
<code>aexp</code>	$a_1 e^{-t/a_0}$	
<code>exp_exp</code>	$a_1 e^{-t/a_0} + (1 - a_1) e^{-t/a_2}$	$a_2 \geq a_0 \geq 0$
<code>exp5</code>	$a_1 e^{-t/a_0} + a_3 e^{-t/a_2} + a_4$	$a_2 \geq a_0 \geq 0$
<code>exp7</code>	$a_1 e^{-t/a_0} + a_3 e^{-t/a_2} + a_5 e^{-t/a_4} + a_6$	$a_4 \geq a_2 \geq a_0 \geq 0$
<code>exp9</code>	$a_1 e^{-t/a_0} + a_3 e^{-t/a_2} + a_5 e^{-t/a_4} + a_7 e^{-t/a_6} + a_8$	$a_6 \geq a_4 \geq a_2 \geq a_0 \geq 0$

8.6.2 Error estimation

Under the hood GROMACS implements some more fitting functions, namely a function to estimate the error in time-correlated data due to Hess [154]:

$$\varepsilon^2(t) = \alpha \tau_1 \left(1 + \frac{\tau_1}{t} (e^{-t/\tau_1} - 1) \right) + (1 - \alpha) \tau_2 \left(1 + \frac{\tau_2}{t} (e^{-t/\tau_2} - 1) \right) \quad (8.18)$$

where τ_1 and τ_2 are time constants (with $\tau_2 \geq \tau_1$) and α usually is close to 1 (in the fitting procedure it is enforced that $0 \leq \alpha \leq 1$). This is used in `gmx analyze` for error estimation using

$$\lim_{t \rightarrow \infty} \varepsilon(t) = \sigma \sqrt{\frac{2(\alpha\tau_1 + (1 - \alpha)\tau_2)}{T}} \quad (8.19)$$

where σ is the standard deviation of the data set and T is the total simulation time [154].

8.6.3 Interphase boundary demarcation

In order to determine the position and width of an interface, Steen-Sæthre *et al.* fitted a density profile to the following function

$$f(x) = \frac{a_0 + a_1}{2} - \frac{a_0 - a_1}{2} \operatorname{erf}\left(\frac{x - a_2}{a_3}\right) \quad (8.20)$$

where a_0 and a_1 are densities of different phases, x is the coordinate normal to the interface, a_2 is the position of the interface and a_3 is the width of the interface [169]. This is implemented in `gmx densorder`.

8.6.4 Transverse current autocorrelation function

In order to establish the transverse current autocorrelation function (useful for computing viscosity [170]) the following function is fitted:

$$f(x) = e^{-\nu} \left(\cosh(\omega\nu) + \frac{\sinh(\omega\nu)}{\omega} \right) \quad (8.21)$$

with $\nu = x/(2a_0)$ and $\omega = \sqrt{1 - a_1}$. This is implemented in `gmx tcacf`.

8.6.5 Viscosity estimation from pressure autocorrelation function

The viscosity is a notoriously difficult property to extract from simulations [154, 171]. It is *in principle* possible to determine it by integrating the pressure autocorrelation function [166], however this is often hampered by the noisy tail of the ACF. A workaround to this is fitting the ACF to the following function [172]:

$$f(t)/f(0) = (1 - C)\cos(\omega t)e^{-(t/\tau_f)^{\beta_f}} + Ce^{-(t/\tau_s)^{\beta_s}} \quad (8.22)$$

where ω is the frequency of rapid pressure oscillations (mainly due to bonded forces in molecular simulations), τ_f and β_f are the time constant and exponent of fast relaxation in a stretched-exponential approximation, τ_s and β_s are constants for slow relaxation and C is the pre-factor that determines the weight between fast and slow relaxation. After a fit, the integral of the function $f(t)$ is used to compute the viscosity:

$$\eta = \frac{V}{k_B T} \int_0^\infty f(t) dt \quad (8.23)$$

This equation has been applied to computing the bulk and shear viscosity using different elements from the pressure tensor [173]. This is implemented in `gmx viscosity`.

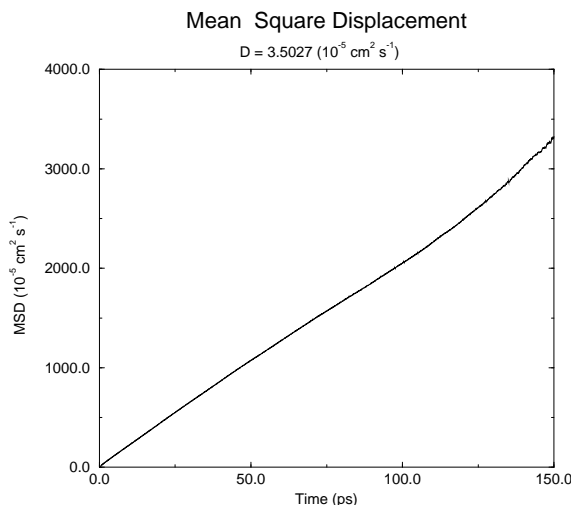


Figure 8.4: Mean Square Displacement of SPC-water.

8.7 Mean Square Displacement

`gmx msd`

To determine the self diffusion coefficient D_A of particles of type A , one can use the Einstein relation [113]:

$$\lim_{t \rightarrow \infty} \langle \|\mathbf{r}_i(t) - \mathbf{r}_i(0)\|^2 \rangle_{i \in A} = 6D_A t \quad (8.24)$$

This *mean square displacement* and D_A are calculated by the program `gmx msd`. Normally an index file containing atom numbers is used and the MSD is averaged over these atoms. For molecules consisting of more than one atom, \mathbf{r}_i can be taken as the center of mass positions of the molecules. In that case, you should use an index file with molecule numbers. The results will be nearly identical to averaging over atoms, however. The `gmx msd` program can also be used for calculating diffusion in one or two dimensions. This is useful for studying lateral diffusion on interfaces.

An example of the mean square displacement of SPC water is given in Fig. 8.4.

8.8 Bonds/distances, angles and dihedrals

`gmx distance`, `gmx angle`, `gmx gangle`

To monitor specific *bonds* in your modules, or more generally distances between points, the program `gmx distance` can calculate distances as a function of time, as well as the distribution of the distance. With a traditional index file, the groups should consist of pairs of atom numbers, for example:

```
[ bonds_1 ]
1      2
3      4
```

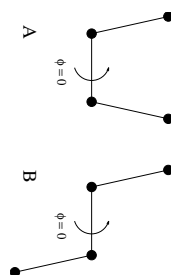


Figure 8.5: Dihedral conventions: A. “Biochemical convention”. B. “Polymer convention”.

```
9      10
```

```
[ bonds_2 ]
12      13
```

Selections are also supported, with first two positions defining the first distance, second pair of positions defining the second distance and so on. You can calculate the distances between CA and CB atoms in all your residues (assuming that every residue either has both atoms, or neither) using a selection such as:

```
name CA CB
```

The selections also allow more generic distances to be computed. For example, to compute the distances between centers of mass of two residues, you can use:

```
com of resname AAA plus com of resname BBB
```

The program `gmx angle` calculates the distribution of *angles* and *dihedrals* in time. It also gives the average angle or dihedral. The index file consists of triplets or quadruples of atom numbers:

```
[ angles ]
1      2      3
2      3      4
3      4      5
```

```
[ dihedrals ]
1      2      3      4
2      3      5      5
```

For the dihedral angles you can use either the “biochemical convention” ($\phi = 0 \equiv cis$) or “polymer convention” ($\phi = 0 \equiv trans$), see Fig. 8.5.

The program `gmx gangle` provides a selection-enabled version to compute angles. This tool can also compute angles and dihedrals, but does not support all the options of `gmx angle`, such as autocorrelation or other time series analyses. In addition, it supports angles between two vectors, a vector and a plane, two planes (defined by 2 or 3 points, respectively), a vector/plane and the

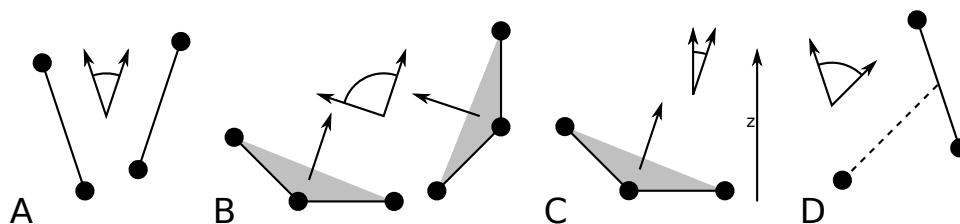


Figure 8.6: Angle options of `gmx gangle`: A. Angle between two vectors. B. Angle between two planes. C. Angle between a vector and the z axis. D. Angle between a vector and the normal of a sphere. Also other combinations are supported: planes and vectors can be used interchangeably.

z axis, or a vector/plane and the normal of a sphere (determined by a single position). Also the angle between a vector/plane compared to its position in the first frame is supported. For planes, `gmx gangle` uses the normal vector perpendicular to the plane. See Fig. 8.6A, B, C) for the definitions.

8.9 Radius of gyration and distances

`gmx gyrate`, `gmx distance`, `gmx mindist`, `gmx mdmat`, `gmx pairdist`, `gmx xpm2ps`

To have a rough measure for the compactness of a structure, you can calculate the *radius of gyration* with the program `gmx gyrate` as follows:

$$R_g = \left(\frac{\sum_i \|\mathbf{r}_i\|^2 m_i}{\sum_i m_i} \right)^{\frac{1}{2}} \quad (8.25)$$

where m_i is the mass of atom i and \mathbf{r}_i the position of atom i with respect to the center of mass of the molecule. It is especially useful to characterize polymer solutions and proteins. The program will also provide the radius of gyration around the coordinate axis (or, optionally, principal axes) by only summing the radii components orthogonal to each axis, for instance

$$R_{g,x} = \left(\frac{\sum_i (r_{i,y}^2 + r_{i,z}^2) m_i}{\sum_i m_i} \right)^{\frac{1}{2}} \quad (8.26)$$

Sometimes it is interesting to plot the *distance* between two atoms, or the *minimum* distance between two groups of atoms (*e.g.*: protein side-chains in a salt bridge). To calculate these distances between certain groups there are several possibilities:

- The *distance between the geometrical centers* of two groups can be calculated with the program `gmx distance`, as explained in sec. 8.8.
- The *minimum distance* between two groups of atoms during time can be calculated with the program `gmx mindist`. It also calculates the *number of contacts* between these groups within a certain radius r_{max} .

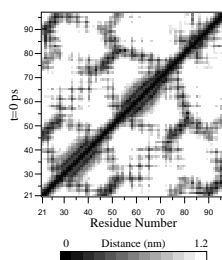


Figure 8.7: A minimum distance matrix for a peptide [174].

- `gmx pairdist` is a selection-enabled version of `gmx mindist`.
- To monitor the *minimum distances between amino acid residues* within a (protein) molecule, you can use the program `gmx mdmat`. This minimum distance between two residues A_i and A_j is defined as the smallest distance between any pair of atoms ($i \in A_i, j \in A_j$). The output is a symmetrical matrix of smallest distances between all residues. To visualize this matrix, you can use a program such as `xv`. If you want to view the axes and legend or if you want to print the matrix, you can convert it with `xpm2ps` into a Postscript picture, see Fig. 8.7.

Plotting these matrices for different time-frames, one can analyze changes in the structure, and *e.g.* forming of salt bridges.

8.10 Root mean square deviations in structure

`gmx rms`, `gmx rmsdist`

The *root mean square deviation (RMSD)* of certain atoms in a molecule with respect to a reference structure can be calculated with the program `gmx rms` by least-square fitting the structure to the reference structure ($t_2 = 0$) and subsequently calculating the *RMSD* (eqn. 8.27).

$$RMSD(t_1, t_2) = \left[\frac{1}{M} \sum_{i=1}^N m_i \|\mathbf{r}_i(t_1) - \mathbf{r}_i(t_2)\|^2 \right]^{\frac{1}{2}} \quad (8.27)$$

where $M = \sum_{i=1}^N m_i$ and $\mathbf{r}_i(t)$ is the position of atom i at time t . **Note** that fitting does not have to use the same atoms as the calculation of the *RMSD*; *e.g.* a protein is usually fitted on

the backbone atoms (N,C α ,C), but the *RMSD* can be computed of the backbone or of the whole protein.

Instead of comparing the structures to the initial structure at time $t = 0$ (so for example a crystal structure), one can also calculate eqn. 8.27 with a structure at time $t_2 = t_1 - \tau$. This gives some insight in the mobility as a function of τ . A matrix can also be made with the *RMSD* as a function of t_1 and t_2 , which gives a nice graphical interpretation of a trajectory. If there are transitions in a trajectory, they will clearly show up in such a matrix.

Alternatively the *RMSD* can be computed using a fit-free method with the program `gmx rmsdist`:

$$RMSD(t) = \left[\frac{1}{N^2} \sum_{i=1}^N \sum_{j=1}^N \|\mathbf{r}_{ij}(t) - \mathbf{r}_{ij}(0)\|^2 \right]^{\frac{1}{2}} \quad (8.28)$$

where the *distance* \mathbf{r}_{ij} between atoms at time t is compared with the distance between the same atoms at time 0.

8.11 Covariance analysis

Covariance analysis, also called principal component analysis or essential dynamics [175], can find correlated motions. It uses the covariance matrix C of the atomic coordinates:

$$C_{ij} = \left\langle M_{ii}^{\frac{1}{2}}(x_i - \langle x_i \rangle) M_{jj}^{\frac{1}{2}}(x_j - \langle x_j \rangle) \right\rangle \quad (8.29)$$

where M is a diagonal matrix containing the masses of the atoms (mass-weighted analysis) or the unit matrix (non-mass weighted analysis). C is a symmetric $3N \times 3N$ matrix, which can be diagonalized with an orthonormal transformation matrix R :

$$R^T C R = \text{diag}(\lambda_1, \lambda_2, \dots, \lambda_{3N}) \quad \text{where } \lambda_1 \geq \lambda_2 \geq \dots \geq \lambda_{3N} \quad (8.30)$$

The columns of R are the eigenvectors, also called principal or essential modes. R defines a transformation to a new coordinate system. The trajectory can be projected on the principal modes to give the principal components $p_i(t)$:

$$\mathbf{p}(t) = R^T M^{\frac{1}{2}}(\mathbf{x}(t) - \langle \mathbf{x} \rangle) \quad (8.31)$$

The eigenvalue λ_i is the mean square fluctuation of principal component i . The first few principal modes often describe collective, global motions in the system. The trajectory can be filtered along one (or more) principal modes. For one principal mode i this goes as follows:

$$\mathbf{x}^f(t) = \langle \mathbf{x} \rangle + M^{-\frac{1}{2}} R_{*i} p_i(t) \quad (8.32)$$

When the analysis is performed on a macromolecule, one often wants to remove the overall rotation and translation to look at the internal motion only. This can be achieved by least square fitting to a reference structure. Care has to be taken that the reference structure is representative for the ensemble, since the choice of reference structure influences the covariance matrix.

One should always check if the principal modes are well defined. If the first principal component resembles a half cosine and the second resembles a full cosine, you might be filtering noise (see below). A good way to check the relevance of the first few principal modes is to calculate the overlap of the sampling between the first and second half of the simulation. **Note** that this can only be done when the same reference structure is used for the two halves.

A good measure for the overlap has been defined in [176]. The elements of the covariance matrix are proportional to the square of the displacement, so we need to take the square root of the matrix to examine the extent of sampling. The square root can be calculated from the eigenvalues λ_i and the eigenvectors, which are the columns of the rotation matrix R . For a symmetric and diagonally-dominant matrix A of size $3N \times 3N$ the square root can be calculated as:

$$A^{\frac{1}{2}} = R \operatorname{diag}(\lambda_1^{\frac{1}{2}}, \lambda_2^{\frac{1}{2}}, \dots, \lambda_{3N}^{\frac{1}{2}}) R^T \quad (8.33)$$

It can be verified easily that the product of this matrix with itself gives A . Now we can define a difference d between covariance matrices A and B as follows:

$$d(A, B) = \sqrt{\operatorname{tr} \left(\left(A^{\frac{1}{2}} - B^{\frac{1}{2}} \right)^2 \right)} \quad (8.34)$$

$$= \sqrt{\operatorname{tr} \left(A + B - 2A^{\frac{1}{2}}B^{\frac{1}{2}} \right)} \quad (8.35)$$

$$= \left(\sum_{i=1}^N (\lambda_i^A + \lambda_i^B) - 2 \sum_{i=1}^N \sum_{j=1}^N \sqrt{\lambda_i^A \lambda_j^B} (R_i^A \cdot R_j^B)^2 \right)^{\frac{1}{2}} \quad (8.36)$$

where tr is the trace of a matrix. We can now define the overlap s as:

$$s(A, B) = 1 - \frac{d(A, B)}{\sqrt{\operatorname{tr}A + \operatorname{tr}B}} \quad (8.37)$$

The overlap is 1 if and only if matrices A and B are identical. It is 0 when the sampled subspaces are completely orthogonal.

A commonly-used measure is the subspace overlap of the first few eigenvectors of covariance matrices. The overlap of the subspace spanned by m orthonormal vectors $\mathbf{w}_1, \dots, \mathbf{w}_m$ with a reference subspace spanned by n orthonormal vectors $\mathbf{v}_1, \dots, \mathbf{v}_n$ can be quantified as follows:

$$\operatorname{overlap}(\mathbf{v}, \mathbf{w}) = \frac{1}{n} \sum_{i=1}^n \sum_{j=1}^m (\mathbf{v}_i \cdot \mathbf{w}_j)^2 \quad (8.38)$$

The overlap will increase with increasing m and will be 1 when set \mathbf{v} is a subspace of set \mathbf{w} . The disadvantage of this method is that it does not take the eigenvalues into account. All eigenvectors are weighted equally, and when degenerate subspaces are present (equal eigenvalues), the calculated overlap will be too low.

Another useful check is the cosine content. It has been proven that the the principal components of random diffusion are cosines with the number of periods equal to half the principal component index [177, 176]. The eigenvalues are proportional to the index to the power -2 . The cosine content is defined as:

$$\frac{2}{T} \left(\int_0^T \cos \left(\frac{i\pi t}{T} \right) p_i(t) dt \right)^2 \left(\int_0^T p_i^2(t) dt \right)^{-1} \quad (8.39)$$

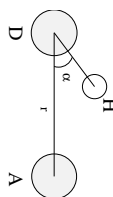


Figure 8.8: Geometrical Hydrogen bond criterion.

When the cosine content of the first few principal components is close to 1, the largest fluctuations are not connected with the potential, but with random diffusion.

The covariance matrix is built and diagonalized by `gmx covar`. The principal components and overlap (and many more things) can be plotted and analyzed with `gmx anaeig`. The cosine content can be calculated with `gmx analyze`.

8.12 Dihedral principal component analysis

`gmx angle`, `gmx covar`, `gmx anaeig`

Principal component analysis can be performed in dihedral space [178] using GROMACS. You start by defining the dihedral angles of interest in an index file, either using `gmx mk_angndx` or otherwise. Then you use the `gmx angle` program with the `-or` flag to produce a new `.trr` file containing the cosine and sine of each dihedral angle in two coordinates, respectively. That is, in the `.trr` file you will have a series of numbers corresponding to: $\cos(\phi_1)$, $\sin(\phi_1)$, $\cos(\phi_2)$, $\sin(\phi_2)$, ..., $\cos(\phi_n)$, $\sin(\phi_n)$, and the array is padded with zeros, if necessary. Then you can use this `.trr` file as input for the `gmx covar` program and perform principal component analysis as usual. For this to work you will need to generate a reference file (`.tpr`, `.gro`, `.pdb` etc.) containing the same number of “atoms” as the new `.trr` file, that is for n dihedrals you need $2n/3$ atoms (rounded up if not an integer number). You should use the `-nofit` option for `gmx covar` since the coordinates in the dummy reference file do not correspond in any way to the information in the `.trr` file. Analysis of the results is done using `gmx anaeig`.

8.13 Hydrogen bonds

`gmx hbond`

The program `gmx hbond` analyzes the *hydrogen bonds* (H-bonds) between all possible donors D and acceptors A. To determine if an H-bond exists, a geometrical criterion is used, see also Fig. 8.8:

$$\begin{aligned} r &\leq r_{HB} = 0.35 \text{ nm} \\ \alpha &\leq \alpha_{HB} = 30^\circ \end{aligned} \quad (8.40)$$

The value of $r_{HB} = 0.35$ nm corresponds to the first minimum of the RDF of SPC water (see also Fig. 8.3).

The program `gmx hbond` analyzes all hydrogen bonds existing between two groups of atoms (which must be either identical or non-overlapping) or in specified donor-hydrogen-acceptor triplets,

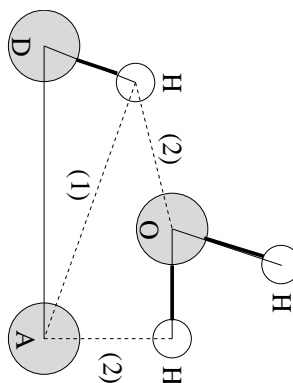


Figure 8.9: Insertion of water into an H-bond. (1) Normal H-bond between two residues. (2) H-bonding bridge via a water molecule.

in the following ways:

- Donor-Acceptor distance (r) distribution of all H-bonds
- Hydrogen-Donor-Acceptor angle (α) distribution of all H-bonds
- The total number of H-bonds in each time frame
- The number of H-bonds in time between residues, divided into groups $n-n+i$ where n and $n+i$ stand for residue numbers and i goes from 0 to 6. The group for $i = 6$ also includes all H-bonds for $i > 6$. These groups include the $n-n+3$, $n-n+4$ and $n-n+5$ H-bonds, which provide a measure for the formation of α -helices or β -turns or strands.
- The lifetime of the H-bonds is calculated from the average over all autocorrelation functions of the existence functions (either 0 or 1) of all H-bonds:

$$C(\tau) = \langle s_i(t) s_i(t + \tau) \rangle \quad (8.41)$$

with $s_i(t) = \{0, 1\}$ for H-bond i at time t . The integral of $C(\tau)$ gives a rough estimate of the average H-bond lifetime τ_{HB} :

$$\tau_{HB} = \int_0^{\infty} C(\tau) d\tau \quad (8.42)$$

Both the integral and the complete autocorrelation function $C(\tau)$ will be output, so that more sophisticated analysis (*e.g.* using multi-exponential fits) can be used to get better estimates for τ_{HB} . A more complete analysis is given in ref. [179]; one of the more fancy options is the Luzar and Chandler analysis of hydrogen bond kinetics [180, 181].

- An H-bond existence map can be generated of dimensions $\#H\text{-bonds} \times \#frames$. The ordering is identical to the index file (see below), but reversed, meaning that the last triplet in the index file corresponds to the first row of the existence map.
- Index groups are output containing the analyzed groups, all donor-hydrogen atom pairs and acceptor atoms in these groups, donor-hydrogen-acceptor triplets involved in hydrogen bonds between the analyzed groups and all solvent atoms involved in insertion.

8.14 Protein-related items

`gmx do_dssp`, `gmx rama`, `gmx wheel`

To analyze structural changes of a protein, you can calculate the radius of gyration or the minimum residue distances over time (see sec. 8.9), or calculate the RMSD (sec. 8.10).

You can also look at the changing of *secondary structure elements* during your run. For this, you can use the program `gmx do_dssp`, which is an interface for the commercial program DSSP [182]. For further information, see the DSSP manual. A typical output plot of `gmx do_dssp` is given in Fig. 8.10.

One other important analysis of proteins is the so-called *Ramachandran plot*. This is the projection of the structure on the two dihedral angles ϕ and ψ of the protein backbone, see Fig. 8.11.

To evaluate this Ramachandran plot you can use the program `gmx rama`. A typical output is given in Fig. 8.12.

When studying α -helices it is useful to have a *helical wheel* projection of your peptide, to see whether a peptide is amphipathic. This can be done using the `gmx wheel` program. Two examples are plotted in Fig. 8.13.

8.15 Interface-related items

`gmx order`, `gmx density`, `gmx potential`, `gmx traj`

When simulating molecules with long carbon tails, it can be interesting to calculate their average orientation. There are several flavors of order parameters, most of which are related. The program `gmx order` can calculate order parameters using the equation:

$$S_z = \frac{3}{2} \langle \cos^2 \theta_z \rangle - \frac{1}{2} \quad (8.43)$$

where θ_z is the angle between the z -axis of the simulation box and the molecular axis under consideration. The latter is defined as the vector from C_{n-1} to C_{n+1} . The parameters S_x and S_y are defined in the same way. The brackets imply averaging over time and molecules. Order parameters can vary between 1 (full order along the interface normal) and $-1/2$ (full order perpendicular to the normal), with a value of zero in the case of isotropic orientation.

The program can do two things for you. It can calculate the order parameter for each CH_2 segment separately, for any of three axes, or it can divide the box in slices and calculate the average value of the order parameter per segment in one slice. The first method gives an idea of the ordering of a molecule from head to tail, the second method gives an idea of the ordering as function of the box length.

The electrostatic potential (ψ) across the interface can be computed from a trajectory by evaluating the double integral of the charge density ($\rho(z)$):

$$\psi(z) - \psi(-\infty) = - \int_{-\infty}^z dz' \int_{-\infty}^{z'} \rho(z'') dz'' / \epsilon_0 \quad (8.44)$$

where the position $z = -\infty$ is far enough in the bulk phase such that the field is zero. With this method, it is possible to “split” the total potential into separate contributions from lipid and water

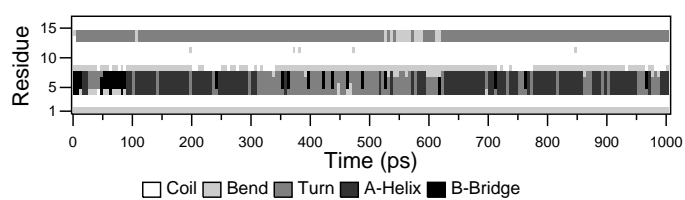


Figure 8.10: Analysis of the secondary structure elements of a peptide in time.

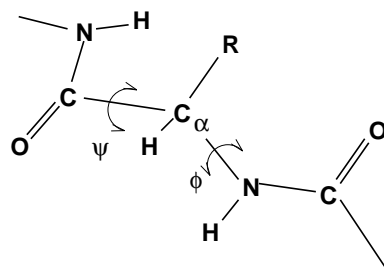


Figure 8.11: Definition of the dihedral angles ϕ and ψ of the protein backbone.

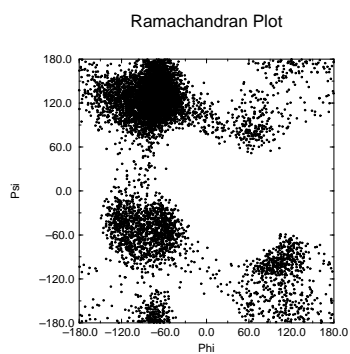


Figure 8.12: Ramachandran plot of a small protein.

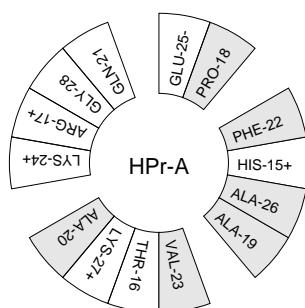


Figure 8.13: Helical wheel projection of the N-terminal helix of HPr.

molecules. The program `gmx potential` divides the box in slices and sums all charges of the atoms in each slice. It then integrates this charge density to give the electric field, which is in turn integrated to give the potential. Charge density, electric field, and potential are written to `xvgr` input files.

The program `gmx traj` is a very simple analysis program. All it does is print the coordinates, velocities, or forces of selected atoms. It can also calculate the center of mass of one or more molecules and print the coordinates of the center of mass to three files. By itself, this is probably not a very useful analysis, but having the coordinates of selected molecules or atoms can be very handy for further analysis, not only in interfacial systems.

The program `gmx density` calculates the mass density of groups and gives a plot of the density against a box axis. This is useful for looking at the distribution of groups or atoms across the interface.

Appendix A

Some implementation details

In this chapter we will present some implementation details. This is far from complete, but we deemed it necessary to clarify some things that would otherwise be hard to understand.

A.1 Single Sum Virial in GROMACS

The virial Ξ can be written in full tensor form as:

$$\Xi = -\frac{1}{2} \sum_{i < j}^N \mathbf{r}_{ij} \otimes \mathbf{F}_{ij} \quad (\text{A.1})$$

where \otimes denotes the *direct product* of two vectors.¹ When this is computed in the inner loop of an MD program 9 multiplications and 9 additions are needed.²

Here it is shown how it is possible to extract the virial calculation from the inner loop [183].

A.1.1 Virial

In a system with periodic boundary conditions, the periodicity must be taken into account for the virial:

$$\Xi = -\frac{1}{2} \sum_{i < j}^N \mathbf{r}_{ij}^n \otimes \mathbf{F}_{ij} \quad (\text{A.2})$$

where \mathbf{r}_{ij}^n denotes the distance vector of the *nearest image* of atom i from atom j . In this definition we add a *shift vector* δ_i to the position vector \mathbf{r}_i of atom i . The difference vector \mathbf{r}_{ij}^n is thus equal to:

$$\mathbf{r}_{ij}^n = \mathbf{r}_i + \delta_i - \mathbf{r}_j \quad (\text{A.3})$$

or in shorthand:

$$\mathbf{r}_{ij}^n = \mathbf{r}_i^n - \mathbf{r}_j \quad (\text{A.4})$$

¹ $(\mathbf{u} \otimes \mathbf{v})^{\alpha\beta} = \mathbf{u}_\alpha \mathbf{v}_\beta$

²The calculation of Lennard-Jones and Coulomb forces is about 50 floating point operations.

In a triclinic system, there are 27 possible images of i ; when a truncated octahedron is used, there are 15 possible images.

A.1.2 Virial from non-bonded forces

Here the derivation for the single sum virial in the *non-bonded force* routine is given. There are a couple of considerations that are special to GROMACS that we take into account:

- When calculating short-range interactions, we apply the *minimum image convention* and only consider the closest image of each neighbor - and in particular we never allow interactions between a particle and any of its periodic images. For all the equations below, this means $i \neq j$.
- In general, either the i or j particle might be shifted to a neighbor cell to get the closest interaction (shift δ_{ij}). However, with minimum image convention there can be at most 27 different shifts for particles in the central cell, and for typical (very short-ranged) biomolecular interactions there are typically only a few different shifts involved for each particle, not to mention that each interaction can only be present for one shift.
- For the GROMACS nonbonded interactions we use this to split the neighborlist of each i particle into multiple separate lists, where each list has a constant shift δ_i for the i particle. We can represent this as a sum over shifts (for which we use index s), with the constraint that each particle interaction can only contribute to one of the terms in this sum, and the shift is no longer dependent on the j particles. For any sum that does not contain complex dependence on s , this means the sum trivially reduces to just the sum over i and/or j .
- To simplify some of the sums, we replace sums over $j < i$ with double sums over all particles (remember, $i \neq j$) and divide by 2.

Starting from the above definition of the virial, we then get

$$\begin{aligned}
\Xi &= -\frac{1}{2} \sum_{i < j}^N \mathbf{r}_{ij}^n \otimes \mathbf{F}_{ij} \\
&= -\frac{1}{2} \sum_{i < j}^N (\mathbf{r}_i + \delta_{ij} - \mathbf{r}_j) \otimes \mathbf{F}_{ij} \\
&= -\frac{1}{4} \sum_{i=1}^N \sum_{j=1}^N (\mathbf{r}_i + \delta_{ij} - \mathbf{r}_j) \otimes \mathbf{F}_{ij} \\
&= -\frac{1}{4} \sum_{i=1}^N \sum_s \sum_{j=1}^N (\mathbf{r}_i + \delta_{i,s} - \mathbf{r}_j) \otimes \mathbf{F}_{ij,s} \\
&= -\frac{1}{4} \sum_{i=1}^N \sum_s \sum_{j=1}^N ((\mathbf{r}_i + \delta_{i,s}) \otimes \mathbf{F}_{ij,s} - \mathbf{r}_j \otimes \mathbf{F}_{ij,s}) \\
&= -\frac{1}{4} \sum_{i=1}^N \sum_s \sum_{j=1}^N (\mathbf{r}_i + \delta_{i,s}) \otimes \mathbf{F}_{ij,s} + \frac{1}{4} \sum_{i=1}^N \sum_s \sum_{j=1}^N \mathbf{r}_j \otimes \mathbf{F}_{ij,s}
\end{aligned}$$

$$\begin{aligned}
&= -\frac{1}{4} \sum_{i=1}^N \sum_s \sum_{j=1}^N (\mathbf{r}_i + \delta_{i,s}) \otimes \mathbf{F}_{ij,s} + \frac{1}{4} \sum_{i=1}^N \sum_{j=1}^N \mathbf{r}_j \otimes \mathbf{F}_{ij} \\
&= -\frac{1}{4} \sum_s \sum_{i=1}^N (\mathbf{r}_i + \delta_{i,s}) \otimes \sum_{j=1}^N \mathbf{F}_{ij,s} + \frac{1}{4} \sum_{j=1}^N \mathbf{r}_j \otimes \sum_{i=1}^N \mathbf{F}_{ij} \\
&= -\frac{1}{4} \sum_s \sum_{i=1}^N (\mathbf{r}_i + \delta_{i,s}) \otimes \sum_{j=1}^N \mathbf{F}_{ij,s} - \frac{1}{4} \sum_{j=1}^N \mathbf{r}_j \otimes \sum_{i=1}^N \mathbf{F}_{ji} \\
&= -\frac{1}{4} \sum_s \sum_{i=1}^N (\mathbf{r}_i + \delta_{i,s}) \otimes \mathbf{F}_{i,s} - \frac{1}{4} \sum_{j=1}^N \mathbf{r}_j \otimes \mathbf{F}_j \\
&= -\frac{1}{4} \left(\sum_{i=1}^N \mathbf{r}_i \otimes \mathbf{F}_i + \sum_{j=1}^N \mathbf{r}_j \otimes \mathbf{F}_j \right) - \frac{1}{4} \sum_s \sum_{i=1}^N \delta_{i,s} \otimes \mathbf{F}_{i,s} \\
&= -\frac{1}{2} \sum_{i=1}^N \mathbf{r}_i \otimes \mathbf{F}_i - \frac{1}{4} \sum_s \sum_{i=1}^N \delta_{i,s} \otimes \mathbf{F}_{i,s} \\
&= -\frac{1}{2} \sum_{i=1}^N \mathbf{r}_i \otimes \mathbf{F}_i - \frac{1}{4} \sum_s \delta_s \otimes \mathbf{F}_s \\
&= \Xi_0 + \Xi_1
\end{aligned} \tag{A.5}$$

In the second-last stage, we have used the property that each shift vector itself does not depend on the coordinates of particle i , so it is possible to sum up all forces corresponding to each shift vector (in the nonbonded kernels), and then just use a sum over the different shift vectors outside the kernels. We have also used

$$\mathbf{F}_i = \sum_{j=1}^N \mathbf{F}_{ij} \tag{A.6}$$

$$\mathbf{F}_j = \sum_{i=1}^N \mathbf{F}_{ji} \tag{A.7}$$

which is the total force on i with respect to j . Because we use Newton's Third Law:

$$\mathbf{F}_{ij} = -\mathbf{F}_{ji} \tag{A.8}$$

we must, in the implementation, double the term containing the shift δ_i . Similarly, in a few places we have summed the shift-dependent force over all shifts to come up with the total force per interaction or particle.

This separates the total virial Ξ into a component Ξ_0 that is a single sum over particles, and a second component Ξ_1 that describes the influence of the particle shifts, and that is only a sum over the different shift vectors.

A.1.3 The intra-molecular shift (mol-shift)

For the bonded forces and SHAKE it is possible to make a *mol-shift* list, in which the periodicity is stored. We simply have an array `mshift` in which for each atom an index in the `shiftvec` array is stored.

The algorithm to generate such a list can be derived from graph theory, considering each particle in a molecule as a bead in a graph, the bonds as edges.

- 1 Represent the bonds and atoms as bidirectional graph
- 2 Make all atoms white
- 3 Make one of the white atoms black (atom i) and put it in the central box
- 4 Make all of the neighbors of i that are currently white, gray
- 5 Pick one of the gray atoms (atom j), give it the correct periodicity with respect to any of its black neighbors and make it black
- 6 Make all of the neighbors of j that are currently white, gray
- 7 If any gray atom remains, go to [5]
- 8 If any white atom remains, go to [3]

Using this algorithm we can

- optimize the bonded force calculation as well as SHAKE
- calculate the virial from the bonded forces in the single sum method again

Find a representation of the bonds as a bidirectional graph.

A.1.4 Virial from Covalent Bonds

Since the covalent bond force gives a contribution to the virial, we have:

$$b = \|\mathbf{r}_{ij}^n\| \quad (\text{A.9})$$

$$V_b = \frac{1}{2}k_b(b - b_0)^2 \quad (\text{A.10})$$

$$\mathbf{F}_i = -\nabla V_b \quad (\text{A.11})$$

$$= k_b(b - b_0)\frac{\mathbf{r}_{ij}^n}{b} \quad (\text{A.12})$$

$$\mathbf{F}_j = -\mathbf{F}_i \quad (\text{A.13})$$

The virial contribution from the bonds then is:

$$\Xi_b = -\frac{1}{2}(\mathbf{r}_i^n \otimes \mathbf{F}_i + \mathbf{r}_j^n \otimes \mathbf{F}_j) \quad (\text{A.14})$$

$$= -\frac{1}{2}\mathbf{r}_{ij}^n \otimes \mathbf{F}_i \quad (\text{A.15})$$

A.1.5 Virial from SHAKE

An important contribution to the virial comes from shake. Satisfying the constraints a force \mathbf{G} that is exerted on the particles “shaken.” If this force does not come out of the algorithm (as in standard SHAKE) it can be calculated afterward (when using *leap-frog*) by:

$$\Delta \mathbf{r}_i = \mathbf{r}_i(t + \Delta t) - [\mathbf{r}_i(t) + \mathbf{v}_i(t - \frac{\Delta t}{2})\Delta t + \frac{\mathbf{F}_i}{m_i}\Delta t^2] \quad (\text{A.16})$$

$$\mathbf{G}_i = \frac{m_i \Delta \mathbf{r}_i}{\Delta t^2} \quad (\text{A.17})$$

This does not help us in the general case. Only when no periodicity is needed (like in rigid water) this can be used, otherwise we must add the virial calculation in the inner loop of SHAKE.

When it is applicable the virial can be calculated in the single sum way:

$$\Xi = -\frac{1}{2} \sum_i^{N_c} \mathbf{r}_i \otimes \mathbf{F}_i \quad (\text{A.18})$$

where N_c is the number of constrained atoms.

A.2 Optimizations

Here we describe some of the algorithmic optimizations used in GROMACS, apart from parallelism.

A.2.1 Inner Loops for Water

GROMACS uses special inner loops to calculate non-bonded interactions for water molecules with other atoms, and yet another set of loops for interactions between pairs of water molecules. There highly optimized loops for two types of water models. For three site models similar to SPC [85], *i.e.*:

1. There are three atoms in the molecule.
2. The whole molecule is a single charge group.
3. The first atom has Lennard-Jones (sec. 4.1.1) and Coulomb (sec. 4.1.3) interactions.
4. Atoms two and three have only Coulomb interactions, and equal charges.

These loops also works for the SPC/E [184] and TIP3P [133] water models. And for four site water models similar to TIP4P [133]:

1. There are four atoms in the molecule.
2. The whole molecule is a single charge group.

3. The first atom has only Lennard-Jones (sec. 4.1.1) interactions.
4. Atoms two and three have only Coulomb (sec. 4.1.3) interactions, and equal charges.
5. Atom four has only Coulomb interactions.

The benefit of these implementations is that there are more floating-point operations in a single loop, which implies that some compilers can schedule the code better. However, it turns out that even some of the most advanced compilers have problems with scheduling, implying that manual tweaking is necessary to get optimum performance. This may include common-sub-expression elimination, or moving code around.

Appendix B

Averages and fluctuations

B.1 Formulae for averaging

Note: this section was taken from ref [185].

When analyzing a MD trajectory averages $\langle x \rangle$ and fluctuations

$$\langle (\Delta x)^2 \rangle^{\frac{1}{2}} = \langle [x - \langle x \rangle]^2 \rangle^{\frac{1}{2}} \quad (\text{B.1})$$

of a quantity x are to be computed. The variance σ_x of a series of N_x values, $\{x_i\}$, can be computed from

$$\sigma_x = \sum_{i=1}^{N_x} x_i^2 - \frac{1}{N_x} \left(\sum_{i=1}^{N_x} x_i \right)^2 \quad (\text{B.2})$$

Unfortunately this formula is numerically not very accurate, especially when $\sigma_x^{\frac{1}{2}}$ is small compared to the values of x_i . The following (equivalent) expression is numerically more accurate

$$\sigma_x = \sum_{i=1}^{N_x} [x_i - \langle x \rangle]^2 \quad (\text{B.3})$$

with

$$\langle x \rangle = \frac{1}{N_x} \sum_{i=1}^{N_x} x_i \quad (\text{B.4})$$

Using eqns. B.2 and B.4 one has to go through the series of x_i values twice, once to determine $\langle x \rangle$ and again to compute σ_x , whereas eqn. B.1 requires only one sequential scan of the series $\{x_i\}$. However, one may cast eqn. B.2 in another form, containing partial sums, which allows for a sequential update algorithm. Define the partial sum

$$X_{n,m} = \sum_{i=n}^m x_i \quad (\text{B.5})$$

and the partial variance

$$\sigma_{n,m} = \sum_{i=n}^m \left[x_i - \frac{X_{n,m}}{m-n+1} \right]^2 \quad (\text{B.6})$$

It can be shown that

$$X_{n,m+k} = X_{n,m} + X_{m+1,m+k} \quad (\text{B.7})$$

and

$$\sigma_{n,m+k} = \sigma_{n,m} + \sigma_{m+1,m+k} + \left[\frac{X_{n,m}}{m-n+1} - \frac{X_{n,m+k}}{m+k-n+1} \right]^2 * \frac{(m-n+1)(m+k-n+1)}{k} \quad (\text{B.8})$$

For $n = 1$ one finds

$$\sigma_{1,m+k} = \sigma_{1,m} + \sigma_{m+1,m+k} + \left[\frac{X_{1,m}}{m} - \frac{X_{1,m+k}}{m+k} \right]^2 \frac{m(m+k)}{k} \quad (\text{B.9})$$

and for $n = 1$ and $k = 1$ (eqn. B.8) becomes

$$\sigma_{1,m+1} = \sigma_{1,m} + \left[\frac{X_{1,m}}{m} - \frac{X_{1,m+1}}{m+1} \right]^2 m(m+1) \quad (\text{B.10})$$

$$= \sigma_{1,m} + \frac{[X_{1,m} - mx_{m+1}]^2}{m(m+1)} \quad (\text{B.11})$$

where we have used the relation

$$X_{1,m+1} = X_{1,m} + x_{m+1} \quad (\text{B.12})$$

Using formulae (eqn. B.11) and (eqn. B.12) the average

$$\langle x \rangle = \frac{X_{1,N_x}}{N_x} \quad (\text{B.13})$$

and the fluctuation

$$\langle (\Delta x)^2 \rangle^{\frac{1}{2}} = \left[\frac{\sigma_{1,N_x}}{N_x} \right]^{\frac{1}{2}} \quad (\text{B.14})$$

can be obtained by one sweep through the data.

B.2 Implementation

In GROMACS the instantaneous energies $E(m)$ are stored in the energy file, along with the values of $\sigma_{1,m}$ and $X_{1,m}$. Although the steps are counted from 0, for the energy and fluctuations steps are counted from 1. This means that the equations presented here are the ones that are implemented. We give somewhat lengthy derivations in this section to simplify checking of code and equations later on.

B.2.1 Part of a Simulation

It is not uncommon to perform a simulation where the first part, *e.g.* 100 ps, is taken as equilibration. However, the averages and fluctuations as printed in the log file are computed over the whole simulation. The equilibration time, which is now part of the simulation, may in such a case invalidate the averages and fluctuations, because these numbers are now dominated by the initial drift towards equilibrium.

Using eqns. B.7 and B.8 the average and standard deviation over part of the trajectory can be computed as:

$$X_{m+1,m+k} = X_{1,m+k} - X_{1,m} \quad (\text{B.15})$$

$$\sigma_{m+1,m+k} = \sigma_{1,m+k} - \sigma_{1,m} - \left[\frac{X_{1,m}}{m} - \frac{X_{1,m+k}}{m+k} \right]^2 \frac{m(m+k)}{k} \quad (\text{B.16})$$

or, more generally (with $p \geq 1$ and $q \geq p$):

$$X_{p,q} = X_{1,q} - X_{1,p-1} \quad (\text{B.17})$$

$$\sigma_{p,q} = \sigma_{1,q} - \sigma_{1,p-1} - \left[\frac{X_{1,p-1}}{p-1} - \frac{X_{1,q}}{q} \right]^2 \frac{(p-1)q}{q-p+1} \quad (\text{B.18})$$

Note that implementation of this is not entirely trivial, since energies are not stored every time step of the simulation. We therefore have to construct $X_{1,p-1}$ and $\sigma_{1,p-1}$ from the information at time p using eqns. B.11 and B.12:

$$X_{1,p-1} = X_{1,p} - x_p \quad (\text{B.19})$$

$$\sigma_{1,p-1} = \sigma_{1,p} - \frac{[X_{1,p-1} - (p-1)x_p]^2}{(p-1)p} \quad (\text{B.20})$$

B.2.2 Combining two simulations

Another frequently occurring problem is, that the fluctuations of two simulations must be combined. Consider the following example: we have two simulations (A) of n and (B) of m steps, in which the second simulation is a continuation of the first. However, the second simulation starts numbering from 1 instead of from $n+1$. For the partial sum this is no problem, we have to add $X_{1,n}^A$ from run A:

$$X_{1,n+m}^{AB} = X_{1,n}^A + X_{1,m}^B \quad (\text{B.21})$$

When we want to compute the partial variance from the two components we have to make a correction $\Delta\sigma$:

$$\sigma_{1,n+m}^{AB} = \sigma_{1,n}^A + \sigma_{1,m}^B + \Delta\sigma \quad (\text{B.22})$$

if we define x_i^{AB} as the combined and renumbered set of data points we can write:

$$\sigma_{1,n+m}^{AB} = \sum_{i=1}^{n+m} \left[x_i^{AB} - \frac{X_{1,n+m}^{AB}}{n+m} \right]^2 \quad (\text{B.23})$$

and thus

$$\sum_{i=1}^{n+m} \left[x_i^{AB} - \frac{X_{1,n+m}^{AB}}{n+m} \right]^2 = \sum_{i=1}^n \left[x_i^A - \frac{X_{1,n}^A}{n} \right]^2 + \sum_{i=1}^m \left[x_i^B - \frac{X_{1,m}^B}{m} \right]^2 + \Delta\sigma \quad (\text{B.24})$$

or

$$\begin{aligned} & \sum_{i=1}^{n+m} \left[(x_i^{AB})^2 - 2x_i^{AB} \frac{X_{1,n+m}^{AB}}{n+m} + \left(\frac{X_{1,n+m}^{AB}}{n+m} \right)^2 \right] - \\ & \sum_{i=1}^n \left[(x_i^A)^2 - 2x_i^A \frac{X_{1,n}^A}{n} + \left(\frac{X_{1,n}^A}{n} \right)^2 \right] - \\ & \sum_{i=1}^m \left[(x_i^B)^2 - 2x_i^B \frac{X_{1,m}^B}{m} + \left(\frac{X_{1,m}^B}{m} \right)^2 \right] = \Delta\sigma \end{aligned} \quad (\text{B.25})$$

all the x_i^2 terms drop out, and the terms independent of the summation counter i can be simplified:

$$\begin{aligned} & \frac{(X_{1,n+m}^{AB})^2}{n+m} - \frac{(X_{1,n}^A)^2}{n} - \frac{(X_{1,m}^B)^2}{m} - \\ & 2 \frac{X_{1,n+m}^{AB}}{n+m} \sum_{i=1}^{n+m} x_i^{AB} + 2 \frac{X_{1,n}^A}{n} \sum_{i=1}^n x_i^A + 2 \frac{X_{1,m}^B}{m} \sum_{i=1}^m x_i^B = \Delta\sigma \end{aligned} \quad (\text{B.26})$$

we recognize the three partial sums on the second line and use eqn. B.21 to obtain:

$$\Delta\sigma = \frac{(mX_{1,n}^A - nX_{1,m}^B)^2}{nm(n+m)} \quad (\text{B.27})$$

if we check this by inserting $m = 1$ we get back eqn. B.11

B.2.3 Summing energy terms

The gmx energy program can also sum energy terms into one, *e.g.* potential + kinetic = total. For the partial averages this is again easy if we have S energy components s :

$$X_{m,n}^S = \sum_{i=m}^n \sum_{s=1}^S x_i^s = \sum_{s=1}^S \sum_{i=m}^n x_i^s = \sum_{s=1}^S X_{m,n}^s \quad (\text{B.28})$$

For the fluctuations it is less trivial again, considering for example that the fluctuation in potential and kinetic energy should cancel. Nevertheless we can try the same approach as before by writing:

$$\sigma_{m,n}^S = \sum_{s=1}^S \sigma_{m,n}^s + \Delta\sigma \quad (\text{B.29})$$

if we fill in eqn. B.6:

$$\sum_{i=m}^n \left[\left(\sum_{s=1}^S x_i^s \right) - \frac{X_{m,n}^S}{m-n+1} \right]^2 = \sum_{s=1}^S \sum_{i=m}^n \left[(x_i^s) - \frac{X_{m,n}^s}{m-n+1} \right]^2 + \Delta\sigma \quad (\text{B.30})$$

which we can expand to:

$$\begin{aligned} & \sum_{i=m}^n \left[\sum_{s=1}^S (x_i^s)^2 + \left(\frac{X_{m,n}^S}{m-n+1} \right)^2 - 2 \left(\frac{X_{m,n}^S}{m-n+1} \sum_{s=1}^S x_i^s + \sum_{s=1}^S \sum_{s'=s+1}^S x_i^s x_i^{s'} \right) \right] \\ & - \sum_{s=1}^S \sum_{i=m}^n \left[(x_i^s)^2 - 2 \frac{X_{m,n}^s}{m-n+1} x_i^s + \left(\frac{X_{m,n}^s}{m-n+1} \right)^2 \right] = \Delta\sigma \end{aligned} \quad (\text{B.31})$$

the terms with $(x_i^s)^2$ cancel, so that we can simplify to:

$$\begin{aligned} & \frac{\left(X_{m,n}^S \right)^2}{m-n+1} - 2 \frac{X_{m,n}^S}{m-n+1} \sum_{i=m}^n \sum_{s=1}^S x_i^s - 2 \sum_{i=m}^n \sum_{s=1}^S \sum_{s'=s+1}^S x_i^s x_i^{s'} - \\ & \sum_{s=1}^S \sum_{i=m}^n \left[-2 \frac{X_{m,n}^s}{m-n+1} x_i^s + \left(\frac{X_{m,n}^s}{m-n+1} \right)^2 \right] = \Delta\sigma \end{aligned} \quad (\text{B.32})$$

or

$$-\frac{\left(X_{m,n}^S \right)^2}{m-n+1} - 2 \sum_{i=m}^n \sum_{s=1}^S \sum_{s'=s+1}^S x_i^s x_i^{s'} + \sum_{s=1}^S \frac{\left(X_{m,n}^s \right)^2}{m-n+1} = \Delta\sigma \quad (\text{B.33})$$

If we now expand the first term using eqn. B.28 we obtain:

$$-\frac{\left(\sum_{s=1}^S X_{m,n}^s \right)^2}{m-n+1} - 2 \sum_{i=m}^n \sum_{s=1}^S \sum_{s'=s+1}^S x_i^s x_i^{s'} + \sum_{s=1}^S \frac{\left(X_{m,n}^s \right)^2}{m-n+1} = \Delta\sigma \quad (\text{B.34})$$

which we can reformulate to:

$$-2 \left[\sum_{s=1}^S \sum_{s'=s+1}^S X_{m,n}^s X_{m,n}^{s'} + \sum_{i=m}^n \sum_{s=1}^S \sum_{s'=s+1}^S x_i^s x_i^{s'} \right] = \Delta\sigma \quad (\text{B.35})$$

or

$$-2 \left[\sum_{s=1}^S X_{m,n}^s \sum_{s'=s+1}^S X_{m,n}^{s'} + \sum_{s=1}^S \sum_{i=m}^n x_i^s \sum_{s'=s+1}^S x_i^{s'} \right] = \Delta\sigma \quad (\text{B.36})$$

which gives

$$-2 \sum_{s=1}^S \left[X_{m,n}^s \sum_{s'=s+1}^S \sum_{i=m}^n x_i^{s'} + \sum_{i=m}^n x_i^s \sum_{s'=s+1}^S x_i^{s'} \right] = \Delta\sigma \quad (\text{B.37})$$

Since we need all data points i to evaluate this, in general this is not possible. We can then make an estimate of $\sigma_{m,n}^S$ using only the data points that are available using the left hand side of eqn. B.30. While the average can be computed using all time steps in the simulation, the accuracy of the fluctuations is thus limited by the frequency with which energies are saved. Since this can be easily done with a program such as xmgr this is not built-in in GROMACS.

Bibliography

- [1] Bekker, H., Berendsen, H. J. C., Dijkstra, E. J., Achterop, S., van Drunen, R., van der Spoel, D., Sijbers, A., Keegstra, H., Reitsma, B., Renardus, M. K. R. Gromacs: A parallel computer for molecular dynamics simulations. In *Physics Computing 92* (Singapore, 1993). de Groot, R. A., Nadrchal, J., eds. . World Scientific.
- [2] Berendsen, H. J. C., van der Spoel, D., van Drunen, R. GROMACS: A message-passing parallel molecular dynamics implementation. *Comp. Phys. Comm.* 91:43–56, 1995.
- [3] Lindahl, E., Hess, B., van der Spoel, D. GROMACS 3.0: A package for molecular simulation and trajectory analysis. *J. Mol. Mod.* 7:306–317, 2001.
- [4] van der Spoel, D., Lindahl, E., Hess, B., Groenhof, G., Mark, A. E., Berendsen, H. J. C. GROMACS: Fast, Flexible and Free. *J. Comp. Chem.* 26:1701–1718, 2005.
- [5] Hess, B., Kutzner, C., van der Spoel, D., Lindahl, E. GROMACS 4: Algorithms for Highly Efficient, Load-Balanced, and Scalable Molecular Simulation. *J. Chem. Theory Comput.* 4(3):435–447, 2008.
- [6] Pronk, S., Páll, S., Schulz, R., Larsson, P., Bjelkmar, P., Apostolov, R., Shirts, M. R., Smith, J. C., Kasson, P. M., van der Spoel, D., Hess, B., Lindahl, E. GROMACS 4.5: a high-throughput and highly parallel open source molecular simulation toolkit. *Bioinformatics* 29(7):845–854, 2013.
- [7] Páll, S., Abraham, M. J., Kutzner, C., Hess, B., Lindahl, E. Tackling exascale software challenges in molecular dynamics simulations with GROMACS. In: *Solving Software Challenges for Exascale*. Vol. 8759. Markidis, S., Laure, E. eds. Vol. 8759. . Springer International Publishing Switzerland London 2015 3–27.
- [8] Abraham, M. J., Murtola, T., Schulz, R., Páll, S., Smith, J. C., Hess, B., Lindahl, E. GROMACS: High performance molecular simulations through multi-level parallelism from laptops to supercomputers. *SoftwareX* 1–2:19–25, 2015.
- [9] van Gunsteren, W. F., Berendsen, H. J. C. Computer simulation of molecular dynamics: Methodology, applications, and perspectives in chemistry. *Angew. Chem. Int. Ed. Engl.* 29:992–1023, 1990.
- [10] Fraaije, J. G. E. M. Dynamic density functional theory for microphase separation kinetics of block copolymer melts. *J. Chem. Phys.* 99:9202–9212, 1993.

- [11] McQuarrie, D. A. *Statistical Mechanics*. New York: Harper & Row. 1976.
- [12] van Gunsteren, W. F., Berendsen, H. J. C. Algorithms for macromolecular dynamics and constraint dynamics. *Mol. Phys.* 34:1311–1327, 1977.
- [13] van Gunsteren, W. F., Karplus, M. Effect of constraints on the dynamics of macromolecules. *Macromolecules* 15:1528–1544, 1982.
- [14] Darden, T., York, D., Pedersen, L. Particle mesh Ewald: An $N \log(N)$ method for Ewald sums in large systems. *J. Chem. Phys.* 98:10089–10092, 1993.
- [15] Essmann, U., Perera, L., Berkowitz, M. L., Darden, T., Lee, H., Pedersen, L. G. A smooth particle mesh ewald potential. *J. Chem. Phys.* 103:8577–8592, 1995.
- [16] Geman, S., Geman, D. Stochastic relaxation, Gibbs distributions and the Bayesian restoration of images. *IEEE Trans. Patt. Anal. Mach. Int.* 6:721, 1984.
- [17] Nilges, M., Clore, G. M., Gronenborn, A. M. Determination of three-dimensional structures of proteins from interproton distance data by dynamical simulated annealing from a random array of atoms. *FEBS Lett.* 239:129–136, 1988.
- [18] van Schaik, R. C., Berendsen, H. J. C., Torda, A. E., van Gunsteren, W. F. A structure refinement method based on molecular dynamics in 4 spatial dimensions. *J. Mol. Biol.* 234:751–762, 1993.
- [19] Zimmerman, K. All purpose molecular mechanics simulator and energy minimizer. *J. Comp. Chem.* 12:310–319, 1991.
- [20] Adams, D. J., Adams, E. M., Hills, G. J. The computer simulation of polar liquids. *Mol. Phys.* 38:387–400, 1979.
- [21] Bekker, H., Dijkstra, E. J., Renardus, M. K. R., Berendsen, H. J. C. An efficient, box shape independent non-bonded force and virial algorithm for molecular dynamics. *Mol. Sim.* 14:137–152, 1995.
- [22] Hockney, R. W., Goel, S. P., Eastwood, J. Quiet High Resolution Computer Models of a Plasma. *J. Comp. Phys.* 14:148–158, 1974.
- [23] Verlet, L. Computer experiments on classical fluids. I. Thermodynamical properties of Lennard-Jones molecules. *Phys. Rev.* 159:98–103, 1967.
- [24] Berendsen, H. J. C., van Gunsteren, W. F. Practical algorithms for dynamics simulations.
- [25] Swope, W. C., Andersen, H. C., Berens, P. H., Wilson, K. R. A computer-simulation method for the calculation of equilibrium-constants for the formation of physical clusters of molecules: Application to small water clusters. *J. Chem. Phys.* 76:637–649, 1982.
- [26] Berendsen, H. J. C., Postma, J. P. M., DiNola, A., Haak, J. R. Molecular dynamics with coupling to an external bath. *J. Chem. Phys.* 81:3684–3690, 1984.
- [27] Andersen, H. C. Molecular dynamics simulations at constant pressure and/or temperature. *J. Chem. Phys.* 72:2384, 1980.

- [28] Nosé, S. A molecular dynamics method for simulations in the canonical ensemble. *Mol. Phys.* 52:255–268, 1984.
- [29] Hoover, W. G. Canonical dynamics: equilibrium phase-space distributions. *Phys. Rev. A* 31:1695–1697, 1985.
- [30] Bussi, G., Donadio, D., Parrinello, M. Canonical sampling through velocity rescaling. *J. Chem. Phys.* 126:014101, 2007.
- [31] Berendsen, H. J. C. Transport properties computed by linear response through weak coupling to a bath. In: *Computer Simulations in Material Science*. Meyer, M., Pontikis, V. eds. . Kluwer 1991 139–155.
- [32] Basconi, J. E., Shirts, M. R. Effects of temperature control algorithms on transport properties and kinetics in molecular dynamics simulations. *J. Chem. Theory Comput.* 9(7):2887–2899, 2013.
- [33] Cooke, B., Schmidler, S. J. Preserving the Boltzmann ensemble in replica-exchange molecular dynamics. *J. Chem. Phys.* 129:164112, 2008.
- [34] Martyna, G. J., Klein, M. L., Tuckerman, M. E. Nosé-Hoover chains: The canonical ensemble via continuous dynamics. *J. Chem. Phys.* 97:2635–2643, 1992.
- [35] Martyna, G. J., Tuckerman, M. E., Tobias, D. J., Klein, M. L. Explicit reversible integrators for extended systems dynamics. *Mol. Phys.* 87:1117–1157, 1996.
- [36] Holian, B. L., Voter, A. F., Ravelo, R. Thermostatted molecular dynamics: How to avoid the Toda demon hidden in Nosé-Hoover dynamics. *Phys. Rev. E* 52(3):2338–2347, 1995.
- [37] Eastwood, M. P., Stafford, K. A., Lippert, R. A., Jensen, M. O., Maragakis, P., Predescu, C., Dror, R. O., Shaw, D. E. Equipartition and the calculation of temperature in biomolecular simulations. *J. Chem. Theory Comput.* ASAP:DOI: 10.1021/ct9002916, 2010.
- [38] Parrinello, M., Rahman, A. Polymorphic transitions in single crystals: A new molecular dynamics method. *J. Appl. Phys.* 52:7182–7190, 1981.
- [39] Nosé, S., Klein, M. L. Constant pressure molecular dynamics for molecular systems. *Mol. Phys.* 50:1055–1076, 1983.
- [40] Liu, G. Dynamical equations for the period vectors in a periodic system under constant external stress. *Can. J. Phys.* 93:974–978, 2015.
- [41] Tuckerman, M. E., Alejandre, J., López-Rendón, R., Jochim, A. L., Martyna, G. J. A Liouville-operator derived measure-preserving integrator for molecular dynamics simulations in the isothermal-isobaric ensemble. *J. Phys. A.* 59:5629–5651, 2006.
- [42] Yu, T.-Q., Alejandre, J., Lopez-Rendon, R., Martyna, G. J., Tuckerman, M. E. Measure-preserving integrators for molecular dynamics in the isothermal-isobaric ensemble derived from the liouville operator. *Chem. Phys.* 370:294–305, 2010.
- [43] Dick, B. G., Overhauser, A. W. Theory of the dielectric constants of alkali halide crystals. *Phys. Rev.* 112:90–103, 1958.

- [44] Jordan, P. C., van Maaren, P. J., Mavri, J., van der Spoel, D., Berendsen, H. J. C. Towards phase transferable potential functions: Methodology and application to nitrogen. *J. Chem. Phys.* 103:2272–2285, 1995.
- [45] van Maaren, P. J., van der Spoel, D. Molecular dynamics simulations of a water with a novel shell-model potential. *J. Phys. Chem. B.* 105:2618–2626, 2001.
- [46] Ryckaert, J. P., Ciccotti, G., Berendsen, H. J. C. Numerical integration of the cartesian equations of motion of a system with constraints; molecular dynamics of n-alkanes. *J. Comp. Phys.* 23:327–341, 1977.
- [47] Miyamoto, S., Kollman, P. A. SETTLE: An analytical version of the SHAKE and RATTLE algorithms for rigid water models. *J. Comp. Chem.* 13:952–962, 1992.
- [48] Andersen, H. C. RATTLE: A “Velocity” version of the SHAKE algorithm for molecular dynamics calculations. *J. Comp. Phys.* 52:24–34, 1983.
- [49] Hess, B., Bekker, H., Berendsen, H. J. C., Fraaije, J. G. E. M. LINCS: A linear constraint solver for molecular simulations. *J. Comp. Chem.* 18:1463–1472, 1997.
- [50] Hess, B. P-LINCS: A parallel linear constraint solver for molecular simulation. *J. Chem. Theory Comput.* 4:116–122, 2007.
- [51] Goga, N., Rzepiela, A. J., de Vries, A. H., Marrink, S. J., Berendsen, H. J. C. Efficient algorithms for Langevin and DPD dynamics. *J. Chem. Theory Comput.* 8:3637–3649, 2012.
- [52] Byrd, R. H., Lu, P., Nocedal, J. A limited memory algorithm for bound constrained optimization. *SIAM J. Scientific. Statistic. Comput.* 16:1190–1208, 1995.
- [53] Zhu, C., Byrd, R. H., Nocedal, J. L-BFGS-B: Algorithm 778: L-BFGS-B, FORTRAN routines for large scale bound constrained optimization. *ACM Trans. Math. Softw.* 23:550–560, 1997.
- [54] Levitt, M., Sander, C., Stern, P. S. The normal modes of a protein: Native bovine pancreatic trypsin inhibitor. *Int. J. Quant. Chem: Quant. Biol. Symp.* 10:181–199, 1983.
- [55] Gō, N., Noguti, T., Nishikawa, T. Dynamics of a small globular protein in terms of low-frequency vibrational modes. *Proc. Natl. Acad. Sci. USA* 80:3696–3700, 1983.
- [56] Brooks, B., Karplus, M. Harmonic dynamics of proteins: Normal modes and fluctuations in bovine pancreatic trypsin inhibitor. *Proc. Natl. Acad. Sci. USA* 80:6571–6575, 1983.
- [57] Hayward, S., Gō, N. Collective variable description of native protein dynamics. *Annu. Rev. Phys. Chem.* 46:223–250, 1995.
- [58] Bennett, C. H. Efficient Estimation of Free Energy Differences from Monte Carlo Data. *J. Comp. Phys.* 22:245–268, 1976.
- [59] Shirts, M. R., Chodera, J. D. Statistically optimal analysis of multiple equilibrium simulations. *J. Chem. Phys.* 129:124105, 2008.

- [60] Hukushima, K., Nemoto, K. Exchange Monte Carlo Method and Application to Spin Glass Simulations. *J. Phys. Soc. Jpn.* 65:1604–1608, 1996.
- [61] Sugita, Y., Okamoto, Y. Replica-exchange molecular dynamics method for protein folding. *Chem. Phys. Lett.* 314:141–151, 1999.
- [62] Seibert, M., Patriksson, A., Hess, B., van der Spoel, D. Reproducible polypeptide folding and structure prediction using molecular dynamics simulations. *J. Mol. Biol.* 354:173–183, 2005.
- [63] Okabe, T., Kawata, M., Okamoto, Y., Mikami, M. Replica-exchange Monte Carlo method for the isobaric-isothermal ensemble. *Chem. Phys. Lett.* 335:435–439, 2001.
- [64] Chodera, J. D., Shirts, M. R. Replica exchange and expanded ensemble simulations as gibbs sampling: Simple improvements for enhanced mixing. *J. Chem. Phys.* 135:194110, 2011.
- [65] de Groot, B. L., Amadei, A., van Aalten, D. M. F., Berendsen, H. J. C. Towards an exhaustive sampling of the configurational spaces of the two forms of the peptide hormone guanylin. *J. Biomol. Str. Dyn.* 13(5):741–751, 1996.
- [66] de Groot, B. L., Amadei, A., Scheek, R. M., van Nuland, N. A. J., Berendsen, H. J. C. An extended sampling of the configurational space of HPr from *E. coli*. *PROTEINS: Struct. Funct. Gen.* 26:314–322, 1996.
- [67] Lange, O. E., Schafer, L. V., Grubmuller, H. Flooding in GROMACS: Accelerated barrier crossings in molecular dynamics. *J. Comp. Chem.* 27:1693–1702, 2006.
- [68] Lyubartsev, A. P., Martsinovski, A. A., Shevkunov, S. V., Vorontsov-Velyaminov, P. N. New approach to Monte Carlo calculation of the free energy: Method of expanded ensembles. *J. Chem. Phys.* 96:1776–1783, 1992.
- [69] Liem, S. Y., Brown, D., Clarke, J. H. R. Molecular dynamics simulations on distributed memory machines. *Comput. Phys. Commun.* 67(2):261–267, 1991.
- [70] Bowers, K. J., Dror, R. O., Shaw, D. E. The midpoint method for parallelization of particle simulations. *J. Chem. Phys.* 124(18):184109–184109, 2006.
- [71] Qiu, D., Shenkin, P., Hollinger, F., Still, W. The GB/SA Continuum Model for Solvation. A Fast Analytical Method for the Calculation of Approximate Born Radii. *J. Phys. Chem. A.* 101:3005–3014, 1997.
- [72] Hawkins, D., Cramer, C., Truhlar, D. Parametrized Models of Aqueous Free Energies of Solvation Based on Pairwise Descreening of Solute Atomic Charges from a Dielectric Medium. *J. Phys. Chem.* 100:19824–19839, 1996.
- [73] Onufriev, A., Bashford, D., Case, D. Exploring protein native states and large-scale conformational changes with a modified Generalized Born model. *PROTEINS: Struct. Funct. Gen.* 55(2):383–394, 2004.

- [74] Larsson, P., Lindahl, E. A High-Performance Parallel-Generalized Born Implementation Enabled by Tabulated Interaction Rescaling. *J. Comp. Chem.* 31(14):2593–2600, 2010.
- [75] Schaefer, M., Bartels, C., Karplus, M. Solution conformations and thermodynamics of structured peptides: molecular dynamics simulation with an implicit solvation model. *J. Mol. Biol.* 284(3):835–848, 1998.
- [76] Tironi, I. G., Sperb, R., Smith, P. E., van Gunsteren, W. F. A generalized reaction field method for molecular dynamics simulations. *J. Chem. Phys.* 102:5451–5459, 1995.
- [77] van der Spoel, D., van Maaren, P. J. The origin of layer structure artifacts in simulations of liquid water. *J. Chem. Theory Comput.* 2:1–11, 2006.
- [78] Ohmine, I., Tanaka, H., Wolynes, P. G. Large local energy fluctuations in water. II. Cooperative motions and fluctuations. *J. Chem. Phys.* 89:5852–5860, 1988.
- [79] Kitchen, D. B., Hirata, F., Westbrook, J. D., Levy, R., Kofke, D., Yarmush, M. Conserving energy during molecular dynamics simulations of water, proteins, and proteins in water. *J. Comp. Chem.* 11:1169–1180, 1990.
- [80] Guenot, J., Kollman, P. A. Conformational and energetic effects of truncating nonbonded interactions in an aqueous protein dynamics simulation. *J. Comp. Chem.* 14:295–311, 1993.
- [81] Steinbach, P. J., Brooks, B. R. New spherical-cutoff methods for long-range forces in macromolecular simulation. *J. Comp. Chem.* 15:667–683, 1994.
- [82] van Gunsteren, W. F., Billeter, S. R., Eising, A. A., Hünenberger, P. H., Krüger, P., Mark, A. E., Scott, W. R. P., Tironi, I. G. *Biomolecular Simulation: The GROMOS96 manual and user guide.* Zürich, Switzerland: Hochschulverlag AG an der ETH Zürich. 1996.
- [83] van Gunsteren, W. F., Berendsen, H. J. C. *Gromos-87 manual.* Biomos BV Nijenborgh 4, 9747 AG Groningen, The Netherlands 1987.
- [84] Morse, P. M. Diatomic molecules according to the wave mechanics. II. vibrational levels. *Phys. Rev.* 34:57–64, 1929.
- [85] Berendsen, H. J. C., Postma, J. P. M., van Gunsteren, W. F., Hermans, J. Interaction models for water in relation to protein hydration. In: *Intermolecular Forces.* Pullman, B. ed. . D. Reidel Publishing Company Dordrecht 1981 331–342.
- [86] Ferguson, D. M. Parametrization and evaluation of a flexible water model. *J. Comp. Chem.* 16:501–511, 1995.
- [87] Warner Jr., H. R. Kinetic theory and rheology of dilute suspensions of finitely extendible dumbbells. *Ind. Eng. Chem. Fundam.* 11(3):379–387, 1972.
- [88] Bulacu, M., Goga, N., Zhao, W., Rossi, G., Monticelli, L., Periolo, X., Tieleman, D., Marrink, S. Improved angle potentials for coarse-grained molecular dynamics simulations. *J. Chem. Phys.* 123(11).

- [89] Brooks, B. R., Bruccoleri, R. E., Olafson, B. D., States, D. J., Swaminathan, S., Karplus, M. CHARMM: a program for macromolecular energy, minimization, and dynamics calculation. *J. Comp. Chem.* 4:187–217, 1983.
- [90] Lawrence, C. P., Skinner, J. L. Flexible TIP4P model for molecular dynamics simulation of liquid water. *Chem. Phys. Lett.* 372:842–847, 2003.
- [91] Jorgensen, W. L., Maxwell, D. S., Tirado-Rives, J. Development and testing of the opls all-atom force field on conformational energetics and properties of organic liquids. *J. Am. Chem. Soc.* 118:11225–11236, 1996.
- [92] Robertson, M. J., Tirado-Rives, J., Jorgensen, W. L. Improved peptide and protein torsional energetics with the opls-aa force field. *J. Chem. Theory Comput.* 11:3499–3509, 2015.
- [93] Bulacu, M., van der Giessen, E. Effect of bending and torsion rigidity on self-diffusion in polymer melts: A molecular-dynamics study. *JCTC* 9(8):3282–3292, 2013.
- [94] Scott, R. A., Scheraga, H. Conformational analysis of macromolecules. *J. Chem. Phys.* 44:3054–3069, 1966.
- [95] Pauling, L. *The nature of chemical bond.* Ithaca and New York: Cornell University Press. 1960.
- [96] Torda, A. E., Scheek, R. M., van Gunsteren, W. F. Time-dependent distance restraints in molecular dynamics simulations. *Chem. Phys. Lett.* 157:289–294, 1989.
- [97] Hess, B., Scheek, R. M. Orientation restraints in molecular dynamics simulations using time and ensemble averaging. *J. Magn. Reson.* 164:19–27, 2003.
- [98] Lopes, P. E. M., Huang, J., Shim, J., Luo, Y., Li, H., Roux, B., MacKerell, J. Alexander D. Polarizable force field for peptides and proteins based on the classical drude oscillator. *J. Chem. Theory Comput* 9:5430–5449, 2013.
- [99] Yu, H., Whitfield, T. W., Harder, E., Lamoureux, G., Vorobyov, I., Anisimov, V. M., MacKerell, Jr., A. D., Roux, B. Simulating Monovalent and Divalent Ions in Aqueous Solution Using a Drude Polarizable Force Field. *J. Chem. Theory Comput.* 6:774–786, 2010.
- [100] Thole, B. T. Molecular polarizabilities with a modified dipole interaction. *Chem. Phys.* 59:341–345, 1981.
- [101] Lamoureux, G., Roux, B. Modeling induced polarization with classical drude oscillators: Theory and molecular dynamics simulation algorithm. *J. Chem. Phys.* 119:3025–3039, 2003.
- [102] Lamoureux, G., MacKerell, A. D., Roux, B. A simple polarizable model of water based on classical drude oscillators. *J. Chem. Phys.* 119:5185–5197, 2003.
- [103] Noskov, S. Y., Lamoureux, G., Roux, B. Molecular dynamics study of hydration in ethanol-water mixtures using a polarizable force field. *J. Phys. Chem. B.* 109:6705–6713, 2005.
- [104] van Gunsteren, W. F., Mark, A. E. Validation of molecular dynamics simulations. *J. Chem. Phys.* 108:6109–6116, 1998.

- [105] Beutler, T. C., Mark, A. E., van Schaik, R. C., Greber, P. R., van Gunsteren, W. F. Avoiding singularities and numerical instabilities in free energy calculations based on molecular simulations. *Chem. Phys. Lett.* 222:529–539, 1994.
- [106] Pham, T. T., Shirts, M. R. Identifying low variance pathways for free energy calculations of molecular transformations in solution phase. *J. Chem. Phys.* 135:034114, 2011.
- [107] Pham, T. T., Shirts, M. R. Optimal pairwise and non-pairwise alchemical pathways for free energy calculations of molecular transformation in solution phase. *J. Chem. Phys.* 136:124120, 2012.
- [108] Jorgensen, W. L., Tirado-Rives, J. The OPLS potential functions for proteins. energy minimizations for crystals of cyclic peptides and crambin. *J. Am. Chem. Soc.* 110:1657–1666, 1988.
- [109] Berendsen, H. J. C., van Gunsteren, W. F. Molecular dynamics simulations: Techniques and approaches. In: *Molecular Liquids-Dynamics and Interactions.* et al., A. J. B. ed. NATO ASI C 135. Reidel Dordrecht, The Netherlands 1984 475–500.
- [110] Ewald, P. P. Die Berechnung optischer und elektrostatischer Gitterpotentiale. *Ann. Phys.* 64:253–287, 1921.
- [111] Hockney, R. W., Eastwood, J. W. *Computer simulation using particles.* New York: McGraw-Hill. 1981.
- [112] Ballenegger, V., Cerdà, J. J., Holm, C. How to convert SPME to P3M: Influence functions and error estimates. *J. Chem. Theory Comput.* 8(3):936–947, 2012.
- [113] Allen, M. P., Tildesley, D. J. *Computer Simulations of Liquids.* Oxford: Oxford Science Publications. 1987.
- [114] Wennberg, C. L., Murtola, T., Hess, B., Lindahl, E. Lennard-Jones Lattice Summation in Bilayer Simulations Has Critical Effects on Surface Tension and Lipid Properties. *J. Chem. Theory Comput.* 9:3527–3537, 2013.
- [115] Oostenbrink, C., Villa, A., Mark, A. E., Van Gunsteren, W. F. A biomolecular force field based on the free enthalpy of hydration and solvation: The GROMOS force-field parameter sets 53A5 and 53A6. *Journal of Computational Chemistry* 25(13):1656–1676, 2004.
- [116] Cornell, W. D., Cieplak, P., Bayly, C. I., Gould, I. R., Merz, K. R. Jr., Ferguson, D. M., Spellmeyer, D. C., Fox, T., Caldwell, J. W., Kollman, P. A. A Second Generation Force Field for the Simulation of Proteins, Nucleic Acids, and Organic Molecules. *J. Am. Chem. Soc.* 117(19):5179–5197, 1995.
- [117] Kollman, P. A. Advances and Continuing Challenges in Achieving Realistic and Predictive Simulations of the Properties of Organic and Biological Molecules. *Acc. Chem. Res.* 29(10):461–469, 1996.
- [118] Wang, J., Cieplak, P., Kollman, P. A. How Well Does a Restrained Electrostatic Potential (RESP) Model Perform in Calculating Conformational Energies of Organic and Biological Molecules? *J. Comp. Chem.* 21(12):1049–1074, 2000.

- [119] Hornak, V., Abel, R., Okur, A., Strockbine, B., Roitberg, A., Simmerling, C. Comparison of Multiple Amber Force Fields and Development of Improved Protein Backbone Parameters. *PROTEINS: Struct. Funct. Gen.* 65:712–725, 2006.
- [120] Lindorff-Larsen, K., Piana, S., Palmo, K., Maragakis, P., Klepeis, J. L., Dorr, R. O., Shaw, D. E. Improved side-chain torsion potentials for the AMBER ff99SB protein force field. *PROTEINS: Struct. Funct. Gen.* 78:1950–1958, 2010.
- [121] Duan, Y., Wu, C., Chowdhury, S., Lee, M. C., Xiong, G., Zhang, W., Yang, R., Cieplak, P., Luo, R., Lee, T., Caldwell, J., Wang, J., Kollman, P. A Point-Charge Force Field for Molecular Mechanics Simulations of Proteins Based on Condensed-Phase Quantum Mechanical Calculations. *J. Comp. Chem.* 24(16):1999–2012, 2003.
- [122] García, A. E., Sanbonmatsu, K. Y. α -Helical stabilization by side chain shielding of backbone hydrogen bonds. *Proc. Natl. Acad. Sci. USA* 99(5):2782–2787, 2002.
- [123] MacKerell, J. A. D., Feig, M., Brooks III, C. L. Extending the treatment of backbone energetics in protein force fields: limitations of gas-phase quantum mechanics in reproducing protein conformational distributions in molecular dynamics simulations. *J. Comp. Chem.* 25(11):1400–15, 2004.
- [124] MacKerell, A. D., Bashford, D., Bellott, Dunbrack, R. L., Evanseck, J. D., Field, M. J., Fischer, S., Gao, J., Guo, H., Ha, S., Joseph-McCarthy, D., Kuchnir, L., Kuczera, K., Lau, F. T. K., Mattos, C., Michnick, S., Ngo, T., Nguyen, D. T., Prodhom, B., Reiher, W. E., Roux, B., Schlenkrich, M., Smith, J. C., Stote, R., Straub, J., Watanabe, M., Wiorkiewicz-Kuczera, J., Yin, D., Karplus, M. All-atom empirical potential for molecular modeling and dynamics studies of proteins. *J. Phys. Chem. B.* 102(18):3586–3616, 1998.
- [125] Feller, S. E., MacKerell, A. D. An improved empirical potential energy function for molecular simulations of phospholipids. *J. Phys. Chem. B.* 104(31):7510–7515, 2000.
- [126] Foloppe, N., MacKerell, A. D. All-atom empirical force field for nucleic acids: I. Parameter optimization based on small molecule and condensed phase macromolecular target data. *J. Comp. Chem.* 21(2):86–104, 2000.
- [127] MacKerell, A. D., Banavali, N. K. All-atom empirical force field for nucleic acids: II. application to molecular dynamics simulations of DNA and RNA in solution. *J. Comp. Chem.* 21(2):105–120, 2000.
- [128] Bjelkmar, P., Larsson, P., Cuendet, M. A., Hess, B., Lindahl, E. Implementation of the CHARMM force field in GROMACS: Analysis of protein stability effects from correction maps, virtual interaction sites, and water models. *J. Chem. Theory Comput.* 6:459–466, 2010.
- [129] Kohlmeyer, A., Vermaas, J. TopoTools: Release 1.6 with CHARMM export in topogromacs. Apr. 2016.
- [130] Rühle, V., Junghans, C., Lukyanov, A., Kremer, K., Andrienko, D. Versatile Object-Oriented toolkit for Coarse-Graining applications. *J. Chem. Theory Comput.* 5(12):3211–3223, 2009.

- [131] Bereau, T., Wang, Z.-J., Deserno, M. Solvent-free coarse-grained model for unbiased high-resolution protein-lipid interactions. (submitted).
- [132] Wang, Z.-J., Deserno, M. A systematically coarse-grained solvent-free model for quantitative phospholipid bilayer simulations. *J. Phys. Chem. B.* 114(34):11207–11220, 2010.
- [133] Jorgensen, W. L., Chandrasekhar, J., Madura, J. D., Impey, R. W., Klein, M. L. Comparison of simple potential functions for simulating liquid water. *J. Chem. Phys.* 79:926–935, 1983.
- [134] IUPAC-IUB Commission on Biochemical Nomenclature. Abbreviations and Symbols for the Description of the Conformation of Polypeptide Chains. Tentative Rules (1969). *Biochemistry* 9:3471–3478, 1970.
- [135] Mahoney, M. W., Jorgensen, W. L. A five-site model for liquid water and the reproduction of the density anomaly by rigid, nonpolarizable potential functions. *J. Chem. Phys.* 112:8910–8922, 2000.
- [136] Ryckaert, J. P., Bellemans, A. Molecular dynamics of liquid alkanes. *Far. Disc. Chem. Soc.* 66:95–106, 1978.
- [137] de Loof, H., Nilsson, L., Rigler, R. Molecular dynamics simulations of galanin in aqueous and nonaqueous solution. *J. Am. Chem. Soc.* 114:4028–4035, 1992.
- [138] van Buuren, A. R., Berendsen, H. J. C. Molecular Dynamics simulation of the stability of a 22 residue alpha-helix in water and 30% trifluoroethanol. *Biopolymers* 33:1159–1166, 1993.
- [139] Neumann, R. M. Entropic approach to Brownian Movement. *Am. J. Phys.* 48:354–357, 1980.
- [140] Jarzynski, C. Nonequilibrium equality for free energy differences. *Phys. Rev. Lett.* 78(14):2690 – 2693, 1997.
- [141] O. Engin, M. S. A. Villa, Hess, B. Driving forces for adsorption of amphiphilic peptides to air-water interface. *J. Phys. Chem. B.*
- [142] Lindahl, V., Lidmar, J., Hess, B. Accelerated weight histogram method for exploring free energy landscapes. *The Journal of chemical physics* 141(4):044110, 2014.
- [143] Wang, F., Landau, D. Efficient, multiple-range random walk algorithm to calculate the density of states. *Physical review letters* 86(10):2050, 2001.
- [144] Huber, T., Torda, A. E., van Gunsteren, W. F. Local elevation: a method for improving the searching properties of molecular dynamics simulation. *Journal of computer-aided molecular design* 8(6):695–708, 1994.
- [145] Laio, A., Parrinello, M. Escaping free-energy minima. *Proceedings of the National Academy of Sciences* 99(20):12562–12566, 2002.
- [146] Belardinelli, R., Pereyra, V. Fast algorithm to calculate density of states. *Physical Review E* 75(4):046701, 2007.

- [147] Barducci, A., Bussi, G., Parrinello, M. Well-tempered metadynamics: a smoothly converging and tunable free-energy method. *Physical review letters* 100(2):020603, 2008.
- [148] Lindahl, V., Villa, A., Hess, B. Sequence dependency of canonical base pair opening in the dna double helix. *PLoS computational biology* 13(4):e1005463, 2017.
- [149] Sivak, D. A., Crooks, G. E. Thermodynamic metrics and optimal paths. *Physical review letters* 108(19):190602, 2012.
- [150] Kutzner, C., Czub, J., Grubmüller, H. Keep it flexible: Driving macromolecular rotary motions in atomistic simulations with GROMACS. *J. Chem. Theory Comput.* 7:1381–1393, 2011.
- [151] Caleman, C., van der Spoel, D. Picosecond Melting of Ice by an Infrared Laser Pulse - A simulation study. *Angew. Chem., Int. Ed. Engl.* 47:1417–1420, 2008.
- [152] Kutzner, C., Grubmüller, H., de Groot, B. L., Zachariae, U. Computational electrophysiology: the molecular dynamics of ion channel permeation and selectivity in atomistic detail. *Biophys. J.* 101:809–817, 2011.
- [153] Feenstra, K. A., Hess, B., Berendsen, H. J. C. Improving efficiency of large time-scale molecular dynamics simulations of hydrogen-rich systems. *J. Comp. Chem.* 20:786–798, 1999.
- [154] Hess, B. Determining the shear viscosity of model liquids from molecular dynamics. *J. Chem. Phys.* 116:209–217, 2002.
- [155] Dewar, M. J. S. Development and status of MINDO/3 and MNDO. *J. Mol. Struct.* 100:41, 1983.
- [156] Guest, M. F., Harrison, R. J., van Lenthe, J. H., van Corler, L. C. H. Computational chemistry on the FPS-X64 scientific computers - Experience on single- and multi-processor systems. *Theor. Chim. Act.* 71:117, 1987.
- [157] Frisch, M. J., Trucks, G. W., Schlegel, H. B., Scuseria, G. E., Robb, M. A., Cheeseman, J. R., Montgomery, J. A. Jr., Vreven, T., Kudin, K. N., Burant, J. C., Millam, J. M., Iyengar, S. S., Tomasi, J., Barone, V., Mennucci, B., Cossi, M., Scalmani, G., Rega, N., Petersson, G. A., Nakatsuji, H., Hada, M., Ehara, M., Toyota, K., Fukuda, R., Hasegawa, J., Ishida, M., Nakajima, T., Honda, Y., Kitao, O., Nakai, H., Klene, M., Li, X., Knox, J. E., Hratchian, H. P., Cross, J. B., Bakken, V., Adamo, C., Jaramillo, J., Gomperts, R., Stratmann, R. E., Yazyev, O., Austin, A. J., Cammi, R., Pomelli, C., Ochterski, J. W., Ayala, P. Y., Morokuma, K., Voth, G. A., Salvador, P., Dannenberg, J. J., Zakrzewski, V. G., Dapprich, S., Daniels, A. D., Strain, M. C., Farkas, O., Malick, D. K., Rabuck, A. D., Raghavachari, K., Foresman, J. B., Ortiz, J. V., Cui, Q., Baboul, A. G., Clifford, S., Cioslowski, J., Stefanov, B. B., Liu, G., Liashenko, A., Piskorz, P., Komaromi, I., Martin, R. L., Fox, D. J., Keith, T., Al-Laham, M. A., Peng, C. Y., Nanayakkara, A., Challacombe, M., Gill, P. M. W., Johnson, B., Chen, W., Wong, M. W., Gonzalez, C., Pople, J. A. *Gaussian 03, Revision C.02.* Gaussian, Inc., Wallingford, CT, 2004.

- [158] Car, R., Parrinello, M. Unified approach for molecular dynamics and density-functional theory. *Phys. Rev. Lett.* 55:2471–2474, 1985.
- [159] Field, M., Bash, P. A., Karplus, M. A combined quantum mechanical and molecular mechanical potential for molecular dynamics simulation. *J. Comp. Chem.* 11:700, 1990.
- [160] Maseras, F., Morokuma, K. IMOMM: A New Ab Initio + Molecular Mechanics Geometry Optimization Scheme of Equilibrium Structures and Transition States. *J. Comp. Chem.* 16:1170–1179, 1995.
- [161] Svensson, M., Humbel, S., Froes, R. D. J., Matsubara, T., Sieber, S., Morokuma, K. ONIOM a multilayered integrated MO + MM method for geometry optimizations and single point energy predictions. a test for Diels-Alder reactions and Pt(P(t-Bu)₃)₂ + H₂ oxidative addition. *J. Phys. Chem.* 100:19357, 1996.
- [162] Yesylevskyy, S. ProtSqueeze: simple and effective automated tool for setting up membrane protein simulations. *J. Chem. Inf. Model.* 47:1986–1994, 2007.
- [163] Wolf, M., Hoefling, M., Aponte-Santamaría, C., Grubmüller, H., Groenhof, G. g_membed: Efficient insertion of a membrane protein into an equilibrated lipid bilayer with minimal perturbation. *J. Comp. Chem.* 31:2169–2174, 2010.
- [164] van der Spoel, D., Berendsen, H. J. C. Molecular dynamics simulations of Leu-enkephalin in water and DMSO. *Biophys. J.* 72:2032–2041, 1997.
- [165] van der Spoel, D., van Maaren, P. J., Berendsen, H. J. C. A systematic study of water models for molecular simulation. *J. Chem. Phys.* 108:10220–10230, 1998.
- [166] Smith, P. E., van Gunsteren, W. F. The Viscosity of SPC and SPC/E Water. *Chem. Phys. Lett.* 215:315–318, 1993.
- [167] Balasubramanian, S., Mundy, C. J., Klein, M. L. Shear viscosity of polar fluids: Molecular dynamics calculations of water. *J. Chem. Phys.* 105:11190–11195, 1996.
- [168] Wuttke, J. Imfit. <http://apps.jcns.fz-juelich.de/doku/sc/Imfit>. 2013.
- [169] Steen-Sæthre, B., Hoffmann, A. C., van der Spoel, D. Order parameters and algorithmic approaches for detection and demarcation of interfaces in hydrate-fluid and ice-fluid systems. *J. Chem. Theor. Comput.* 10:5606–5615, 2014.
- [170] Palmer, B. J. Transverse-current autocorrelation-function calculations of the shear viscosity for molecular liquids. *Phys. Rev. E* 49:359–366, 1994.
- [171] Wensink, E. J. W., Hoffmann, A. C., van Maaren, P. J., van der Spoel, D. Dynamic properties of water/alcohol mixtures studied by computer simulation. *J. Chem. Phys.* 119:7308–7317, 2003.
- [172] Guo, G.-J., Zhang, Y.-G., Refson, K., Zhao, Y.-J. Viscosity and stress autocorrelation function in supercooled water: a molecular dynamics study. *Mol. Phys.* 100:2617–2627, 2002.

- [173] Fanourgakis, G. S., Medina, J. S., Prosmi, R. Determining the bulk viscosity of rigid water models. *J. Phys. Chem. A* 116:2564–2570, 2012.
- [174] van der Spoel, D., Vogel, H. J., Berendsen, H. J. C. Molecular dynamics simulations of N-terminal peptides from a nucleotide binding protein. *PROTEINS: Struct. Funct. Gen.* 24:450–466, 1996.
- [175] Amadei, A., Linssen, A. B. M., Berendsen, H. J. C. Essential dynamics of proteins. *PROTEINS: Struct. Funct. Gen.* 17:412–425, 1993.
- [176] Hess, B. Convergence of sampling in protein simulations. *Phys. Rev. E* 65:031910, 2002.
- [177] Hess, B. Similarities between principal components of protein dynamics and random diffusion. *Phys. Rev. E* 62:8438–8448, 2000.
- [178] Mu, Y., Nguyen, P. H., Stock, G. Energy landscape of a small peptide revealed by dihedral angle principal component analysis. *PROTEINS: Struct. Funct. Gen.* 58:45–52, 2005.
- [179] van der Spoel, D., van Maaren, P. J., Larsson, P., Timneanu, N. Thermodynamics of hydrogen bonding in hydrophilic and hydrophobic media. *J. Phys. Chem. B.* 110:4393–4398, 2006.
- [180] Luzar, A., Chandler, D. Hydrogen-bond kinetics in liquid water. *Nature* 379:55–57, 1996.
- [181] Luzar, A. Resolving the hydrogen bond dynamics conundrum. *J. Chem. Phys.* 113:10663–10675, 2000.
- [182] Kabsch, W., Sander, C. Dictionary of protein secondary structure: Pattern recognition of hydrogen-bonded and geometrical features. *Biopolymers* 22:2577–2637, 1983.
- [183] Bekker, H., Berendsen, H. J. C., Dijkstra, E. J., Achterop, S., v. Drunen, R., v. d. Spoel, D., Sijbers, A., Keegstra, H., Reitsma, B., Renardus, M. K. R. Gromacs Method of Virial Calculation Using a Single Sum. In *Physics Computing 92* (Singapore, 1993). de Groot, R. A., Nadrchal, J., eds. . World Scientific.
- [184] Berendsen, H. J. C., Grigera, J. R., Straatsma, T. P. The missing term in effective pair potentials. *J. Phys. Chem.* 91:6269–6271, 1987.
- [185] van Gunsteren, W. F., Berendsen, H. J. C. Molecular dynamics of simple systems. *Practicum Handleiding voor MD Practicum Nijenborgh 4*, 9747 AG, Groningen, The Netherlands 1994.

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